

Sergio A. Quezada
Karl S. Peggs
Tyler R. Simpson
James P. Allison

Shifting the equilibrium in cancer immunoediting: from tumor tolerance to eradication

Authors' addresses

Sergio A. Quezada¹, Karl S. Peggs², Tyler R. Simpson¹, James P. Allison¹

¹Ludwig Center for Cancer Immunotherapy, Howard Hughes Medical Institute, Department of Immunology, Memorial Sloan-Kettering Cancer Center, New York, NY, USA.

²Department of Haematology, UCL Cancer Institute, University College London, London, UK.

Correspondence to:

James P. Allison

Ludwig Center for Cancer Immunotherapy
Howard Hughes Medical Institute
Department of Immunology
Memorial Sloan-Kettering Cancer Center
1275 York Avenue
New York, NY 10021, USA
Tel.: +1 212 122 6971
Fax: +1 212 717 3212
e-mail: allisonj@mskcc.org

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Summary: The continual interaction of the immune system with a developing tumor is thought to result in the establishment of a dynamic state of equilibrium. This equilibrium depends on the balance between effector and regulatory T-cell compartments. Whereas regulatory T cells can infiltrate and accumulate within tumors, effector T cells fail to efficiently do so. Furthermore, effector T cells that do infiltrate the tumor become tightly controlled by different regulatory cellular subsets and inhibitory molecules. The outcome of this balance is critical to survival, and whereas in some cases the equilibrium can rapidly result in the elimination of the transformed cells by the immune system, in many other cases the tumor manages to escape immune control. In this review, we discuss relevant work focusing on the establishment of the intratumor balance, the dynamic changes in the populations of effector and regulatory T cells within the tumor, and the role of the tumor vasculature and its activation state in the recruitment of different T-cell subsets. Finally, we also discuss work associated to the manipulation of the immune response to tumors and its impact on the infiltration, accumulation, and function of tumor-reactive lymphocytes within the tumor microenvironment.

Keywords: costimulation, cancer, tumor microenvironment, immunotherapy, vaccines

The immune system and cancer

The relative contribution of the immune system to the control of cancer growth and its spread has been debated for many years. The 'cancer immunosurveillance' hypothesis, initially postulated by Burnet and Thomas in the late 1950s, proposed that as tumors grew they could elicit efficient immunity which prevented clinical manifestation and that the immune system had evolved, at least in part, to control malignant cell outgrowth (1). Subsequent attempts to prove this hypothesis showed that mice with an impaired immune system were more susceptible to tumors, but controversy persisted as these findings were mostly limited to chemically or virally induced tumors. In the case of virus-associated tumors, it was argued that the results could be attributed to virus-mediated transformation consequent upon impaired immunity against the virus rather than as a direct effect of the impaired immune response

directed towards the cancer cells *per se*. Later work further fueled the debate, as a series of experiments comparing wild-type and nude mice showed no difference in the incidence of non-virally derived tumors (2, 3). Only in the last two decades has the concept of cancer immunosurveillance been more fully accepted following a series of publications demonstrating that mice genetically deficient in an array of key components of the immune response (RAG^{-/-}, RAG^{-/-}STAT^{-/-}, PFN^{-/-}, IFN γ ^{-/-}, and IFN γ R^{-/-}) had higher susceptibility to spontaneous, transplantable, and chemically induced tumors (4–7). The concept of cancer immunosurveillance has evolved into a larger and more complex ‘cancer immunoediting’ model, initially introduced by Ikeda, Old, and Schreiber (1, 8), and defined by three key events: elimination, equilibrium and escape. In this model, the ‘elimination’ phase corresponds to cancer immunosurveillance, where tumors are detected and destroyed by various components of the immune response. During the ‘equilibrium’ phase, a balance is established between the tumor and the immune system, during which both tumors and immune cells are shaped reciprocally by each other. Finally, the immune system contributes to the selection of tumor variants that will then grow uncontrollably and ‘escape’ immune control (9).

It is during the equilibrium phase that the interplay between several components of the immune system and the tumor will define the final outcome of the immune response. It is now clear that as tumors develop they can be infiltrated by different subsets of effector, helper, and regulatory T cells (Treg) which, together with myeloid derived suppressor cells (MDSCs), can shape the microenvironment into one less permissive for effector T-cell (T_H1) function. Furthermore, transition through the equilibrium phase not only depends on the extrinsic control exerted by Treg cells and MDSCs but also on the intrinsic regulation of T-cell function by co-inhibitory and costimulatory receptor–ligand pairs. Understanding the key factors involved in maintaining the balance during the equilibrium phase and recognizing ways to interfere with them will help us devise new therapeutic strategies capable of tilting this balance towards elimination instead of escape.

Tumor-specific tolerance – general principles

Progression of cancer may depend on multiple changes within the tumor, including changes intrinsic to the tumor cells resulting in loss or attenuation of immunogenicity (as proposed in immunoediting models), and changes that the tumor cells induce in the surrounding microenvironment or more broadly exert on host immunity to induce a state of immunological tolerance. Support for these latter mechanisms comes

from studies such as those of Willmsky and Blankenstein (10) using a mouse model in which the viral SV40 large T cancer-promoting gene was controlled to activate rarely in random tissues. Although immune responses to the SV40 large T protein were initially detected in such mice, they subsequently developed immune tolerance, whereas the tumors remained capable of eliciting vigorous immunity when transferred into identical but tumor-free mice. Although tumor-specific immunity is compromised in tumor-bearing mice, there is often not generalized immune deficiency (11), indicating that tumors can specifically suppress the induction of effective antitumor immunity, subjugating host responses to create isolated nodes of immune privilege within otherwise immunologically intact hosts. Thus, in models of concomitant immunity, mice injected with a tumor are capable of rejecting a subsequent challenge with the same tumor at a distant site, despite continued growth at the site of initial challenge (12–14). Such concomitant immunity is eventually subverted during primary tumor progression by the establishment of CD4⁺ T-cell-mediated immune suppression (15), which has more recently been shown to be mediated largely by CD4⁺ CD25⁺ Foxp3⁺ Treg cells (16).

Changes occurring during the escape phase of tumor growth that contribute to the development of functional tolerance may be broadly considered as those intrinsic to the tumor cells and those involving the local tumor microenvironment. For example, increased expression of T-cell inhibitory molecules, such as programmed cell death ligand 1 (PD-L1), B7-H3, B7x, HLA-G and HLA-E, by the tumor cells or surrounding parenchyma [stromal or antigen-presenting cells (APCs)] can directly inhibit T_H1 cell function, and expression levels by the tumor or its microenvironment correlate inversely with outcomes in many epithelial tumors (17–25). Similarly, soluble suppressive factors such as interleukin-10 (IL-10), transforming growth factor- β (TGF- β), vascular endothelial growth factor (VEGF), or gangliosides may be elaborated by either the tumor cells or parenchyma (26–32). Indoleamine 2,3-dioxygenase (IDO) expression by tumor cells or IDO-competent APCs can also contribute to acquired tolerance, both by direct suppression of T cells and by enhancement of local regulatory T-cell-mediated suppression (33, 34). IDO catalyzes the rate-limiting step in tryptophan degradation, and the combination of local reduction in tryptophan levels and the production of immunomodulatory tryptophan metabolites appears to exert tolerogenic activity. Furthermore, IDO-expressing plasmacytoid dendritic cells (pDCs) resident within tumor-draining lymph nodes appear to directly activate mature Treg cells, which can subsequently

cause upregulation of PD-L1 by other DCs which in turn inhibits Teff proliferation (35). The presence of an array of other cell types capable of actively suppressing immune responses such as CD4⁺ CD25⁺ Foxp3⁺ Treg cells, IL-10-secreting Treg, CD1d-restricted natural killer T (NKT) cells, immature DCs and pDCs, and MDSCs has been demonstrated to be pivotal for the induction and/or maintenance of local immune privilege in a number of animal models (34, 36, 37). Such cells may be preferentially recruited to these sites or expanded or induced therein.

Extrinsic suppressors: T cells

CD4⁺ T cells can in many ways be considered as master regulators of immune responses, contributing both to development of effector and suppressor activities. The dominant inhibitory potential of Treg cell populations in murine models of malignancy is well established (38), and a similar potential role in human malignancies has been suggested (39). The mechanisms driving Treg cell expansion and accumulation in patients with cancer are not fully understood, but both proliferation of pre-existing Treg and conversion from naive precursors are likely to be involved (40–42). Suppressor populations fall broadly into one of two categories: a thymically derived population that appear crucially dependent on the expression of the X-linked forkhead/winged helix transcription factor Foxp3 for their development (so-called ‘naturally occurring’ Treg) (43–49), and a peripherally induced population which arise from naïve CD4⁺ T cells as a result of ‘tolerogenic’ encounters. These ‘inducible’ Treg include IL-10-producing Tr1 cells (50–52), TGF- β -producing Th3 cells, which are mostly associated with oral tolerance (53, 54), and extrathymically or *de novo* generated CD4⁺ CD25⁺ Foxp3⁺ inducible Treg (iTreg) cells (55–60). The acquisition of regulatory phenotype by conventional non-regulatory CD4⁺ T cells appears important for the maintenance of T-cell homeostasis and control of inflammation. Assuming antigen encounter is required for conversion, it is likely that the regulatory pool expands at the expense of potential Teff, since precursors recognizing tumor antigens may be redirected into a suppressor rather than effector phenotype (40, 61, 62). Factors such as suboptimal antigen stimulation in combination with TGF- β appear to be important in driving peripheral conversion, both of which are likely to be relevant within the tumor microenvironment (57, 63).

Extrinsic suppressors: APCs

Suppressive APC populations have been postulated to play a part in the generation of local immune privilege within

tumors. Developing tumors may selectively recruit suppressive APCs or convert stimulatory APCs into suppressors, mirroring the situation with suppressive T-cell populations. The molecular mechanisms underpinning active immune suppression by DC and myeloid populations have not been fully elucidated but include secretion of IL-10 and TGF- β , expression of FAS ligand, PDL1, and elaboration of intracellular IDO (64–68). IDO-competent DCs can induce apoptosis of activated T cells or either T-cell anergy or conversion of effectors into iTregs, as previously outlined (35, 69–71). The local balance of stimulatory versus suppressive APCs is probably critical in determining the eventual outcome of T-cell encounter with antigen in these sites. It has also become clear that the interaction between DCs and Tregs is likely a two-way process (72–74). MDSCs are a heterogeneous group of cellular precursors of macrophages, granulocytes, DCs, and myeloid cells at earlier stages of differentiation (75–77). Specific phenotypic markers that are reflective of suppressor function remain relatively poorly defined (78). MDSC numbers may correlate with clinical outcomes in human cancer (79). Several tumor-derived cytokines have been implicated in the expansion of MDSCs, including VEGF, IL-1 β , and granulocyte-macrophage colony-stimulating factor (GM-CSF) (80–82). The mechanism of MDSC-mediated suppression is complex, involving contributions from either inducible nitric oxide synthase or arginase 1 (65, 83–86), which enable MDSCs to inhibit T-cell responses in various ways, including induction of apoptosis, inhibition of proliferation, or induction of a regulatory phenotype. Type 2 macrophages found at tumor sites have also been implicated in suppression of tumor immunity and seem to share some functional properties with immature myeloid cells (87, 88).

Effector-intrinsic tolerance

The existence of co-inhibitory receptors mediating direct downregulation of lymphocyte activation and/or effector function is a well-recognized feature of the immunoglobulin superfamily. Cytotoxic T-lymphocyte antigen-4 (CTLA-4) is expressed by activated CD4⁺ and CD8⁺ T cells, though its surface expression is tightly regulated with a short half-life. It influences some the earliest events in T-cell activation (89, 90), being rapidly mobilized from intracellular vesicles to the immune synapse after T-cell receptor (TCR) engagement (91). It is constitutively expressed by natural and inducible Foxp3⁺ Tregs, although the majority of CTLA-4 is again found intracellularly. CTLA-4 shares the B7-1 (CD80) and B7-2 (CD86) ligands with CD28, a critical costimulatory molecule. Ligation of CD28 in concert with TCR stimulation enhances

T-cell proliferation by inducing production of IL-2 and antiapoptotic factors, decreasing the number of ligated TCRs that are required for a given biological response (92). CTLA-4 has significantly higher affinities for both B7 ligands than does CD28 and may therefore outcompete CD28 for the ligand (93). Furthermore, CD28 recruitment to the immunological synapse can be disrupted by CTLA-4, which forms extended high affinity lattices of alternating CTLA-4 and B7-1 homodimers (94). CTLA-4 ligation by B7 ligands also induces decreased production of cytokines (particularly IL-2 and its receptor) and cell cycle arrest. Finally, in addition to its role in controlling T_{eff} function, CTLA-4 has an important role in T_{reg}-mediated suppression (95), as further evidenced by the recent demonstration that T_{reg}-specific CTLA-4 deficiency in conditional knockout mice is associated with a profound reduction in their suppressive capacity (96). The function of CTLA-4 as a negative regulator of CD28-dependent T-cell responses is most strikingly demonstrated by the phenotype

of CTLA-4 knockout mice, which succumb to a rapidly lethal polyclonal CD4-dependent lymphoproliferation within 3–4 weeks of birth (97, 98). The role of CTLA-4 in controlling antitumor responses has been demonstrated in many preclinical models of cancer, thus suggesting that interfering with this pathway in cancer patients may also result in improved survival. This was recently demonstrated in a randomized phase III trial in advanced melanoma. Patients receiving a blocking monoclonal antibody against human CTLA-4 (ipilimumab), either alone or in combination with a gp100 peptide vaccine, demonstrated superior overall survival when compared to patients receiving only the vaccine (99). This is the first randomized trial to ever demonstrate that blockade of an immune inhibitory pathway can be used as an effective cancer therapeutic.

PD-1 is more broadly expressed than CD28 or CTLA-4. It can be detected on activated CD4⁺ and CD8⁺ T cells, as well as B cells, monocytes, and at lower levels on NKT cells. It binds

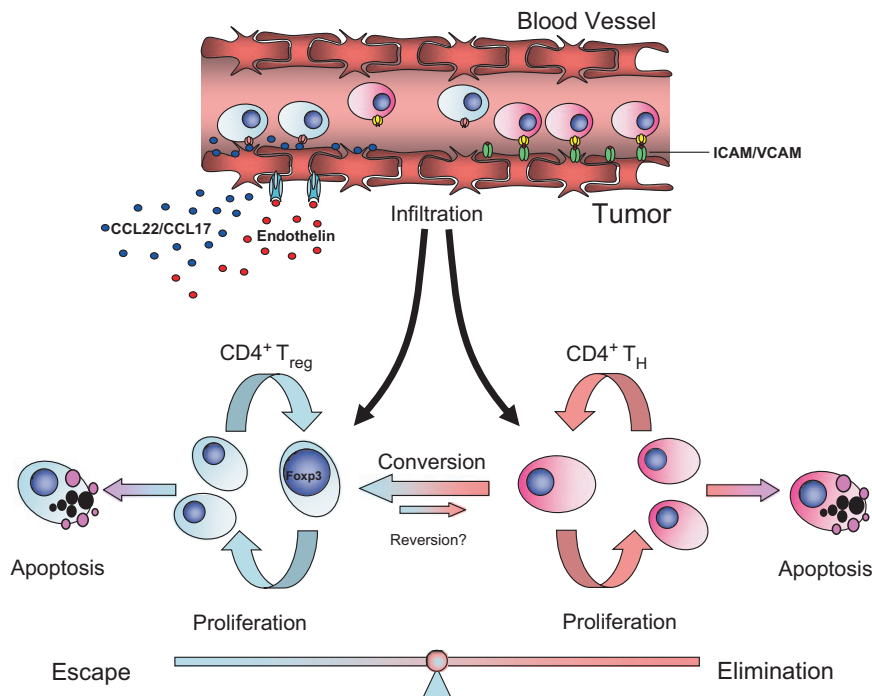


Fig. 1. The intratumor balance of effector and regulatory T cells (T_{reg}). The infiltration of tumors by different subsets of lymphocytes can be initially affected by the activation state of the tumor vasculature as well as by the tumor microenvironment. During tumor progression and under non-inflammatory conditions, a microenvironment rich on CCL22 and or CCL17 will favor recruitment of CD4⁺ Foxp3⁺ T_{reg} cells via engagement of chemokine receptor 4 (CCR4) on their cell surface. Furthermore, endothelin-1 produced by tumors will engage endothelin B receptor on the tumor endothelium and reduce infiltration by activated effector T cells (T_{eff}) through downregulation of intercellular adhesion molecule (ICAM) on the tumor vasculature. Conversely, activation of the tumor vasculature by inflammatory mediators or by immunotherapy can increase the expression of adhesion molecules such as ICAM and vascular cell adhesion molecule (VCAM), thus favoring tumor infiltration by T_{eff}. Once within the tumor, the balance is maintained by expansion and contraction of both regulatory and effector T cells. Low levels of costimulation, absence of Th1-type inflammation, and presence of cytokines such as TGF- β may further contribute to conversion of CD4⁺ Foxp3⁺ naive or helper T cells into CD4⁺ Foxp3⁺ T_{reg}, thus tipping the balance towards tumor tolerance and escape. Although still debated, changes in the stability of Foxp3 may result in T_{reg} reverting into T_{eff}. In addition, an increase on the levels of costimulatory molecules or inflammatory intermediates will also contribute to the accumulation of T_{eff} and reduction of T_{reg} within the tumor, consequently leaning the balance towards tumor elimination.

to two ligands, PD-L1 and PD-L2, which exhibit distinct expression profiles (68). PD-L1 is broadly expressed and can be detected on resting and activated T cells (including CD4⁺ CD25⁺ Foxp3⁺ Tregs), B cells, macrophages, DCs, and mast cells. In addition, its expression on non-hematopoietic cells (including cornea, lung, pancreatic islets, placental syncytiotrophoblast, keratinocytes, and vascular endothelium) may have relevance to the function of this receptor–ligand pair. This broad non-hematopoietic expression pattern suggests that inhibition through the PD-L1/PD-1 axis may not be restricted solely to the interaction of T cells and professional APCs but that it may also be relevant during the effector phase of the immune response in peripheral tissues, perhaps helping to prevent immune-mediated tissue damage directly at the tissue interface. By comparison, PD-L2 has a much more limited expression profile. It is not expressed on naive or activated T cells but is instead restricted to activated macrophages, myeloid DCs, and mast cells, suggesting that it fulfills a role that differs from that of PD-L1. The phenotype of PD-1^{-/-} mice provides perhaps the most direct evidence for an inhibitory role of this receptor (100, 101). These mice can develop an array of autoimmune pathologies characterized by high titers of auto-antibodies.

PD-L1 is expressed by a variety of human and murine tumors, and PD-1 expressed by tumor-infiltrating lymphocytes, suggesting that they may be important in restricting intratumor Teff responses. In humans, myeloid DCs isolated from tumor or lymph nodes from ovarian carcinoma patients express high levels of PD-L1 and are capable of enhancing T-cell activity only following PD-L1 blockade (102). Likewise, pDCs in tumor-draining lymph nodes produce high levels of IDO, which results in Treg cell activation, upregulation of PD-L1 on the DCs, and negative regulation of T-cell responses (35). PD-L1 is expressed on several human carcinomas (mammary, cervical, lung, ovarian, colonic, renal), as well as melanoma, glioblastoma and some hematopoietic malignancies (18, 103–107). Its expression has been directly correlated with poor prognosis in bladder, breast, kidney, gastric, and pancreatic cancer (19, 105, 108). Forced expression of PD-L1 on murine tumor lines diminished T-cell activation and tumor killing *in vitro* and markedly enhanced tumor growth *in vivo*, while anti-PD-L1 antibodies blocked these effects (109, 110).

PD-L1 was recently demonstrated to bind B7-1 with an affinity intermediate between those of CTLA-4 and CD28 for B7-1 (111). This interaction is specific and bidirectional, allowing suppression of T-cell proliferation and cytokine production either through B7-1 or PD-L1. T-cell activation signals delivered through the TCR and CD28 will thus be integrated

with cell-intrinsic co-inhibitory signals delivered through CTLA-4 and PD-L1 (via B7-1 on the APC and potentially also via B7-1 and PD-1 on other T cells), and PD-1 and B7-1 (via PD-L1 on the APC and potentially via CTLA-4, PD-1, and PD-L1 on other T cells). Finally, inhibitory signaling through PD-1 and B7-1 (via PD-L1 on non-hematopoietic tissues) may influence the final outcome of antigen encounter in the periphery. It is likely that there is some redundancy within such complex and apparently overlapping systems. The physiological relevance of some of these findings remains uncertain, but members of the PD-1:PD-L1/PD-L2 grouping clearly make attractive therapeutic targets for attempts to enhance antitumor immunity. Recent data highlight the relevance of this pathway to chronic T-cell responses to pathogens (112–115). During chronic viral infection, antigen-specific CD8⁺ T cells are impaired. These ‘exhausted’ T cells demonstrate a selective upregulation of PD-1, and *in vivo* administration of anti-PD-L1 antibodies restores their activity, as indicated by increased proliferation and cytokine production, and by a significant reduction in viral load (112). Similarly, upregulation of PD-1 on HIV-specific CD8⁺ T cells has been associated with T-cell exhaustion and disease progression in humans (113, 116). Together these data suggest that blockade of PD-1 and/or PD-L1 can restore functionality of the T-cell compartment and could be applied not only to reinvigorate responses to chronic infections but also to enhance T-cell activity towards other chronic pathologies such as cancer.

The intratumor balance of Teff and Treg cells

The Teff/Treg ratio

As mentioned earlier, several layers of regulation can restrict or prevent immunity against tumors. Tregs play a pivotal role in the control of autoimmune diseases and infections, and several studies have also demonstrated their role controlling antitumor immunity (39, 61, 117–121). Together with controlling the initiation of the immune response in peripheral lymphoid organs, Tregs also accumulate at tumor sites in mice and in humans (118, 121–125) where they can regulate helper and Teff responses (126, 127). The attention of immunotherapists has therefore been focusing on the events taking place within the tumor microenvironment. Whereas the proportion of Tregs in peripheral lymphoid organs averages 5–10% of the total CD4⁺ T-cell compartment, this proportion is significantly increased at tumor sites, amounting to 20–30% dependent on the type of tumor (121). This is an important observation, since all *in vitro* and *in vivo* data suggest that Tregs suppress in a dose-dependent manner. Nevertheless, tumor

infiltration is not restricted to Tregs, as other T-cell subsets such as CD4⁺ Foxp3⁻ as well as CD8⁺ T cells can also be found within tumors. Prior to the description of Foxp3 as a key marker for Tregs, many studies had demonstrated that the presence of tumor-infiltrating lymphocytes (TILs) correlated with a favorable overall survival (128–130). Later work from Ohtani's group (131, 132) studying both murine and human cancers went further in the analysis of the TILs and concluded that whereas tumors lacking T-cell infiltrates were most likely to progress, it was the presence of a CD8⁺ T-cell infiltrate and proliferation of such cells that best correlated with favorable prognosis. Further critical insights were provided by Sato et al. (133), who incorporated an additional variable to the analysis of TILs. By analyzing the levels of Foxp3⁺ T-cell infiltrates, they demonstrated that broad characterization of CD3⁺ T-cell infiltrates was not sufficient to determine correlation with survival but instead a high ratio of CD8⁺ T cells to Foxp3⁺ Treg cells was clearly associated with favorable prognosis in epithelial ovarian cancers (133). This major breakthrough in the characterization of TILs was not limited to humans. It was also observed in murine models of cancer. Using the transplantable B16 melanoma (121) and TRAMP-C2 prostate cancer cell lines (authors' unpublished data), we observed that untreated tumors were predominantly infiltrated by CD4⁺ T cells, of which the majority were CD4⁺ Foxp3⁺ Tregs. The relative abundance of CD8⁺ T cells was severely reduced within tumors, where CD8⁺ T cells co-existed with Tregs in similar numbers. Together these data underscored the relevance of the intratumor balance between effector and Treg (Fig. 1) and posed the question of whether tipping this balance towards the Teff compartment would prevent tumor escape while favoring elimination.

Modifying the intratumor balance through costimulation

Both we and others have demonstrated that therapeutic interventions that significantly increase the intratumor Teff/Treg ratio are most likely to result in effective tumor rejection. Combination of a GM-CSF-secreting tumor cell-based vaccine (Gvax) with a blocking anti-CTLA-4 antibody induced substantial tumor infiltration by CD8⁺ Teff cells, which increased the intratumor ratio of CD8⁺ Teff/Treg cells and directly correlated with tumor rejection. In contrast, mice treated with either Gvax or anti-CTLA-4 monotherapy showed only a partial increase in the intratumor CD8⁺ Teff/Treg cell ratio and failed to reject tumors (121). In keeping with this observation, CTLA-4-blockade in combination with a FLT3L-secreting tumor-cell-based vaccine also resulted in significant increases

in the CD8⁺ Teff/Treg ratios and potent tumor rejection (134). Similar results have been observed in cancer patients, where CTLA-4 blockade resulted in increased ratios of effector to Treg (135, 136). The capacity to change the intratumor balance is not restricted to CTLA-4-blockade, as blocking inhibitory signals via PD-1/PD-L1 interactions also resulted in increased Teff/Treg ratios and tumor rejection (137). Combinatorial blockade of both CTLA-4 and PD-1 pathways resulted in an additive effect with significantly higher Teff/Treg ratios and potent tumor rejection. This is encouraging, as a recent study in melanoma patients demonstrated that the majority to tumor infiltrating CD8⁺ T cells expressed high levels of PD-1, thus suggesting this as a relevant pathway in the regulation of intratumor responses in cancer patients (138).

Enhancing stimulation of T-cell function via the tumor necrosis factor receptor family also modifies the intratumor balance of T cells, as treatment of established tumors with agonistic anti-GITR (139) or with a combination of cyclophosphamide and an OX40 agonistic antibody (140) resulted in significant CD8⁺ T-cell infiltration with a concomitant reduction in Foxp3⁺ Treg cells within the tumors.

Although numerous studies have demonstrated a correlation between the changes in the intratumor balance of Teff/Treg cells and tumor rejection, we still lack a clear understanding of the mechanisms driving such changes. In most cases, overt accumulation of Teff cells and reduction of Treg cells at the tumor site are most likely explanations for the increase in the Teff/Treg ratio, but the cellular and molecular mechanisms underpinning these changes remain less clear.

Dynamic changes in the frequency of tumor infiltrating Treg

Treg accumulation

The description of an intratumor balance favored by the natural accumulation of CD4⁺ Foxp3⁺ Treg cells and the impact of modification of such tumor balance through immunotherapy gives rise to two major questions: (i) what drives the accumulation of Tregs within tumors during tumor development and (ii) what are the mechanisms driving the increase in the intratumor Teff/Treg cell ratio following immunotherapy.

The most likely explanations for Treg accumulation during tumorigenesis include an increase in Treg infiltration, enhanced proliferation, reduced apoptosis, or *de novo* induction (or conversion) of CD4⁺ Foxp3⁻ cells into CD4⁺ Foxp3⁺ Treg cells. Clearly these mechanisms are not mutually exclusive. Tumor infiltration driven by the expression of the chemokine receptor 4 (CCR4) on Tregs is considered a major contributor

in some settings. Seminal studies by Curiel et al. (39) demonstrated that in human ovarian carcinoma, a high frequency of tumor-infiltrating Treg cells correlated with poor survival. They were able to demonstrate *in vitro* and *in vivo* that Treg infiltration (but not Teff infiltration) depended on CCL22/CCR4 interactions with CCL22 being produced both by tumor cells and by tumor-infiltrating macrophages (39). Subsequent studies in melanoma (141), breast cancer (142), Hodgkin's lymphoma (143, 144), and most recently in human glioblastoma where the presence and frequency of Tregs also correlated with the WHO tumor grade (145), further support a role for TARC/CCL17 and MDC/CCL22 (specific ligands for CCR4) in tumor infiltration by CD4⁺ Foxp3⁺ CCR4⁺ Treg cells. Several strategies including monoclonal antibodies or receptor antagonists are being developed to target CCR4⁺ Treg cells and prevent tumor infiltration, although their efficacy at preventing or, more importantly, reverting Treg accumulation remains to be fully demonstrated (144, 146, 147).

Less is known about changes of Treg proliferation within tumors. In a model of murine autoimmune diabetes, low levels of IL-2 are required for maintenance of intra-islet Treg homeostasis and survival (148). An equivalent scenario could occur in tumors where low levels of IL-2 would help sustain Treg proliferation and homeostasis. Although this hypothesis has not been formally tested, we have previously demonstrated that untreated B16 melanoma is infiltrated by CD4⁺ Foxp3⁺ Treg cells as well as by CD4⁺ Teff cells (120, 121). Teff cells could be providing the low levels of IL-2 required for Treg survival and proliferation within the tumor microenvironment. In keeping with this, analysis of untreated tumors demonstrates high levels of KI-67 expression by tumor infiltrating Treg cells (120). Finally, IDO produced by either tumor cells or parenchyma also favors the activation and expansion of Treg cells (35, 71, 149).

Numerous studies now support the idea that *de novo* induction (conversion) of CD4⁺ Foxp3⁺ Treg cells from CD4⁺ Foxp3⁻ precursors contributes significantly to Treg accumulation within tumors. However, distinction between conversion and expansion can be technically difficult due to a requirement for highly purified populations of CD4⁺ Foxp3⁻ precursors in many conversion models. Initial studies were based on CD4⁺ CD25⁻ purification strategies where there was still a chance for contaminating Foxp3⁺ cells within the CD25⁻ population. The use of Foxp3 green fluorescence protein (GFP) knockin mice as more reliable sources of Foxp3⁻ precursors has only been possible more recently. Studies by the Levitsky group (40) were among the first to demonstrate conversion of CD4⁺ CD25⁻ GITR⁻ HA-reactive CD4⁺ T cells into Foxp3⁺ Treg cells in response to HA-expressing B-cell lymphomas

(40). Nevertheless, since conversion was induced in response to tumors expressing MHC class II which could be directly presenting antigen to CD4⁺ T cells, it remains unclear whether this observation is fully translatable to non-hematopoietic class II-negative tumors. Recent studies using Foxp3GFP cells demonstrated that lamina propria DCs could promote *de novo* generation of Foxp3⁺ Tregs upon oral exposure to antigen in a retinoic acid-dependent manner (150). In this model, *in vivo* conversion depended on TGF- β as well as on a lymphopenic environment. Two recent studies suggest conversion as a main mechanism for Treg generation in response to tumors. The first, using a murine pancreatic tumor cell line (Pan02) showed an increase in *de novo* induction of Treg *in vivo* in a TGF- β dependent manner (151). The second demonstrated that conversion of OVA-reactive CD4⁺ T cells in response to OVA-expressing B16 melanoma was dependent on PD-1/PD-L1 interaction (152). Interestingly in both studies, conversion seemed to depend on host lymphodepletion. TGF- β emerges from these studies as a common requirement for conversion. It is provided by many tumor types, supporting the idea that the intratumor microenvironment can drive conversion. The apparent requirement for lymphopenia in driving conversion in many of these models remains intriguing. Is lymphopenia really required, or are there conditions within the tumor microenvironment that resemble those of a lymphopenic environment? Interestingly, under many circumstances, cancer patients can be partially lymphodepleted either by the effect of chemo-therapeutic or radio-therapeutic interventions or in response to the tumor itself. Perhaps it is in these conditions where *de novo* induction of Treg cells becomes more relevant in the expansion of the regulatory compartment.

Treg reduction

How can this tolerogenic state or balance in the tumors be broken? Increased tumor infiltration by activated Teff cells clearly contributes (121, 139, 140). But besides Teff infiltration, a decrease in the absolute number of Tregs within the tumor can also account for at least part of the increase in the Teff/Treg cell ratio (140, 153, 154). It is not clear what mediates this reduction in Treg numbers. Reduced conversion, increased Treg cell death, impaired infiltration, or even a reduction in the stability of Foxp3 in the regulatory T-cell compartment are all considered potential mechanisms.

Although some studies suggest that manipulation of costimulatory pathways can lead to a reduction in conversion (155–157), it is not clear in the tumor setting if interfering with

those pathways changes the T_{eff}/T_{reg} cell ratio by reducing conversion or by increasing T_{eff} infiltration. An alternative to reduced conversion is the loss of Foxp3 expression, perhaps representing reversion. Treatment of tumor-bearing Foxp3GFP mice with GITR agonistic antibodies resulted in loss of intratumoral Tregs, apparently due to loss of Foxp3. This was verified in histological analysis of tumors, where cells expressing GFP but not Foxp3 were detected, thus supporting the idea of Foxp3 instability (154). In keeping with this finding, a recent study demonstrated that a substantial proportion of CD4⁺ Foxp3⁺ cells have unstable expression of Foxp3. Remarkably, loss of Foxp3 expression renders these cells autoreactive as demonstrated by their capacity to mediate autoimmune diabetes (158). The issue remains controversial, however, as a newer study demonstrated great stability of Foxp3 *in vivo* during steady state and after different inflammatory stimuli including autoimmune diabetes (159). Here, the authors point out that one potential explanation for the discrepant outcomes of these studies may reside in different regulation of Foxp3 expression in the bacterial artificial chromosome transgenic mouse (158) versus the endogenous regulation found in the knock in mouse model (159). Finally there is evidence that Foxp3 stability may be dependent on the cell in which it is expressed. A recent study demonstrated that whereas the majority of CD4⁺ Foxp3⁺ cells are stable (most of them CD25⁺), a much smaller subset within the CD25⁻ population can actually lose and re-acquire Foxp3 expression depending on the environmental cues (160).

Finally, increased Treg death has also been postulated as a potential mechanism accounting for reduced Treg numbers. Although finding apoptotic Tregs has been challenging, a recent paper elegantly demonstrated that a combination of OX40 agonistic antibodies and cyclophosphamide resulted in Treg apoptosis in a murine model of melanoma (140). FAS/FASL interactions have been implicated in increasing Treg apoptosis. In animal models of colitis, inflammatory stimuli were capable of inducing a local FAS-dependent depletion of Tregs without significantly affecting CD4⁺ T_{eff} cells (161). In a recent publication using a murine model of breast cancer, immunizations with both effector and helper epitopes resulted in significant antitumor responses characterized by a striking change in the intratumor balance of T_{eff} and Treg due to a reduction in tumor-infiltrating Tregs. This reduction was due to apoptosis induced by CD4⁺ FASL⁺ T-helper cells induced by the vaccination strategy, and further corroborated after intratumor administration of anti-FASL blocking antibodies prevented Treg apoptosis enabling tumor progression (162). The FAS/FASL hypothesis offers some additional insights into

the establishment and resolution of the equilibrium phase. As a tumor develops and T_{eff} cells recognize antigen in absence of costimulation or inflammation, they will succumb to activation-induced cell death, whereas in this same microenvironment, Tregs will tend to infiltrate, expand, and convert, thus tilting the balance towards tolerance and escape. Conversely during immunotherapy, T_{eff} cells will infiltrate and contribute to the elimination of Tregs through FAS-mediated apoptosis. Importantly, several publications suggest that T_{eff} cells primed *in vivo* become resistant to FAS-mediated killing, thus allowing them to survive within the inflammatory milieu generated in the rejecting tumor (163, 164).

Dissociation of systemic and local responses

Tumor vasculature and microenvironment as barriers to T-cell infiltration

Despite attempts to elicit potent antitumor reactivity through targeting cell-intrinsic and cell-extrinsic regulatory circuits, the responses we are able to generate and quantify in the periphery (i.e. blood and lymphoid structures) have not been mirrored by such promising outcomes in the clinic. In some cases, this could still reflect less than optimal T-cell activation and a lack of durable immunity, but as our understanding of tumor biology grows, we have realized that barriers other than immune-regulatory checkpoints exist. One such barrier is the restriction of efficient redistribution and accumulation of T_{eff} within tumor lesions. Studies in both mice and humans have demonstrated that tumors can continue to grow regardless of detectable levels of tumor-reactive CD4⁺ and CD8⁺ T cells in peripheral blood (165–169). One potential explanation is inability of the tumor or tumor antigen to prime a proper T-cell response, either due to lack of costimulatory signals or due to antigen sequestration. This is supported by work in the RIP-Tag5 pancreatic cancer model, the Lewis lung carcinoma model and the B16/BL6 melanoma model showing that presence of tumor did not result in activation of TCR transgenic tumor-reactive CD8⁺ T cells (168–170). Furthermore, another study by Mark Davis' group (167) identified a group of melanoma patients in whom tumor-specific CD8⁺ T cells identified by MART1 tetramers displayed a naive phenotype, suggesting lack of T-cell activation even in presence of active disease. Nevertheless, there are studies where good levels of T-cell activation can be detected in absence of tumor regression (171, 172). For instance, in the RIP1-Tag5 model, tumor reactive T cells upregulate the activation marker CD44, dilute CFSE, and acquire cytotoxic activity, suggesting that tumors are capable of eliciting T-cell

activation even in the absence of immunization (172). Despite the presence of activated cytotoxic T cells, tumors continued to grow, suggesting that acquisition of potent effector function by the T-cell compartment is insufficient to drive tumor rejection. On the clinical front, immunizations with the MHC I-restricted peptide from the melanoma differentiation antigen gp100 elicit robust CD8⁺ T-cell responses against gp100 in peripheral blood samples (173). Strikingly, however, none of the tumor-reactive lymphocytes isolated from the tumors recognized the gp100 peptide used in the vaccine. This data underscores the schism that can exist between responses measured in the blood and those taking place in the tumors.

As novel therapeutics become available, immunotherapists are attempting to better model treatment of disease by treating fully established vascularized tumors. During our attempts to further increase the potency of CTLA-4-blockade to enable the rejection of larger more established tumors, we combined Gvax/anti-CTLA-4 with Treg depletion. Depletion of Tregs is being investigated in pre-clinical models and in clinical trials as a cancer immunotherapy (174, 175). Despite enhancing T-cell activity against a tumor antigen (175), Treg depletion does not increase survival in melanoma patients (174). In agreement, we found that Treg depletion following tumor establishment significantly enhanced peripheral antitumor activity, although increased activity did not protect against tumor outgrowth owing to a lack of intercellular adhesion molecule (ICAM) and vascular cell adhesion molecule (VCAM) expression by the tumor vasculature and poor infiltration of Teff into the tumor (120). Similar observations were made by the Ganss group (176) in the RIP1-Tag5 pancreatic cancer model where they found that treatment with the Toll-like receptor ligand CpG and vaccination against the tumor antigen Tag prevents outgrowth of tumors in 5-week-old mice, whereas therapy fails in 23-week old-mice bearing established tumors despite similar levels of systemic cytotoxicity towards Tag targets. The failure in tumor protection correlated with reduced infiltration of vaccine-generated T cells into the tumor.

Approaches to overcome barriers in T-cell infiltration into tumors

Since irradiation has been demonstrated to enhance infiltration of T cells into tumors and T-cell effector function (177, 178), we adoptively transferred polyclonal CD4⁺ and CD8⁺ T cells harvested from mice previously treated with Gvax, anti-CTLA-4, and depleted of Tregs into irradiated mice bearing large tumors (120). This treatment protected mice against tumor outgrowth and was associated with expression of ICAM

and VCAM on the tumor vasculature and infiltration of Teff into the tumor. Once again, these findings were in agreement with those from the RIP1-Tag5 model, where irradiation of tumor-bearing mice prior to transfer of tumor-reactive CD4⁺ T cells resulted in increased infiltration of T cells into the tumor, pro-inflammatory chemokine expression, and ICAM and VCAM expression on the tumor vasculature, resulting in slower tumor progression (179). Although our observations regarding ICAM and VCAM and tumor rejection currently represent correlation rather than causation, it is likely they play a key role in tumor rejection by mediating diapedesis of Teff into the tumor parenchyma (180). The apparent dependence of tumor rejection on ICAM and VCAM expression observed in our model was in keeping with a previous studies demonstrating for the first time an inverse correlation between the expression of the endothelin B receptor (ET_BR) on tumor vasculature and survival of ovarian carcinoma patients (181, 182). Furthermore, they demonstrated that ligation of ET_BR by its ligand ET-1 downregulates ICAM expression and that neutralization of ET_BR using a small molecule inhibitor restores ICAM expression and adhesion of T cells to vessels. When tested in mouse models of cancer, the combination of immunotherapy and blockade of ET_BR synergizes to greatly increase infiltration of T cells into the tumor and reduce tumor outgrowth. Together our data suggest that ICAM expression is an important and perhaps limiting step in tumor elimination, and therapies aiming at increasing its expression on the vasculature may produce better antitumor responses. As a further example, a recent study reported that IL-2 and agonistic anti-CD40 antibodies targeted selectively to the tumor vasculature with a peptide results in enhanced accumulation of T cells in tumors and protection against tumor outgrowth (183). The mechanism of protection involves the anti-CD40 antibody acting directly on CD40⁺ vessels to upregulate ICAM and VCAM, thus making the tumor receptive to T-cell infiltration. A similar approach involving targeting CpG-loaded liposomes to the tumor vasculature with a peptide also elevated ICAM expression by tumor-associated vessels and made tumors receptive to treatment with immunotherapy (184).

Accessibility to tumors is not only regulated by ICAM. Regulator of G protein signaling (RGS5) appears to be an additional key player controlling vasculature sprouting and growth (185). In the RIP1-Tag5 system, genetic deletion of RGS5 normalizes tumor vasculature resulting in improved CD8⁺ T-cell infiltration into tumors after immunotherapy (185, 186). These data suggest that therapeutic manipulation of the RGS5 pathway in combination with immunotherapy may enhance infiltration of vaccine-generated lymphocytes.

Although a great part of many current immunotherapeutic strategies focuses on the generation of more robust T-cell responses, these considerations suggest that combination of such therapies with strategies capable of sensitizing the tumor vasculature and microenvironment will significantly synergize to produce maximal T-cell infiltration and tumor destruction, thus overcoming the observed discordance between local intratumor responses and systemic T-cell activity.

Adoptive cell therapy (ACT) to overcome tolerance to tumors

ACT generally consists of the transfer of large number of activated T_{eff} into lymphopenic tumor-bearing recipients (187, 188). Although ACT may be considered a 'brute force' approach that simply depends on saturating the patient with T_{eff}, study of the mechanisms underpinning the efficacy of ACT have generated significant insights into some of the basic components required for effective rejection of established tumors. A key component of ACT strategies is the state of lymphodepletion induced in the host prior to T-cell transfer. Lymphodepletion eliminates cytokine sinks, myeloid suppressor cells (189), and T_{reg} cells at the same time as providing an environment favorable to homeostatic proliferation (190). Host irradiation, used in many cases to induce lymphodepletion, also contributes by sensitizing the tumor stroma (191) and by inducing the upregulation of adhesion molecules on tumor vasculature, thus rendering the tumor susceptible to T-cell infiltration (177, 179). Furthermore, LPS is released from commensal gut flora following radiation therapy. This allows efficient maturation of DCs carrying tumor antigens which can also be generated as a consequence of irradiation (192). Hence, lymphodepletion acts like a 'reset' button capable of breaking the tolerogenic state originally induced and maintained by the growing tumor. Based on these mechanistic insights, we can postulate that approaches inducing short-lasting lymphodepletion (i.e. radio- or chemo-therapy) will efficiently synergize with active immunization strategies aiming at enhancing T-cell function. As a successful example, combination of an agonistic anti-OX-40 antibody with cyclophosphamide resulted in effective eradication of established melanoma (140).

In addition to lymphodepletion, transfer of large numbers of T_{eff} in the correct stage of activation is crucial for the efficacy of classical ACT (193). For years, ACT and the field of tumor immunology in general have focused on the function of tumor-reactive CD8⁺ cytotoxic T cells (CTLs) (194) reflecting the fact that CD8⁺ T cells are considered the ultimate effectors of the immune system, capable of directly engaging and killing their targets. Although it is well known that CD4⁺ T cells contribute to CD8⁺ T-cell function (190, 195), more recent studies attribute a potentially more direct role for the CD4 compartment to antitumor immunity (196–198). Thus, *in vitro* expanded and differentiated tumor-reactive Th17 cells are capable of rejecting established melanoma tumors in mice (199). Furthermore, the transfer of high numbers of tumor-reactive CD4⁺ T cells into a patient with melanoma resulted in a complete response (200). In addition, two recent studies demonstrated that transfer of a small number of naive tumor-specific CD4⁺ T cells into lymphopenic mice results in rejection of large vascularized melanoma lesions (153, 201). Surprisingly, the antitumor activity was not based in classical T help but dependent in the acquisition of granzyme B-dependent cytotoxic activity by tumor reactive CD4⁺ T cells (153). The acquisition of cytotoxic activity by transferred tumor-reactive CD4⁺ T cells distinguished our findings from previous work showing that CD4⁺ T cells can help rejection of less well-established tumors through indirect effects of IFN- γ (198) on NK cells (197) and tumor-infiltrating macrophages (196, 202–204). Remarkably, CD4⁺ Trp1⁺ cells developed all the hallmarks of CD4⁺ T-helper cells in addition to cytolytic activity. Although CD4⁺ CTLs targeting viral (205–207) and allo-antigens (208, 209) have been described previously, the demonstration of similar activity in a more physiological model for self/tumor antigen emphasizes the promise of these cells in cancer immunotherapy.

Together these new advances in the understanding of tumor-reactive CD4⁺ T cells demonstrate their capacity to modify T-cell function as well as the tumor microenvironment, thus becoming a powerful tool in the fight against cancer. Perhaps a better understanding and manipulation of the function of this T-cell subset will provide all the necessary components for the adequate resolution of the equilibrium phase established during cancer immune-editing.

References

1. Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoediting: from immunosurveillance to tumor escape. *Nat Immunol* 2002;3:991–998.
2. Rygaard J, Povlsen CO. Is immunological surveillance not a cell-mediated immune function? *Transplantation* 1974;17:135–136.
3. Rygaard J, Povlsen CO. The mouse mutant nude does not develop spontaneous tumours. An argument against immunological surveillance. *Acta Pathol Microbiol*

- Scand B *Microbiol Immunol* 1974;**82**:99–106.
4. Shankaran V, et al. IFN γ and lymphocytes prevent primary tumour development and shape tumour immunogenicity. *Nature* 2001;**410**:1107–1111.
 5. Street SE, Trapani JA, MacGregor D, Smyth MJ. Suppression of lymphoma and epithelial malignancies effected by interferon gamma. *J Exp Med* 2002;**196**:129–134.
 6. Street SE, Cretney E, Smyth MJ. Perforin and interferon-gamma activities independently control tumor initiation, growth, and metastasis. *Blood* 2001;**97**:192–197.
 7. Kaplan DH, et al. Demonstration of an interferon gamma-dependent tumor surveillance system in immunocompetent mice. *Proc Natl Acad Sci USA* 1998;**95**:7556–7561.
 8. Ikeda H, Old LJ, Schreiber RD. The roles of IFN gamma in protection against tumor development and cancer immunoeediting. *Cytokine Growth Factor Rev* 2002;**13**:95–109.
 9. Koebel CM, et al. Adaptive immunity maintains occult cancer in an equilibrium state. *Nature* 2007;**450**:903–907.
 10. Willimsky G, Blankenstein T. Sporadic immunogenic tumours avoid destruction by inducing T-cell tolerance. *Nature* 2005;**437**:141–146.
 11. Radoja S, Rao TD, Hillman D, Frey AB. Mice bearing late-stage tumors have normal functional systemic T cell responses *in vitro* and *in vivo*. *J Immunol* 2000;**164**:2619–2628.
 12. Kurt RA, et al. T lymphocytes infiltrating sites of tumor rejection and progression display identical V beta usage but different cytotoxic activities. *J Immunol* 1995;**154**:3969–3974.
 13. Kurt RA, Park JA, Schluter SF, Marchalonis JJ, Akporiaye ET. TCR v(beta) usage and clonality of T cells isolated from progressing and rejected tumor sites before and after *in vitro* culture. *Int Immunol* 2000;**12**:639–646.
 14. Blohm U, Roth E, Brommer K, Dumrese T, Rosenthal FM, Pircher H. Lack of effector cell function and altered tetramer binding of tumor-infiltrating lymphocytes. *J Immunol* 2002;**169**:5522–5530.
 15. North RJ, Bursuker I. Generation and decay of the immune response to a progressive fibrosarcoma. I. Ly-1+2- suppressor T cells down-regulate the generation of Ly-1-2+effector T cells. *J Exp Med* 1984;**159**:1295–1311.
 16. Turk MJ, Guevara-Patino JA, Rizzuto GA, Engelhorn ME, Houghton AN. Concomitant tumor immunity to a poorly immunogenic melanoma is prevented by regulatory T cells. *J Exp Med* 2004;**200**:771–782.
 17. Dong H, Chen L. B7-H1 pathway and its role in the evasion of tumor immunity. *J Mol Med* 2003;**81**:281–287.
 18. Dong H, et al. Tumor-associated B7-H1 promotes T-cell apoptosis: a potential mechanism of immune evasion. *Nat Med* 2002;**8**:793–800.
 19. Inman BA, et al. PD-L1 (B7-H1) expression by urothelial carcinoma of the bladder and BCG-induced granulomata: associations with localized stage progression. *Cancer* 2007;**109**:1499–1505.
 20. Thompson RH, et al. B7-H1 glycoprotein blockade: a novel strategy to enhance immunotherapy in patients with renal cell carcinoma. *Urology* 2005;**66**:10–14.
 21. Roth TJ, et al. B7-H3 ligand expression by prostate cancer: a novel marker of prognosis and potential target for therapy. *Cancer Res* 2007;**67**:7893–7900.
 22. Sun Y, et al. B7-H3 and B7-H4 expression in non-small-cell lung cancer. *Lung Cancer* 2006;**53**:143–151.
 23. Zang X, et al. B7-H3 and B7x are highly expressed in human prostate cancer and associated with disease spread and poor outcome. *Proc Natl Acad Sci USA* 2007;**104**:19458–19463.
 24. Tripathi P, Agrawal S. Non-classical HLA-G antigen and its role in the cancer progression. *Cancer Invest* 2006;**24**:178–186.
 25. Derré L, et al. Expression and release of HLA-E by melanoma cells and melanocytes: potential impact on the response of cytotoxic effector cells. *J Immunol* 2006;**177**:3100–3107.
 26. Kawamura K, Bahar R, Natsume W, Sakiyama S, Tagawa M. Secretion of interleukin-10 from murine colon carcinoma cells suppresses systemic antitumor immunity and impairs protective immunity induced against the tumors. *Cancer Gene Ther* 2002;**9**:109–115.
 27. Zhang Q, et al. Adoptive transfer of tumor-reactive transforming growth factor-beta-insensitive CD8+ T cells: eradication of autologous mouse prostate cancer. *Cancer Res* 2005;**65**:1761–1769.
 28. Chen ML, et al. Regulatory T cells suppress tumor-specific CD8 T cell cytotoxicity through TGF-beta signals *in vivo*. *Proc Natl Acad Sci USA* 2005;**102**:419–424.
 29. Gorelik L, Flavell RA. Immune-mediated eradication of tumors through the blockade of transforming growth factor-beta signaling in T cells. *Nat Med* 2001;**7**:1118–1122.
 30. Fahlen L, et al. T cells that cannot respond to TGF- β escape control by CD4+CD25+ regulatory T cells. *J Exp Med* 2005;**201**:737–746.
 31. Gabrilovich DI, Ishida T, Nadaf S, Ohm JE, Carbone DP. Antibodies to vascular endothelial growth factor enhance the efficacy of cancer immunotherapy by improving endogenous dendritic cell function. *Clin Cancer Res* 1999;**5**:2963–2970.
 32. McKallip R, Li R, Ladisch S. Tumor gangliosides inhibit the tumor-specific immune response. *J Immunol* 1999;**163**:3718–3726.
 33. Munn DH, Mellor AL. Indoleamine 2,3-dioxygenase and tumor-induced tolerance. *J Clin Invest* 2007;**117**:1147–1154.
 34. Mellor AL, Munn DH. Creating immune privilege: active local suppression that benefits friends, but protects foes. *Nat Rev Immunol* 2008;**8**:74–80.
 35. Sharma MD, et al. Plasmacytoid dendritic cells from mouse tumor-draining lymph nodes directly activate mature Tregs via indoleamine 2,3-dioxygenase. *J Clin Invest* 2007;**117**:2570–2582.
 36. Zitvogel L, Tesniere A, Kroemer G. Cancer despite immunosurveillance: immunoselection and immunosubversion. *Nat Rev Immunol* 2006;**6**:715–727.
 37. Nagaraj S, et al. Altered recognition of antigen is a mechanism of CD8+ T cell tolerance in cancer. *Nat Med* 2007;**13**:828–835.
 38. Shimizu J, Yamazaki S, Sakaguchi S. Induction of tumor immunity by removing CD25+CD4+ T cells: a common basis between tumor immunity and autoimmunity. *J Immunol* 1999;**163**:5211–5218.
 39. Curiel TJ, et al. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat Med* 2004;**10**:942–949.
 40. Zhou G, Levitsky HI. Natural regulatory T cells and de novo-induced regulatory T cells contribute independently to tumor-specific tolerance. *J Immunol* 2007;**178**:2155–2162.
 41. Liu VC, et al. Tumor evasion of the immune system by converting CD4+CD25- T cells into CD4+CD25+ T regulatory cells: role of tumor-derived TGF-beta. *J Immunol* 2007;**178**:2883–2892.
 42. Ghiringhelli F, et al. Tumor cells convert immature myeloid dendritic cells into TGF- β -secreting cells inducing CD4+CD25+ regulatory T cell proliferation. *J Exp Med* 2005;**202**:919–929.
 43. Takahashi T, et al. Immunologic self-tolerance maintained by CD25+CD4+ naturally anergic and suppressive T cells: induction of autoimmune disease by breaking their anergic/suppressive state. *Int Immunol* 1998;**10**:1969–1980.
 44. Takahashi T, et al. Immunologic self-tolerance maintained by CD25(+)CD4(+) regulatory T cells constitutively expressing cytotoxic T lymphocyte-associated antigen 4. *J Exp Med* 2000;**192**:303–310.
 45. Shimizu J, Yamazaki S, Takahashi T, Ishida Y, Sakaguchi S. Stimulation of CD25(+)CD4(+) regulatory T cells through GITR breaks immunological self-tolerance. *Nat Immunol* 2002;**3**:135–142.

46. Piconese S, Valzasina B, Colombo MP. OX40 triggering blocks suppression by regulatory T cells and facilitates tumor rejection. *J Exp Med* 2008;**205**:825–839.
47. Fontenot JD, Gavin MA, Rudensky AY. Foxp3 programs the development and function of CD4⁺CD25⁺ regulatory T cells. *Nat Immunol* 2003;**4**:330–336.
48. Gavin MA, et al. Foxp3-dependent programme of regulatory T-cell differentiation. *Nature* 2007;**445**:771–775.
49. Zheng Y, Rudensky AY. Foxp3 in control of the regulatory T cell lineage. *Nat Immunol* 2007;**8**:457–462.
50. Groux H, et al. A CD4⁺ T-cell subset inhibits antigen-specific T-cell responses and prevents colitis. *Nature* 1997;**389**:737–742.
51. Levings MK, Sangregorio R, Galbiati F, Squadrone S, de Waal MR, Roncarolo MG. IFN- α and IL-10 induce the differentiation of human type 1 T regulatory cells. *J Immunol* 2001;**166**:5530–5539.
52. Roncarolo MG, Gregori S, Battaglia M, Bacchetta R, Fleischhauer K, Levings MK. Interleukin-10-secreting type 1 regulatory T cells in rodents and humans. *Immunol Rev* 2006;**212**:28–50.
53. Weiner HL. Induction and mechanism of action of transforming growth factor- β -secreting Th3 regulatory cells. *Immunol Rev* 2001;**182**:207–214.
54. Faria AM, Weiner HL. Oral tolerance and TGF- β -producing cells. *Inflamm Allergy Drug Targets* 2006;**5**:179–190.
55. Walker MR, et al. Induction of FoxP3 and acquisition of T regulatory activity by stimulated human CD4⁺CD25⁺ T cells. *J Clin Invest* 2003;**112**:1437–1443.
56. Apostolou I, von Boehmer H. In vivo instruction of suppressor commitment in naive T cells. *J Exp Med* 2004;**199**:1401–1408.
57. Kretschmer K, Apostolou I, Hawiger D, Khazaie K, Nussenzweig MC, von Boehmer H. Inducing and expanding regulatory T cell populations by foreign antigen. *Nat Immunol* 2005;**6**:1219–1227.
58. Curotto de Lafaille MA, Lino AC, Kutchukhidze N, Lafaille JJ. CD25⁺ T cells generate CD25⁺Foxp3⁺ regulatory T cells by peripheral expansion. *J Immunol* 2004;**173**:7259–7568.
59. Liang S, Alard P, Zhao Y, Parnell S, Clark SL, Kosiewicz MM. Conversion of CD4⁺CD25⁺ cells into CD4⁺CD25⁺ regulatory T cells in vivo requires B7 costimulation, but not the thymus. *J Exp Med* 2005;**201**:127–137.
60. Chen W, et al. Conversion of peripheral CD4⁺CD25⁺ naive T cells to CD4⁺CD25⁺ regulatory T cells by TGF- β induction of transcription factor Foxp3. *J Exp Med* 2003;**198**:1875–1886.
61. Valzasina B, Piconese S, Guiducci C, Colombo MP. Tumor-induced expansion of regulatory T cells by conversion of CD4⁺CD25⁺ lymphocytes is thymus and proliferation independent. *Cancer Res* 2006;**66**:4488–4495.
62. Zhou G, Drake CG, Levitsky HI. Amplification of tumor-specific regulatory T cells following therapeutic cancer vaccines. *Blood* 2006;**107**:628–636.
63. Apostolou I, Verginis P, Kretschmer K, Polansky J, Huhn J, von Boehmer H. Peripherally induced Treg: mode, stability, and role in specific tolerance. *J Clin Immunol* 2008;**28**:619–624.
64. Mellor AL, Munn DH. IDO expression by dendritic cells: tolerance and tryptophan catabolism. *Nat Rev Immunol* 2004;**4**:762–774.
65. Bronte V, Serafini P, Mazzoni A, Segal DM, Zanovello P. L-arginine metabolism in myeloid cells controls T-lymphocyte functions. *Trends Immunol* 2003;**24**:302–306.
66. Ferguson TA, Griffith TS. A vision of cell death: Fas ligand and immune privilege 10 years later. *Immunol Rev* 2006;**213**:228–238.
67. Okazaki T, Honjo T. PD-1 and PD-1 ligands: from discovery to clinical application. *Int Immunol* 2007;**19**:813–824.
68. Keir ME, Butte MJ, Freeman GJ, Sharpe AH. PD-1 and its ligands in tolerance and immunity. *Annu Rev Immunol* 2008;**26**:677–704.
69. Munn DH, et al. Expression of indoleamine 2,3-dioxygenase by plasmacytoid dendritic cells in tumor-draining lymph nodes. *J Clin Invest* 2004;**114**:280–290.
70. Munn DH, et al. GCN2 kinase in T cells mediates proliferative arrest and anergy induction in response to indoleamine 2,3-dioxygenase. *Immunity* 2005;**22**:633–642.
71. Fallarino F, et al. The combined effects of tryptophan starvation and tryptophan catabolites down-regulate T cell receptor zeta-chain and induce a regulatory phenotype in naive T cells. *J Immunol* 2006;**176**:6752–6761.
72. Mahnke K, Johnson TS, Ring S, Enk AH. Tolerogenic dendritic cells and regulatory T cells: a two-way relationship. *J Dermatol Sci* 2007;**46**:159–167.
73. Mahnke K, Ring S, Bedke T, Karakhanova S, Enk AH. Interaction of regulatory T cells with antigen-presenting cells in health and disease. *Chem Immunol Allergy* 2008;**94**:29–39.
74. Mahnke K, et al. Induction of immunosuppressive functions of dendritic cells in vivo by CD4⁺CD25⁺ regulatory T cells: role of B7-H3 expression and antigen presentation. *Eur J Immunol* 2007;**37**:2117–2126.
75. Rabinovich GA, Gabrilovich D, Sotomayor EM. Immunosuppressive strategies that are mediated by tumor cells. *Annu Rev Immunol* 2007;**25**:267–296.
76. Marigo I, Dolcetti L, Serafini P, Zanovello P, Bronte V. Tumor-induced tolerance and immune suppression by myeloid derived suppressor cells. *Immunol Rev* 2008;**222**:162–179.
77. Nagaraj S, Gabrilovich DI. Tumor escape mechanism governed by myeloid-derived suppressor cells. *Cancer Res* 2008;**68**:2561–2563.
78. Youn JI, Nagaraj S, Collazo M, Gabrilovich DI. Subsets of myeloid-derived suppressor cells in tumor-bearing mice. *J Immunol* 2008;**181**:5791–5802.
79. Diaz-Montero CM, Salem ML, Nishimura MI, Garrett-Mayer E, Cole DJ, Montero AJ. Increased circulating myeloid-derived suppressor cells correlate with clinical cancer stage, metastatic tumor burden, and doxorubicin-cyclophosphamide chemotherapy. *Cancer Immunol Immunother* 2009;**58**:49–59.
80. Melani C, Chiodoni C, Forni G, Colombo MP. Myeloid cell expansion elicited by the progression of spontaneous mammary carcinomas in c-erbB-2 transgenic BALB/c mice suppresses immune reactivity. *Blood* 2003;**102**:2138–2145.
81. Serafini P, Carbley R, Noonan KA, Tan G, Bronte V, Borrello I. High-dose granulocyte-macrophage colony-stimulating factor-producing vaccines impair the immune response through the recruitment of myeloid suppressor cells. *Cancer Res* 2004;**64**:6337–6343.
82. Song X, et al. CD11b⁺/Gr-1⁺ immature myeloid cells mediate suppression of T cells in mice bearing tumors of IL-1 β -secreting cells. *J Immunol* 2005;**175**:8200–8208.
83. Serafini P, Borrello I, Bronte V. Myeloid suppressor cells in cancer: recruitment, phenotype, properties, and mechanisms of immune suppression. *Semin Cancer Biol* 2006;**16**:53–65.
84. Mazzoni A, et al. Myeloid suppressor lines inhibit T cell responses by an NO-dependent mechanism. *J Immunol* 2002;**168**:689–695.
85. Kusmartsev S, Nefedova Y, Yoder D, Gabrilovich DI. Antigen-specific inhibition of CD8⁺ T cell response by immature myeloid cells in cancer is mediated by reactive oxygen species. *J Immunol* 2004;**172**:989–999.
86. Bronte V, et al. IL-4-induced arginase 1 suppresses alloreactive T cells in tumor-bearing mice. *J Immunol* 2003;**170**:270–278.
87. Sica A, et al. Macrophage polarization in tumour progression. *Semin Cancer Biol* 2008;**18**:349–355.
88. Allavena P, Sica A, Garlanda C, Mantovani A. The Yin–Yang of tumor-associated macrophages in neoplastic progression and immune surveillance. *Immunol Rev* 2008;**222**:155–161.

89. Brunner MC, Chambers CA, Chan FK, Hanke J, Winoto A, Allison JP. CTLA-4-Mediated inhibition of early events of T cell proliferation. *J Immunol* 1999;**162**:5813–5820.
90. Blair PJ, et al. CTLA-4 ligation delivers a unique signal to resting human CD4 T cells that inhibits interleukin-2 secretion but allows Bcl-X(L) induction. *J Immunol* 1998;**160**:12–15.
91. Egen JG, Allison JP. Cytotoxic T lymphocyte antigen-4 accumulation in the immunological synapse is regulated by TCR signal strength. *Immunity* 2002;**16**:23–35.
92. Viola A, Lanzavecchia A. T cell activation determined by T cell receptor number and tunable thresholds. *Science* 1996;**273**:104–106.
93. Collins AV, et al. The interaction properties of costimulatory molecules revisited. *Immunity* 2002;**17**:201–210.
94. Greene JL, et al. Covalent dimerization of CD28/CTLA-4 and oligomerization of CD80/CD86 regulate T cell costimulatory interactions. *J Biol Chem* 1996;**271**:26762–26771.
95. Peggs KS, Quezada SA, Chambers CA, Korman AJ, Allison JP. Blockade of CTLA-4 on both effector and regulatory T cell compartments contributes to the antitumor activity of anti-CTLA-4 antibodies. *J Exp Med* 2009;**205**:1717–1725.
96. Wing K, et al. CTLA-4 control over Foxp3+ regulatory T cell function. *Science* 2008;**322**:271–275.
97. Waterhouse P, et al. Lymphoproliferative disorders with early lethality in mice deficient in C_tla-4. *Science* 1995;**270**:985–988.
98. Tivol EA, Borriello F, Schweitzer AN, Lynch WP, Bluestone JA, Sharpe AH. Loss of CTLA-4 leads to massive lymphoproliferation and fatal multiorgan tissue destruction, revealing a critical negative regulatory role of CTLA-4. *Immunity* 1995;**3**:541–547.
99. Hodi FS, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 2010;**363**:711–723.
100. Nishimura H, Nose M, Hiai H, Minato N, Honjo T. Development of lupus-like autoimmune diseases by disruption of the PD-1 gene encoding an ITIM motif-carrying immunoreceptor. *Immunity* 1999;**11**:141–151.
101. Nishimura H, et al. Autoimmune dilated cardiomyopathy in PD-1 receptor-deficient mice. *Science* 2001;**291**:319–322.
102. Curiel TJ, et al. Blockade of B7-1 improves myeloid dendritic cell-mediated antitumor immunity. *Nat Med* 2003;**9**:562–567.
103. Blank C, et al. PD-L1/B7H-1 inhibits the effector phase of tumor rejection by T cell receptor (TCR) transgenic CD8+ T cells. *Cancer Res* 2004;**64**:1140–1145.
104. Brown JA, et al. Blockade of programmed death-1 ligands on dendritic cells enhances T cell activation and cytokine production. *J Immunol* 2003;**170**:1257–1266.
105. Thompson RH, et al. Costimulatory B7-H1 in renal cell carcinoma patients: indicator of tumor aggressiveness and potential therapeutic target. *Proc Natl Acad Sci USA* 2004;**101**:17174–17179.
106. Dorfman DM, Brown JA, Shahsafaei A, Freeman GJ. Programmed death-1 (PD-1) is a marker of germinal center-associated T cells and angioimmunoblastic T-cell lymphoma. *Am J Surg Pathol* 2006;**30**:802–810.
107. Liu J, et al. Plasma cells from multiple myeloma patients express B7-H1 (PD-L1) and increase expression after stimulation with IFN- γ and TLR ligands via a MyD88-, TRAF6-, and MEK-dependent pathway. *Blood* 2007;**110**:296–304.
108. Hamanishi J, et al. Programmed cell death 1 ligand 1 and tumor-infiltrating CD8+ T lymphocytes are prognostic factors of human ovarian cancer. *Proc Natl Acad Sci USA* 2007;**104**:3360–3365.
109. Iwai Y, Ishida M, Tanaka Y, Okazaki T, Honjo T, Minato N. Involvement of PD-L1 on tumor cells in the escape from host immune system and tumor immunotherapy by PD-L1 blockade. *Proc Natl Acad Sci USA* 2002;**99**:12293–12297.
110. Strome SE, et al. B7-H1 blockade augments adoptive T-cell immunotherapy for squamous cell carcinoma. *Cancer Res* 2003;**63**:6501–6505.
111. Butte MJ, Keir ME, Phamduy TB, Sharpe AH, Freeman GJ. Programmed death-1 ligand 1 interacts specifically with the B7-1 costimulatory molecule to inhibit T cell responses. *Immunity* 2007;**27**:111–122.
112. Barber DL, et al. Restoring function in exhausted CD8 T cells during chronic viral infection. *Nature* 2006;**439**:682–687.
113. Day CL, et al. PD-1 expression on HIV-specific T cells is associated with T-cell exhaustion and disease progression. *Nature* 2006;**443**:350–354.
114. Radziewicz H, et al. Liver-infiltrating lymphocytes in chronic human hepatitis C virus infection display an exhausted phenotype with high levels of PD-1 and low levels of CD127 expression. *J Virol* 2007;**81**:2545–2553.
115. Wherry EJ, et al. Molecular signature of CD8+ T cell exhaustion during chronic viral infection. *Immunity* 2007;**27**:670–684.
116. Kaufmann DE, et al. Upregulation of CTLA-4 by HIV-specific CD4+ T cells correlates with disease progression and defines a reversible immune dysfunction. *Nat Immunol* 2007;**8**:1246–1254.
117. Mempel TR, et al. Regulatory T cells reversibly suppress cytotoxic T cell function independent of effector differentiation. *Immunity* 2006;**25**:129–141.
118. Gao Q, et al. Intratumoral balance of regulatory and cytotoxic T cells is associated with prognosis of hepatocellular carcinoma after resection. *J Clin Oncol* 2007;**25**:2586–2593.
119. Alvaro T, et al. Outcome in Hodgkin's lymphoma can be predicted from the presence of accompanying cytotoxic and regulatory T cells. *Clin Cancer Res* 2005;**11**:1467–1473.
120. Quezada SA, Peggs KS, Simpson TR, Shen Y, Littman DR, Allison JP. Limited tumor infiltration by activated T effector cells restricts the therapeutic activity of regulatory T cell depletion against established melanoma. *J Exp Med* 2008;**205**:2125–2138.
121. Quezada SA, Peggs KS, Curran MA, Allison JP. CTLA4 blockade and GM-CSF combination immunotherapy alters the intratumor balance of effector and regulatory T cells. *J Clin Invest* 2006;**116**:1935–1945.
122. Sinicrope FA, Rego RL, Ansell SM, Knutson KL, Foster NR, Sargent DJ. Intraepithelial effector (CD3+)/regulatory (FoxP3+) T-cell ratio predicts a clinical outcome of human colon carcinoma. *Gastroenterology* 2009;**137**:1270–1279.
123. Jandus C, Bioley G, Speiser DE, Romero P. Selective accumulation of differentiated FOXP3(+) CD4(+) T cells in metastatic tumor lesions from melanoma patients compared to peripheral blood. *Cancer Immunol Immunother* 2008;**57**:1795–1805.
124. Ahmadzadeh M, et al. FOXP3 expression accurately defines the population of intratumoral regulatory T cells that selectively accumulate in metastatic melanoma lesions. *Blood* 2008;**112**:4953–4960.
125. Zhang HH, et al. Regulatory T cell depletion enhances tumor specific CD8 T-cell responses, elicited by tumor antigen NY-ESO-1b in hepatocellular carcinoma patients, in vitro. *Int J Oncol* 2010;**36**:841–848.
126. Yu P, et al. Intratumor depletion of CD4+ cells unmasks tumor immunogenicity leading to the rejection of late-stage tumors. *J Exp Med* 2005;**201**:779–791.
127. Albers AE, Ferris RL, Kim GG, Chikamatsu K, DeLeo AB, Whiteside TL. Immune responses to p53 in patients with cancer: enrichment in tetramer+ p53 peptide-specific T cells and regulatory T cells at tumor sites. *Cancer Immunol Immunother* 2005;**54**:1072–1081.
128. Mihm MC Jr, Clemente CG, Cascinelli N. Tumor infiltrating lymphocytes in lymph node melanoma metastases: a histopathologic prognostic indicator and an expression of local immune response. *Lab Invest* 1996;**74**:43–47.
129. Clemente CG, Mihm MC Jr, Bufalino R, Zurrada S, Collini P, Cascinelli N. Prognostic

- value of tumor infiltrating lymphocytes in the vertical growth phase of primary cutaneous melanoma. *Cancer* 1996;**77**:1303–1310.
130. Clark WH Jr, et al. Model predicting survival in stage I melanoma based on tumor progression. *J Natl Cancer Inst* 1989;**81**:1893–1904.
 131. Naito Y, et al. CD8+ T cells infiltrated within cancer cell nests as a prognostic factor in human colorectal cancer. *Cancer Res* 1998;**58**:3491–3494.
 132. Nakano O, et al. Proliferative activity of intratumoral CD8(+) T-lymphocytes as a prognostic factor in human renal cell carcinoma: clinicopathologic demonstration of antitumor immunity. *Cancer Res* 2001;**61**:5132–5136.
 133. Sato E, et al. Intraepithelial CD8+ tumor-infiltrating lymphocytes and a high CD8+/regulatory T cell ratio are associated with favorable prognosis in ovarian cancer. *Proc Natl Acad Sci USA* 2005;**102**:18538–18543.
 134. Curran MA, Allison JP. Tumor vaccines expressing flt3 ligand synergize with ctla-4 blockade to reject preimplanted tumors. *Cancer Res* 2009;**69**:7747–7755.
 135. Chen H, et al. Anti-CTLA-4 therapy results in higher CD4+ICOShi T cell frequency and IFN-gamma levels in both nonmalignant and malignant prostate tissues. *Proc Natl Acad Sci USA* 2009;**106**:2729–2734.
 136. Liakou CI, et al. CTLA-4 blockade increases IFN-gamma-producing CD4+ICOShi cells to shift the ratio of effector to regulatory T cells in cancer patients. *Proc Natl Acad Sci USA* 2008;**105**:14987–14992.
 137. Curran MA, Montalvo W, Yagita H, Allison JP. PD-1 and CTLA-4 combination blockade expands infiltrating T cells and reduces regulatory T and myeloid cells within B16 melanoma tumors. *Proc Natl Acad Sci USA* 2010;**107**:4275–4280.
 138. Ahmadzadeh M, et al. Tumor antigen-specific CD8 T cells infiltrating the tumor express high levels of PD-1 and are functionally impaired. *Blood* 2009;**114**:1537–1544.
 139. Ko K, et al. Treatment of advanced tumors with agonistic anti-GITR mAb and its effects on tumor-infiltrating Foxp3+CD25+CD4+ regulatory T cells. *J Exp Med* 2005;**202**:885–891.
 140. Hirschhorn-Cymerman D, et al. OX40 engagement and chemotherapy combination provides potent antitumor immunity with concomitant regulatory T cell apoptosis. *J Exp Med* 2009;**206**:1103–1116.
 141. Kimpfler S, et al. Skin melanoma development in ret transgenic mice despite the depletion of CD25+Foxp3+ regulatory T cells in lymphoid organs. *J Immunol* 2009;**183**:6330–6337.
 142. Gobert M, et al. Regulatory T cells recruited through CCL22/CCR4 are selectively activated in lymphoid infiltrates surrounding primary breast tumors and lead to an adverse clinical outcome. *Cancer Res* 2009;**69**:2000–2009.
 143. Ishida T, et al. Specific recruitment of CC chemokine receptor 4-positive regulatory T cells in Hodgkin lymphoma fosters immune privilege. *Cancer Res* 2006;**66**:5716–5722.
 144. Ishida T, Ueda R. CCR4 as a novel molecular target for immunotherapy of cancer. *Cancer Sci* 2006;**97**:1139–1146.
 145. Jacobs JF, et al. Prognostic significance and mechanism of Treg infiltration in human brain tumors. *J Neuroimmunol* 2010;**225**:195–199.
 146. Ito A, et al. Defucosylated anti-CCR4 monoclonal antibody exerts potent ADCC against primary ATLL cells mediated by autologous human immune cells in NOD/Shi-scid, IL-2R gamma(null) mice in vivo. *J Immunol* 2009;**183**:4782–4791.
 147. Davies MN, et al. Toward the discovery of vaccine adjuvants: coupling in silico screening and in vitro analysis of antagonist binding to human and mouse CCR4 receptors. *PLoS ONE* 2009;**4**:e8084.
 148. Tang Q, et al. Central role of defective interleukin-2 production in the triggering of islet autoimmune destruction. *Immunity* 2008;**28**:687–697.
 149. Curti A, et al. Modulation of tryptophan catabolism by human leukemic cells results in the conversion of CD25- into CD25+ T regulatory cells. *Blood* 2007;**109**:2871–2877.
 150. Sun CM, et al. Small intestine lamina propria dendritic cells promote de novo generation of Foxp3 T reg cells via retinoic acid. *J Exp Med* 2007;**204**:1775–1785.
 151. Moo-Young TA, et al. Tumor-derived TGF-beta mediates conversion of CD4+Foxp3+ regulatory T cells in a murine model of pancreas cancer. *J Immunother* 2009;**32**:12–21.
 152. Wang L, Pino-Lagos K, de Vries VC, Guleria I, Sayegh MH, Noelle RJ. Programmed death 1 ligand signaling regulates the generation of adaptive Foxp3+CD4+ regulatory T cells. *Proc Natl Acad Sci USA* 2008;**105**:9331–9336.
 153. Quezada SA, et al. Tumor-reactive CD4(+) T cells develop cytotoxic activity and eradicate large established melanoma after transfer into lymphopenic hosts. *J Exp Med* 2010;**207**:637–650.
 154. Cohen AD, et al. Agonist anti-GITR monoclonal antibody induces melanoma tumor immunity in mice by altering regulatory T cell stability and intra-tumor accumulation. *PLoS ONE* 2010;**5**:e10436.
 155. Zheng SG, Wang JH, Stohl W, Kim KS, Gray JD, Horwitz DA. TGF-beta requires CTLA-4 early after T cell activation to induce FoxP3 and generate adaptive CD4+CD25+ regulatory cells. *J Immunol* 2006;**176**:3321–3329.
 156. So T, Croft M. Cutting edge: OX40 inhibits TGF-beta- and antigen-driven conversion of naive CD4 T cells into CD25+Foxp3+ T cells. *J Immunol* 2007;**179**:1427–1430.
 157. Xiao X, Kroemer A, Gao W, Ishii N, Demirci G, Li XC. OX40/OX40L costimulation affects induction of Foxp3+ regulatory T cells in part by expanding memory T cells in vivo. *J Immunol* 2008;**181**:3193–3201.
 158. Zhou X, et al. Instability of the transcription factor Foxp3 leads to the generation of pathogenic memory T cells in vivo. *Nat Immunol* 2009;**10**:1000–1007.
 159. Rubtsov YP, et al. Stability of the regulatory T cell lineage in vivo. *Science* 2010;**329**:1667–1671.
 160. Komatsu N, Mariotti-Ferrandiz ME, Wang Y, Malissen B, Waldmann H, Hori S. Heterogeneity of natural Foxp3+ T cells: a committed regulatory T-cell lineage and an uncommitted minor population retaining plasticity. *Proc Natl Acad Sci USA* 2009;**106**:1903–1908.
 161. Reardon C, Wang A, McKay DM. Transient local depletion of Foxp3+ regulatory T cells during recovery from colitis via Fas/Fas ligand-induced death. *J Immunol* 2008;**180**:8316–8326.
 162. Gritzapis AD, Voutsas IF, Lekka E, Papamichail M, Baxevanis CN. Peptide vaccination breaks tolerance to HER-2/neu by generating vaccine-specific FasL(+) CD4(+) T cells: first evidence for intratumor apoptotic regulatory T cells. *Cancer Res* 2010;**70**:2686–2696.
 163. Ehl S, Hoffmann-Rohrer U, Nagata S, Hengartner H, Zinkernagel R. Different susceptibility of cytotoxic T cells to CD95 (Fas/Apo-1) ligand-mediated cell death after activation in vitro versus in vivo. *J Immunol* 1996;**156**:2357–2360.
 164. Rivoltini L, et al. Human melanoma-reactive CD4+ and CD8+ CTL clones resist Fas ligand-induced apoptosis and use Fas/Fas ligand-independent mechanisms for tumor killing. *J Immunol* 1998;**161**:1220–1230.
 165. Romero P, et al. Ex vivo staining of metastatic lymph nodes by class I major histocompatibility complex tetramers reveals high numbers of antigen-experienced tumor-specific cytolytic T lymphocytes. *J Exp Med* 1998;**188**:1641–1650.
 166. Bioley G, et al. Melan-A/MART-1-specific CD4 T cells in melanoma patients: identification of new epitopes and ex vivo visualization of specific T cells by MHC class II tetramers. *J Immunol* 2006;**177**:6769–6779.
 167. Lee PP, et al. Characterization of circulating T cells specific for tumor-associated antigens in melanoma patients. *Nat Med* 1999;**5**:677–685.

168. Hermans IF, Daish A, Yang J, Ritchie DS, Ronchese F. Antigen expressed on tumor cells fails to elicit an immune response, even in the presence of increased numbers of tumor-specific cytotoxic T lymphocyte precursors. *Cancer Res* 1998;**58**:3909–3917.
169. Prévost-Blondel A, Zimmermann C, Stemmer C, Kulmburg P, Rosenthal FM, Pircher H. Tumor-infiltrating lymphocytes exhibiting high ex vivo cytolytic activity fail to prevent murine melanoma tumor growth in vivo. *J Immunol* 1998;**161**:2187–2194.
170. Lyman MA, Aung S, Biggs JA, Sherman LA. A spontaneously arising pancreatic tumor does not promote the differentiation of naive CD8+ T lymphocytes into effector CTL. *J Immunol* 2004;**172**:6558–6567.
171. Wick M, et al. Antigenic cancer cells grow progressively in immune hosts without evidence for T cell exhaustion or systemic anergy. *J Exp Med* 1997;**186**:229–238.
172. Nguyen LT, et al. Tumor growth enhances cross-presentation leading to limited T cell activation without tolerance. *J Exp Med* 2002;**195**:423–435.
173. Lee KH, et al. Functional dissociation between local and systemic immune response during anti-melanoma peptide vaccination. *J Immunol* 1998;**161**:4183–4194.
174. Powell DJ, et al. Administration of a CD25-directed immunotoxin, LMB-2, to patients with metastatic melanoma induces a selective partial reduction in regulatory T cells in vivo. *J Immunol* 2007;**179**:4919–4928.
175. Morse MA, et al. Depletion of human regulatory T cells specifically enhances antigen-specific immune responses to cancer vaccines. *Blood* 2008;**112**:610–618.
176. Garbi N, Arnold B, Gordon S, Hämmerling GJ, Ganss R. CpG motifs as proinflammatory factors render autochthonous tumors permissive for infiltration and destruction. *J Immunol* 2004;**172**:5861–5869.
177. Lugade AA, Moran JP, Gerber SA, Rose RC, Frelinger JG, Lord EM. Local radiation therapy of B16 melanoma tumors increases the generation of tumor antigen-specific effector cells that traffic to the tumor. *J Immunol* 2005;**174**:7516–7523.
178. Gattinoni L, et al. Removal of homeostatic cytokine sinks by lymphodepletion enhances the efficacy of adoptively transferred tumor-specific CD8+ T cells. *J Exp Med* 2005;**202**:907–912.
179. Ganss R, Ryschich E, Klar E, Arnold B, Hämmerling GJ. Combination of T-cell therapy and trigger of inflammation induces remodeling of the vasculature and tumor eradication. *Cancer Res* 2002;**62**:1462–1470.
180. Muller WA. Mechanisms of transendothelial migration of leukocytes. *Circ Res* 2009;**105**:223–230.
181. Kandalafi LE, Facciabene A, Buckanovich RJ, Coukos G. Endothelin B receptor, a new target in cancer immune therapy. *Clin Cancer Res* 2009;**15**:4521–4528.
182. Buckanovich RJ, et al. Endothelin B receptor mediates the endothelial barrier to T cell homing to tumors and disables immune therapy. *Nat Med* 2008;**14**:28–36.
183. Hamzah J, Nelson D, Moldenhauer G, Arnold B, Hämmerling GJ, Ganss R. Vascular targeting of anti-CD40 antibodies and IL-2 into autochthonous tumors enhances immunotherapy in mice. *J Clin Invest* 2008;**118**:1691–1699.
184. Hamzah J, et al. Targeted liposomal delivery of TLR9 ligands activates spontaneous antitumor immunity in an autochthonous cancer model. *J Immunol* 2009;**183**:1091–1098.
185. Berger M, Bergers G, Arnold B, Hämmerling GJ, Ganss R. Regulator of G-protein signaling-5 induction in pericytes coincides with active vessel remodeling during neovascularization. *Blood* 2005;**105**:1094–1101.
186. Hamzah J, et al. Vascular normalization in Rgs5-deficient tumours promotes immune destruction. *Nature* 2008;**453**:410–414.
187. Dudley ME, et al. Adoptive cell therapy for patients with metastatic melanoma: evaluation of intensive myeloablative chemoradiation preparative regimens. *J Clin Oncol* 2008;**26**:5233–5239.
188. Dudley ME, et al. Cancer regression and autoimmunity in patients after clonal repopulation with antitumor lymphocytes. *Science* 2002;**298**:850–854.
189. Wrzesinski C, et al. Increased intensity lymphodepletion enhances tumor treatment efficacy of adoptively transferred tumor-specific T cells. *J Immunother* 2010;**33**:1–7.
190. Antony PA, et al. CD8+ T cell immunity against a tumor/self-antigen is augmented by CD4+ T helper cells and hindered by naturally occurring T regulatory cells. *J Immunol* 2005;**174**:2591–2601.
191. Zhang B, et al. Induced sensitization of tumor stroma leads to eradication of established cancer by T cells. *J Exp Med* 2007;**204**:49–55.
192. Paulos CM, et al. Microbial translocation augments the function of adoptively transferred self/tumor-specific CD8 T cells via TLR4 signaling. *J Clin Invest* 2007;**117**:2197–2204.
193. Gattinoni L, et al. Acquisition of full effector function in vitro paradoxically impairs the in vivo antitumor efficacy of adoptively transferred CD8+ T cells. *J Clin Invest* 2005;**115**:1616–1626.
194. Yee C, et al. Adoptive T cell therapy using antigen-specific CD8+ T cell clones for the treatment of patients with metastatic melanoma: in vivo persistence, migration, and antitumor effect of transferred T cells. *Proc Natl Acad Sci USA* 2002;**99**:16168–16173.
195. Pardoll DM, Topalian SL. The role of CD4+ T cell responses in antitumor immunity. *Curr Opin Immunol* 1998;**10**:588–594.
196. Corthay A, et al. Primary antitumor immune response mediated by CD4+ T cells. *Immunity* 2005;**22**:371–383.
197. Perez-Diez A, et al. CD4 cells can be more efficient at tumor rejection than CD8 cells. *Blood* 2007;**109**:5346–5354.
198. Mumberg D, et al. CD4(+) T cells eliminate MHC class II-negative cancer cells in vivo by indirect effects of IFN-gamma. *Proc Natl Acad Sci USA* 1999;**96**:8633–8638.
199. Muranski P, et al. Tumor-specific Th17-polarized cells eradicate large established melanoma. *Blood* 2008;**112**:362–373.
200. Hunder NN, et al. Treatment of metastatic melanoma with autologous CD4+ T cells against NY-ESO-1. *N Engl J Med* 2008;**358**:2698–2703.
201. Xie Y, et al. Naive tumor-specific CD4(+) T cells differentiated in vivo eradicate established melanoma. *J Exp Med* 2010;**207**:651–667.
202. Greenberg PD, Kern DE, Cheever MA. Therapy of disseminated murine leukemia with cyclophosphamide and immune Lyt-1+, 2– T cells. Tumor eradication does not require participation of cytotoxic T cells. *J Exp Med* 1985;**161**:1122–1134.
203. Hung K, Hayashi R, Lafond-Walker A, Lowenstein C, Pardoll D, Levitsky H. The central role of CD4(+) T cells in the antitumor immune response. *J Exp Med* 1998;**188**:2357–2368.
204. Corthay A. CD4+ T cells cooperate with macrophages for specific elimination of MHC class II-negative cancer cells. *Adv Exp Med Biol* 2007;**590**:195–208.
205. Heller KN, Gurer C, Munz C. Virus-specific CD4+ T cells: ready for direct attack. *J Exp Med* 2006;**203**:805–808.
206. Paludan C, et al. Epstein-Barr nuclear antigen 1-specific CD4(+) Th1 cells kill Burkitt's lymphoma cells. *J Immunol* 2002;**169**:1593–1603.
207. Hegde NR, Dunn C, Lewinsohn DM, Jarvis MA, Nelson JA, Johnson DC. Endogenous human cytomegalovirus gB is presented efficiently by MHC class II molecules to CD4+ CTL. *J Exp Med* 2005;**202**:1109–1119.
208. Holloway PA, et al. A class II-restricted cytotoxic T-cell clone recognizes a human minor histocompatibility antigen with a restricted tissue distribution. *Br J Haematol* 2005;**128**:73–81.
209. Spaapen RM, et al. Toward targeting B cell cancers with CD4+ CTLs: identification of a CD19-encoded minor histocompatibility antigen using a novel genome-wide analysis. *J Exp Med* 2008;**205**:2863–2872.