



ΔΙΑΤΜΗΜΑΤΙΚΟ ΠΡΟΓΡΑΜΜΑ ΜΕΤΑΠΤΥΧΙΑΚΩΝ ΣΠΟΥΔΩΝ
«ΕΦΑΡΜΟΓΕΣ ΤΗΣ ΒΙΟΛΟΓΙΑΣ ΣΤΗΝ ΙΑΤΡΙΚΗ»
Μάθημα: Μικροβιολογία και Δημόσια Υγεία



Μέθοδοι διάγνωσης της μικροβιακής αντοχής

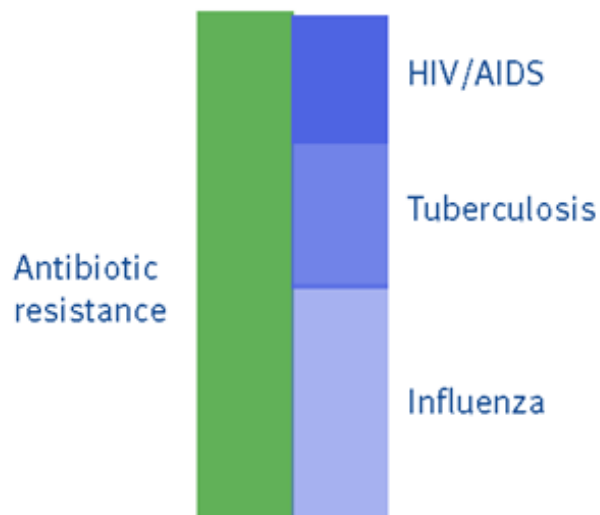
Ολυμπία Ζαρκωτού,

Διευθύντρια Μικροβιολογικού Τμήματος, Γ.Ν. Πειραιά «Τζάνειο»

Antibiotic resistance: a growing threat to human health

Antibiotic resistance is the ability of bacteria to combat the action of one or more antibiotics. Bacteria, not humans or animals, become antibiotic-resistant.

In Europe, the health impact of antibiotic resistant infections is comparable to that of influenza, tuberculosis and HIV/AIDS combined.

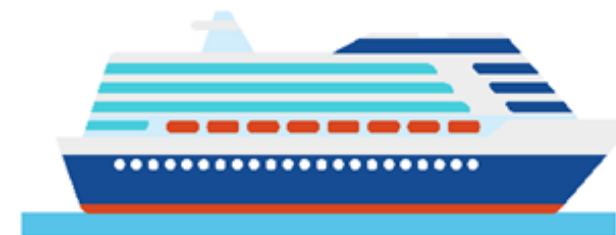


More than
35000 deaths

Each year, more than 35 000 people die from antibiotic-resistant infections in the European Union, Iceland and Norway. This is **equivalent to the number of passengers on 13 cruise ships.**

Antibiotic resistance is a silent pandemic and a growing threat to human health.

13 cruise ships



Over **70 %**
healthcare-associated
infections

Over 70% of the health impact of antibiotic-resistant infections is directly linked to healthcare-associated infections. This could be minimized through adequate infection prevention and control measures, as well as antibiotic stewardship in healthcare settings.

Increasing burden

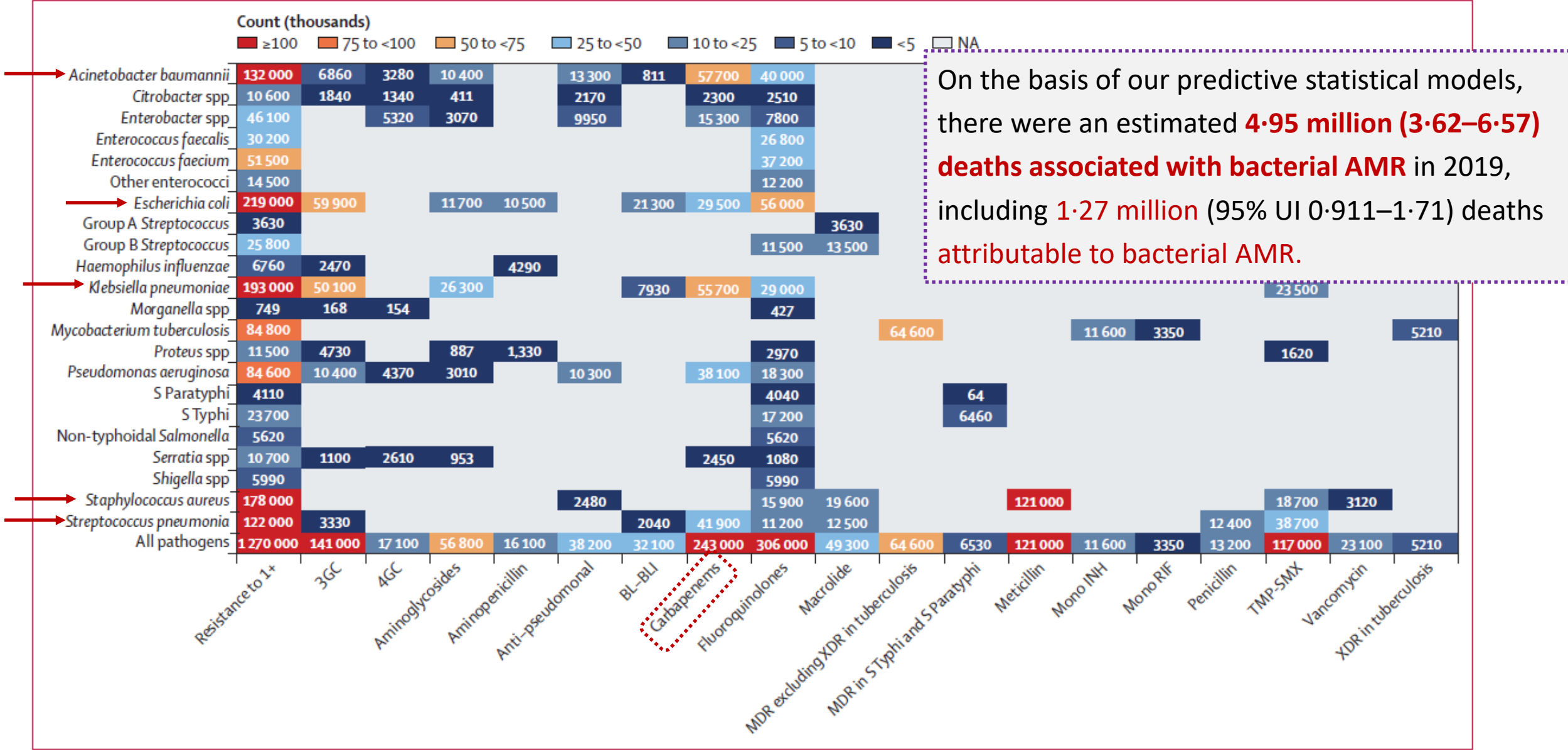
Resistance to antibiotics that are used as last line for treatment of infections, such as the carbapenems, has the highest health impact.

Between 2016 and 2020, the overall number of deaths caused by antibiotic-resistant bacteria under study has increased.

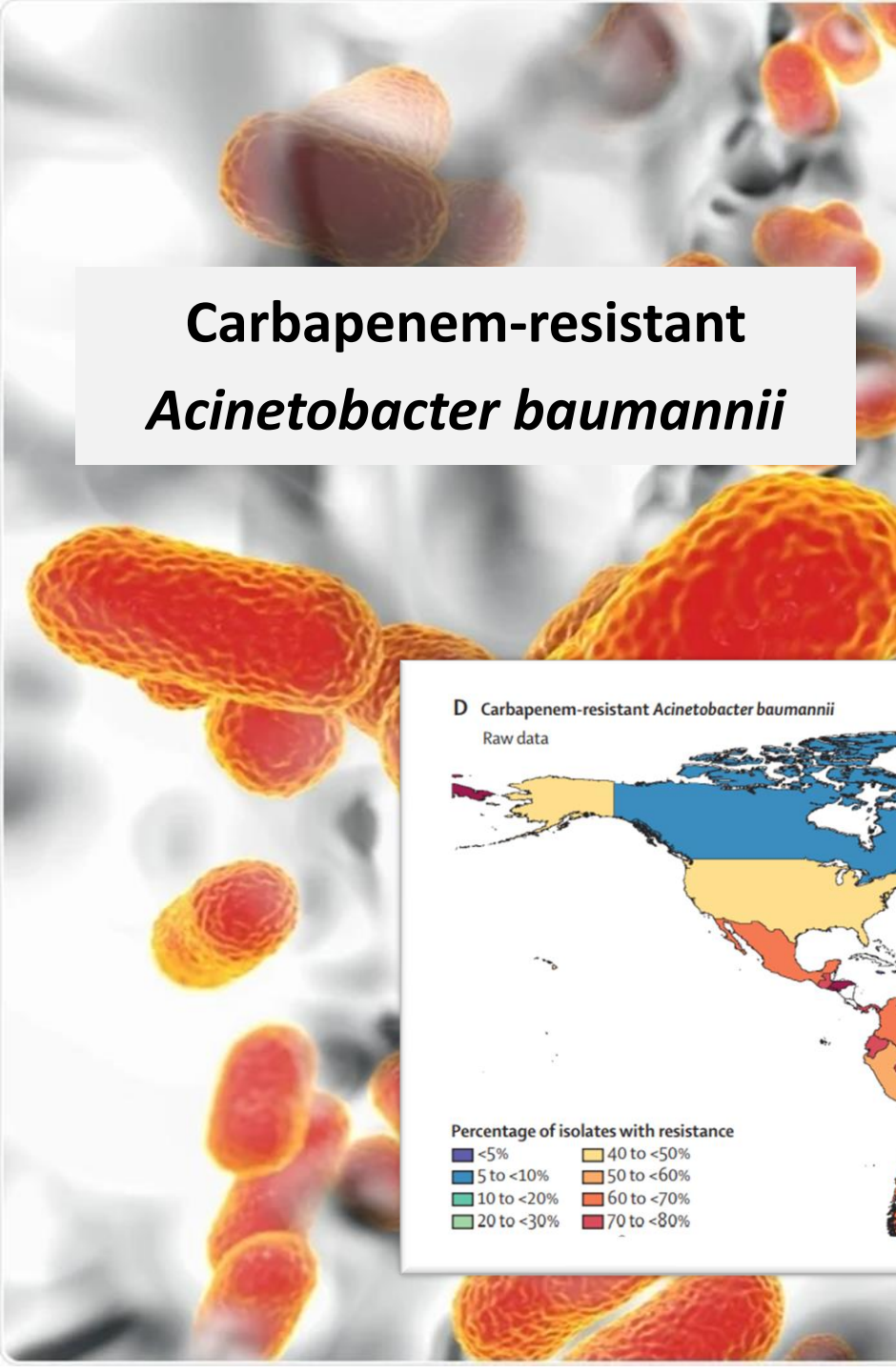
For carbapenem-resistant *Klebsiella pneumoniae* and *Acinetobacter* spp, commonly causing healthcare-associated infections, the number of attributable deaths increased by approximately 50%.



Global deaths (counts) attributable to bacterial AMR by pathogen–drug combination, 2019



Carbapenem-resistant *Acinetobacter baumannii*

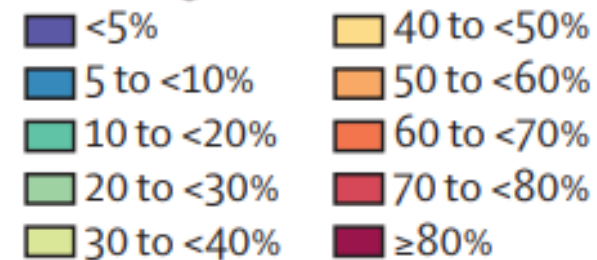


D Carbapenem-resistant *Acinetobacter baumannii*
Raw data

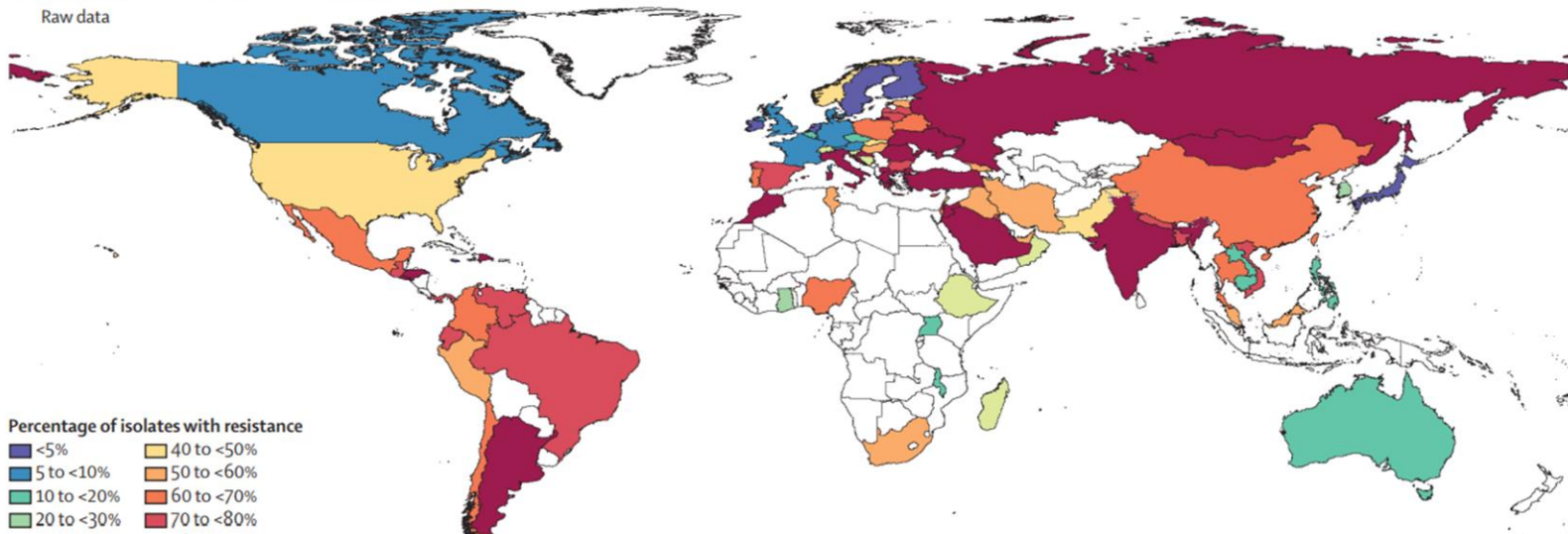
Percentage of isolates with resistance

<5%	40 to <50%
5 to <10%	50 to <60%
10 to <20%	60 to <70%
20 to <30%	70 to <80%

A map of the Balkan Peninsula and surrounding regions. The study area, which includes Bulgaria, Romania, and parts of Greece and Turkey, is highlighted in red. Other countries shown in different colors include Hungary (orange), Serbia (yellow), Croatia (blue), and Slovenia (green). The map shows the Black Sea to the east and the Aegean Sea to the south.



Raw data

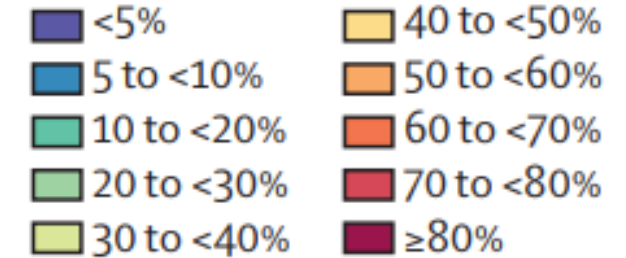


Carbapenem-resistant *Klebsiella pneumoniae*

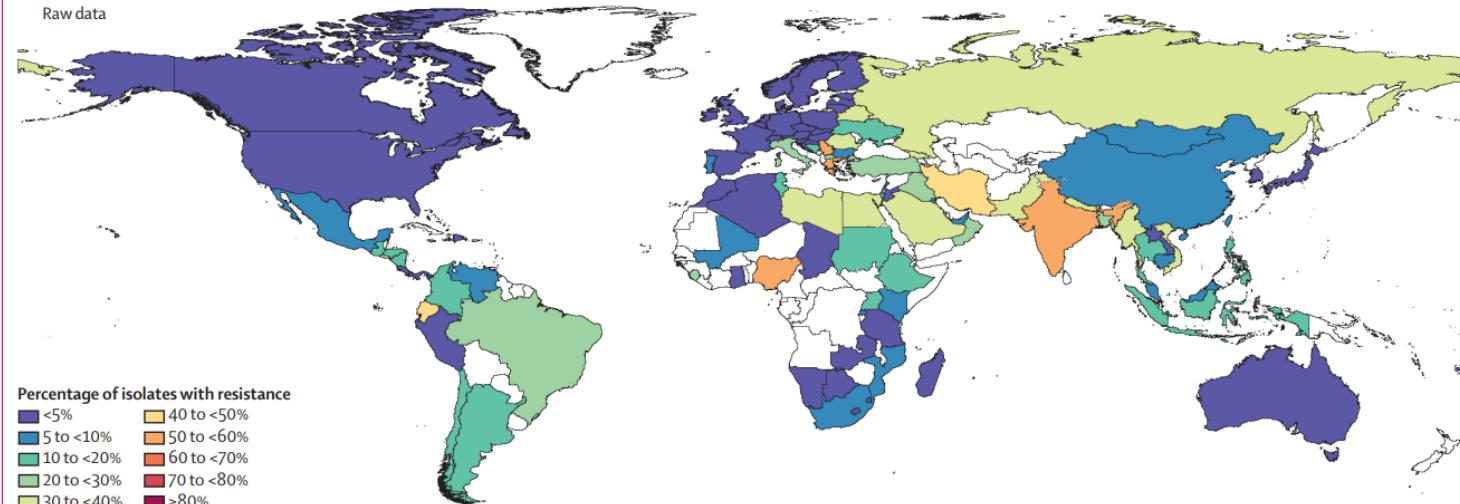
Balkan Peninsula



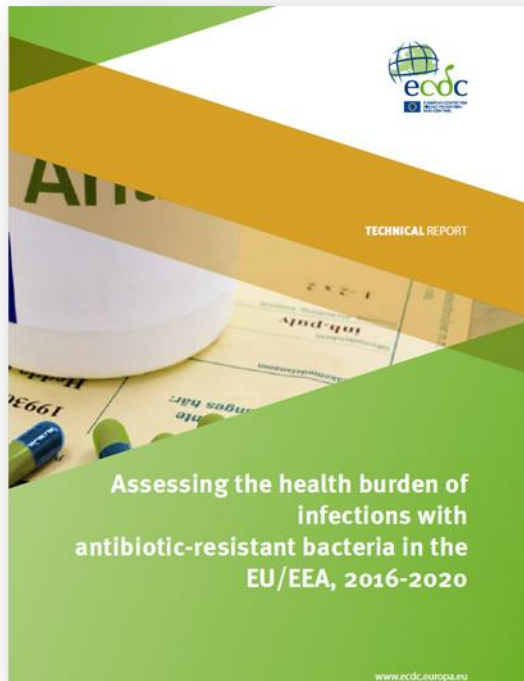
Percentage of isolates with resistance



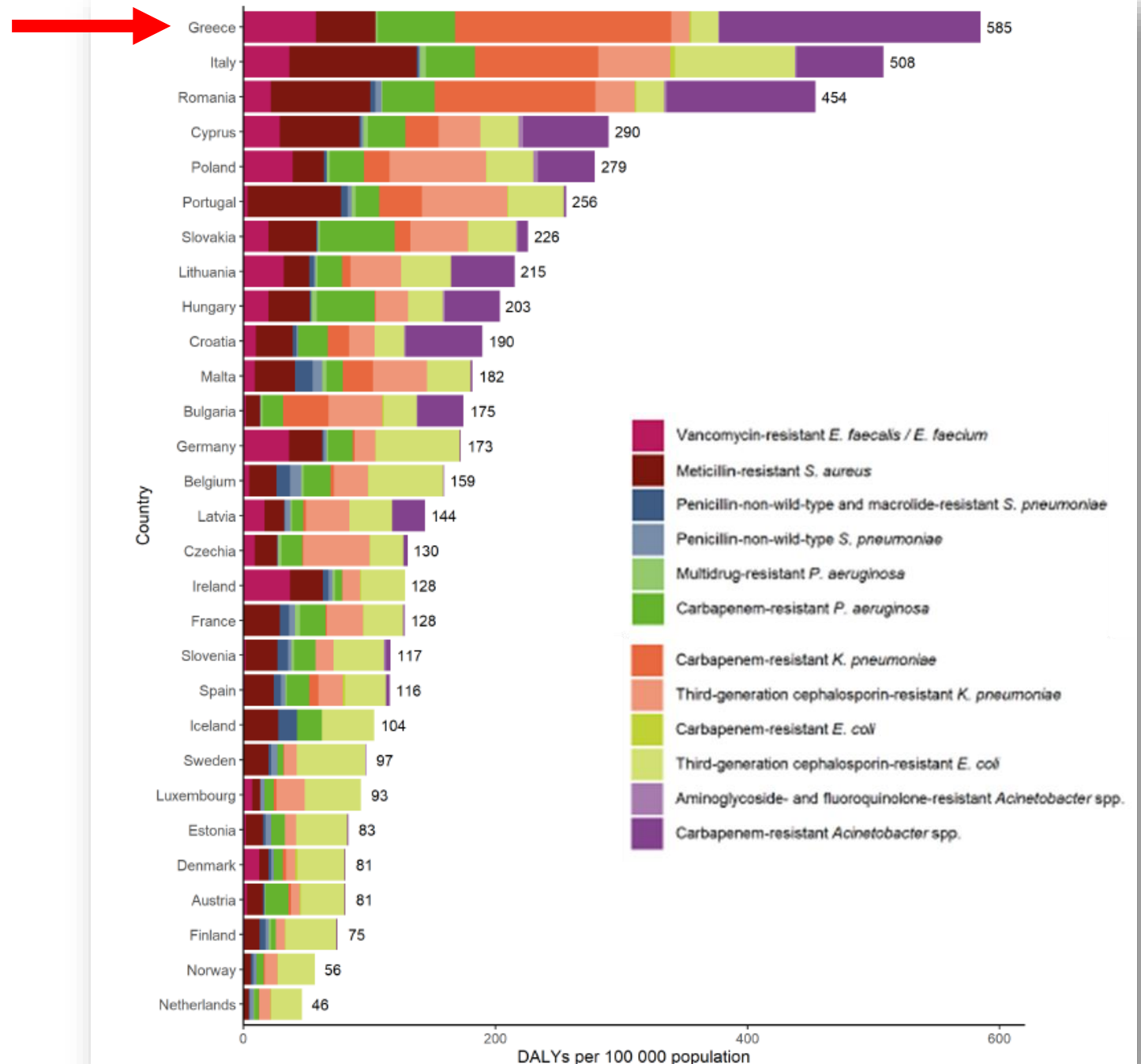
F Carbapenem-resistant *Klebsiella pneumoniae*
Raw data



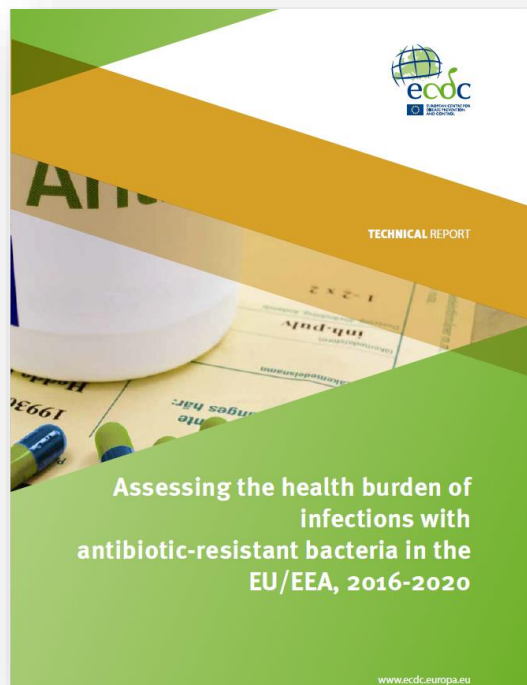
Estimations of the burden of infections with antibiotic-resistant bacteria presented as disability-adjusted life years (DALYs) per 100 000 population by country*, EU/EEA, 2020



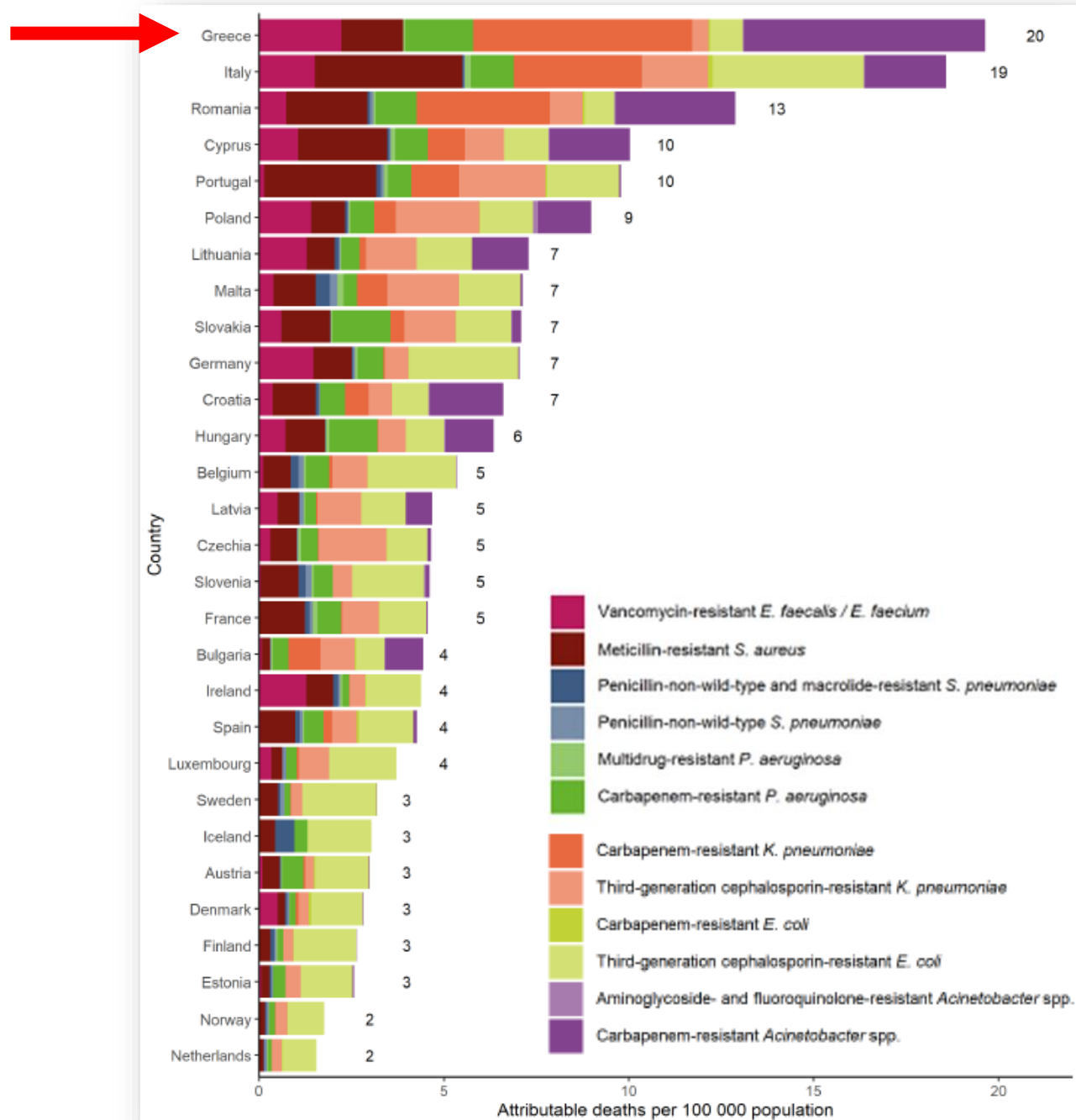
Assessing the health burden of infections with antibiotic-resistant bacteria in the EU/EEA, 2016-2020. Stockholm: ECDC; 2022.



Estimations of the burden of infections with antibiotic-resistant bacteria presented as attributable deaths per 100 000 population by country*, EU/EEA, 2020



Assessing the health burden of infections with antibiotic-resistant bacteria in the EU/EEA, 2016-2020. Stockholm: ECDC; 2022.





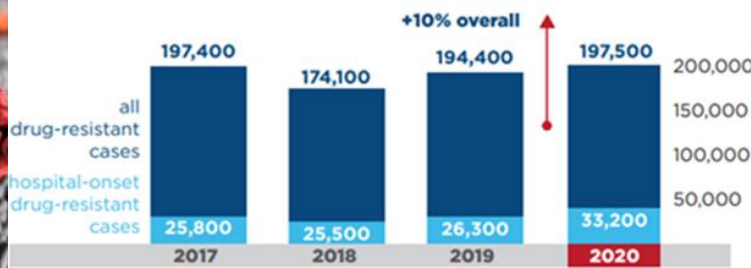
COVID-19 Impacts on 18 Antimicrobial-Resistant Bacteria and Fungi: Threat Estimates

Carbapenem-resistant Enterobacterales (CRE)



The rate of CRE infections increased 35% in hospitals in 2020, emphasizing the important role these difficult-to-treat pathogens play in hospital infections and the need to contain further spread.

Extended Spectrum Beta-lactamase (ESBL) producing Enterobacterales



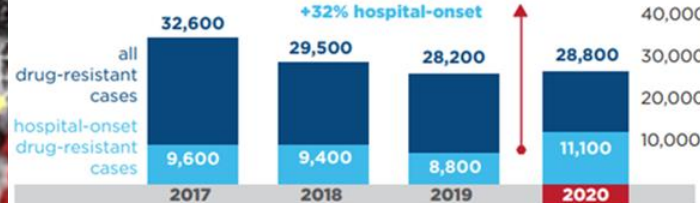
The rate of ESBL-producing Enterobacterales cases increased from 2019 to 2020, with an increased rate in both hospital-onset (32%) and community-onset (7%).

Carbapenem-resistant *Acinetobacter*



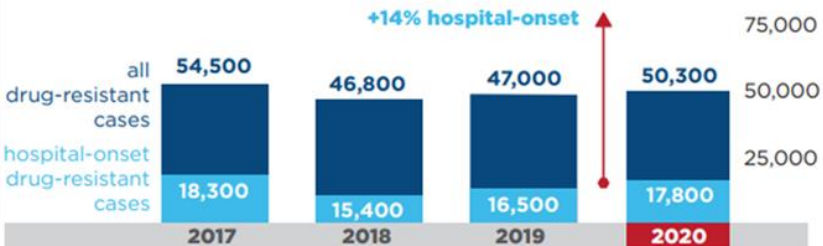
The rate of carbapenem-resistant *Acinetobacter* cases increased overall by 35% in 2020 compared with 2019, driven by hospital-onset cases.

Multidrug-resistant (MDR) *Pseudomonas aeruginosa*



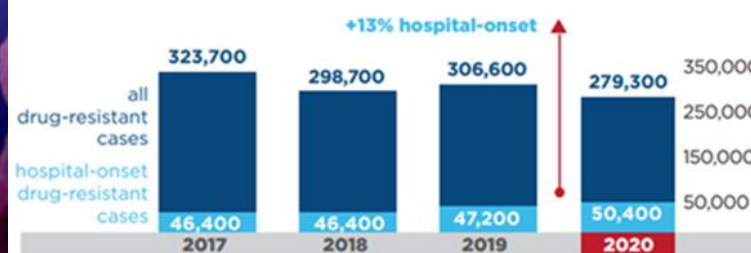
In 2020, the rate of cases of hospital-onset MDR *P. aeruginosa* increased 32% compared to 2019.

Vancomycin-resistant Enterococcus (VRE)



The rate of VRE cases increased 16% from 2019 to 2020, reversing substantial decreases since 2012.

Methicillin-resistant *Staphylococcus aureus* (MRSA)



After years of decreasing, the overall rate of MRSA cases stabilized in 2017 through 2020. The rate of hospital-onset cases increased 13% in 2020, while the rate of community-onset MRSA cases decreased 5% compared with 2019.

TACKLING ANTIMICROBIAL RESISTANCE ON TEN FRONTS



Public awareness



Sanitation and hygiene



Antibiotics in agriculture and the environment



Vaccines and alternatives



Surveillance



Rapid diagnostics



Human capital



Drugs



Global Innovation Fund



International coalition for action

Review on Antimicrobial Resistance



A European One Health Action Plan against Antimicrobial Resistance (AMR).
European Commission, 2017.

Σύσταση του Συμβουλίου σχετικά με την ενίσχυση των δράσεων της ΕΕ για την καταπολέμηση της μικροβιακής αντοχής στο πλαίσιο της προσέγγισης «Μία υγεία» 2023/C 220/01

‘Best buy’ interventions to tackle AMR



- improving hygiene in healthcare facilities, including promotion of handwashing
- stewardship programmes promoting more prudent use of antibiotics to end decades of over-prescription

These will decrease the health burden of AMR – measured in DALYs – by about 40%

- **use of rapid diagnostic tests**
- delayed prescription
- public awareness campaigns

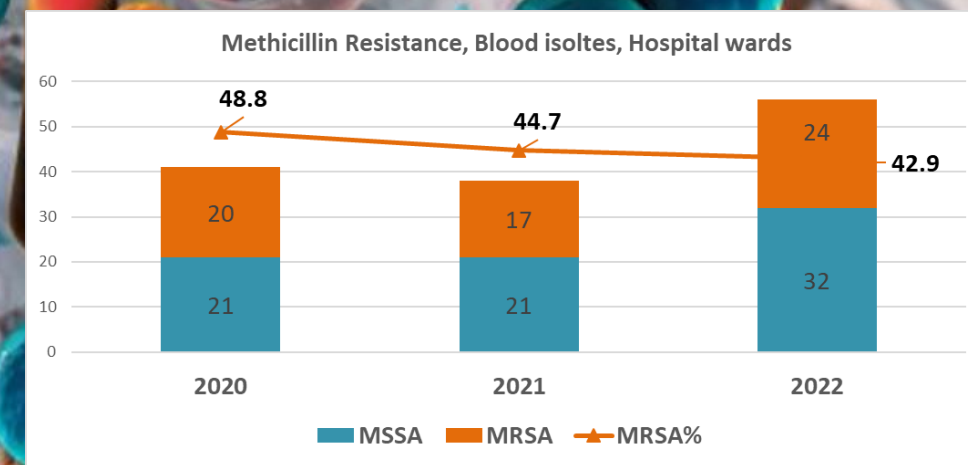
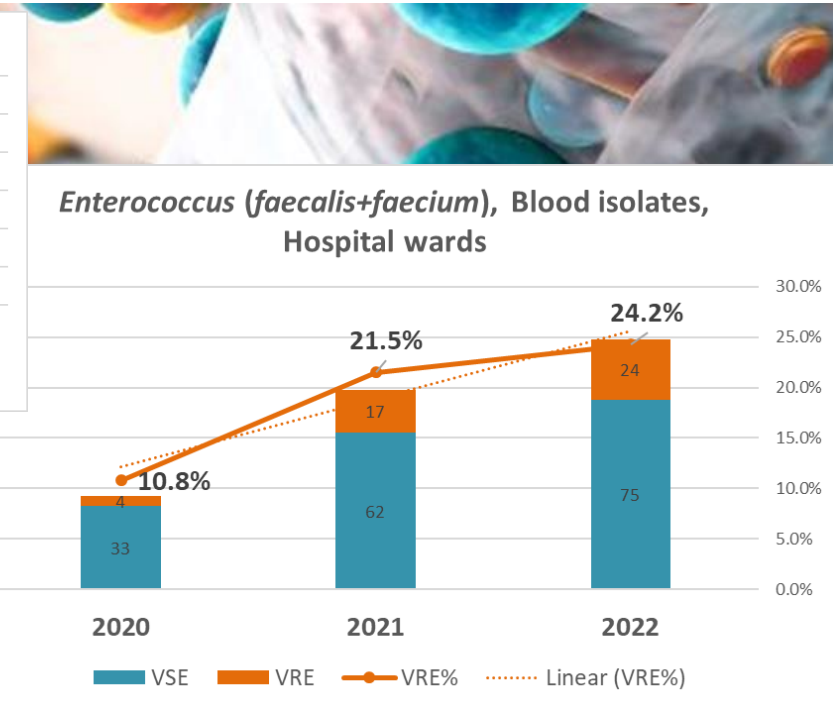
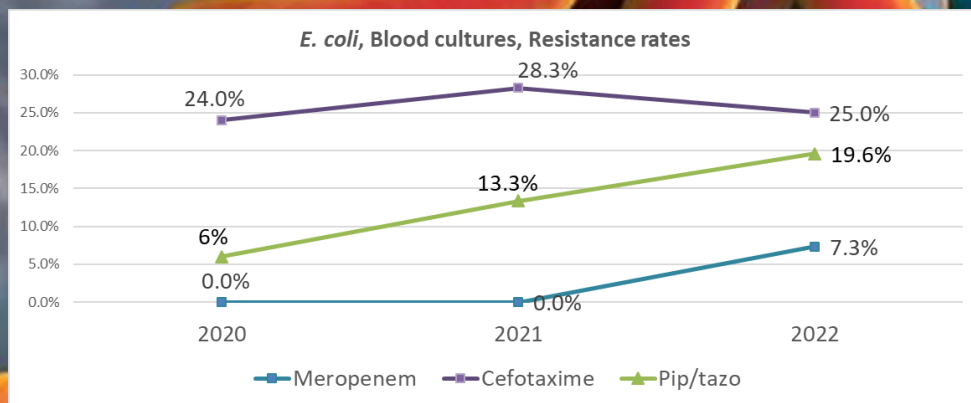
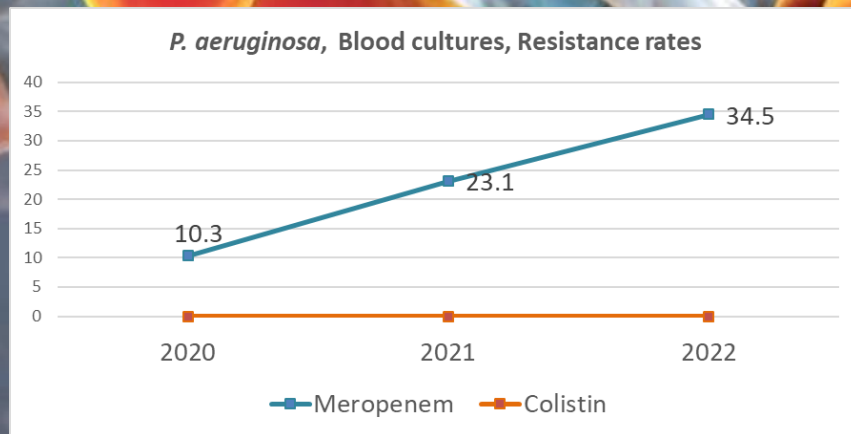
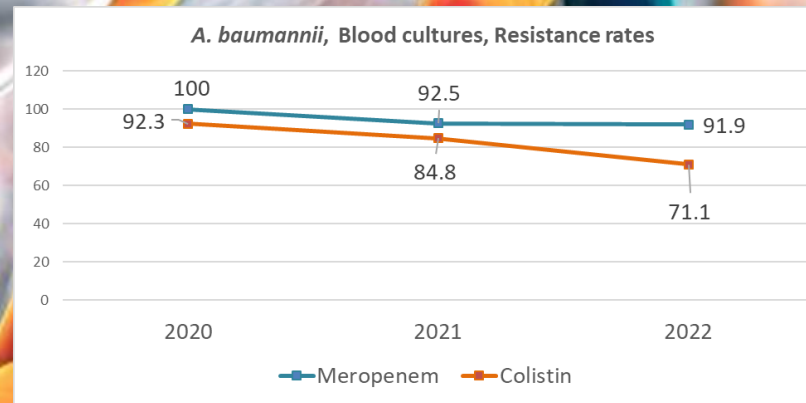
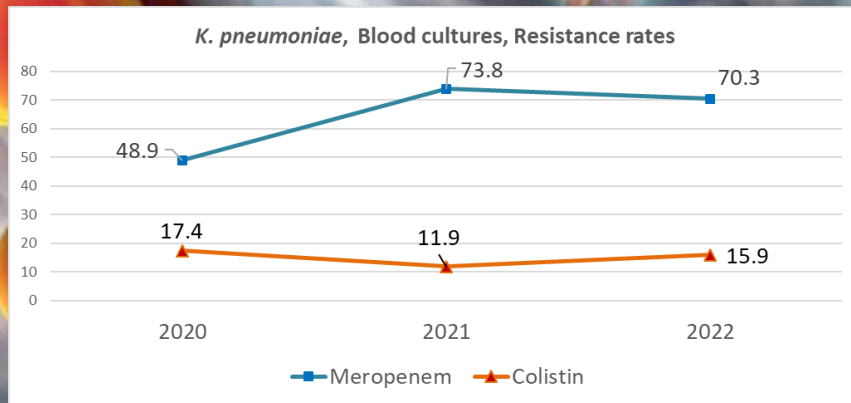
Μικροβιολογικό εργαστήριο και ανίχνευση αντοχής

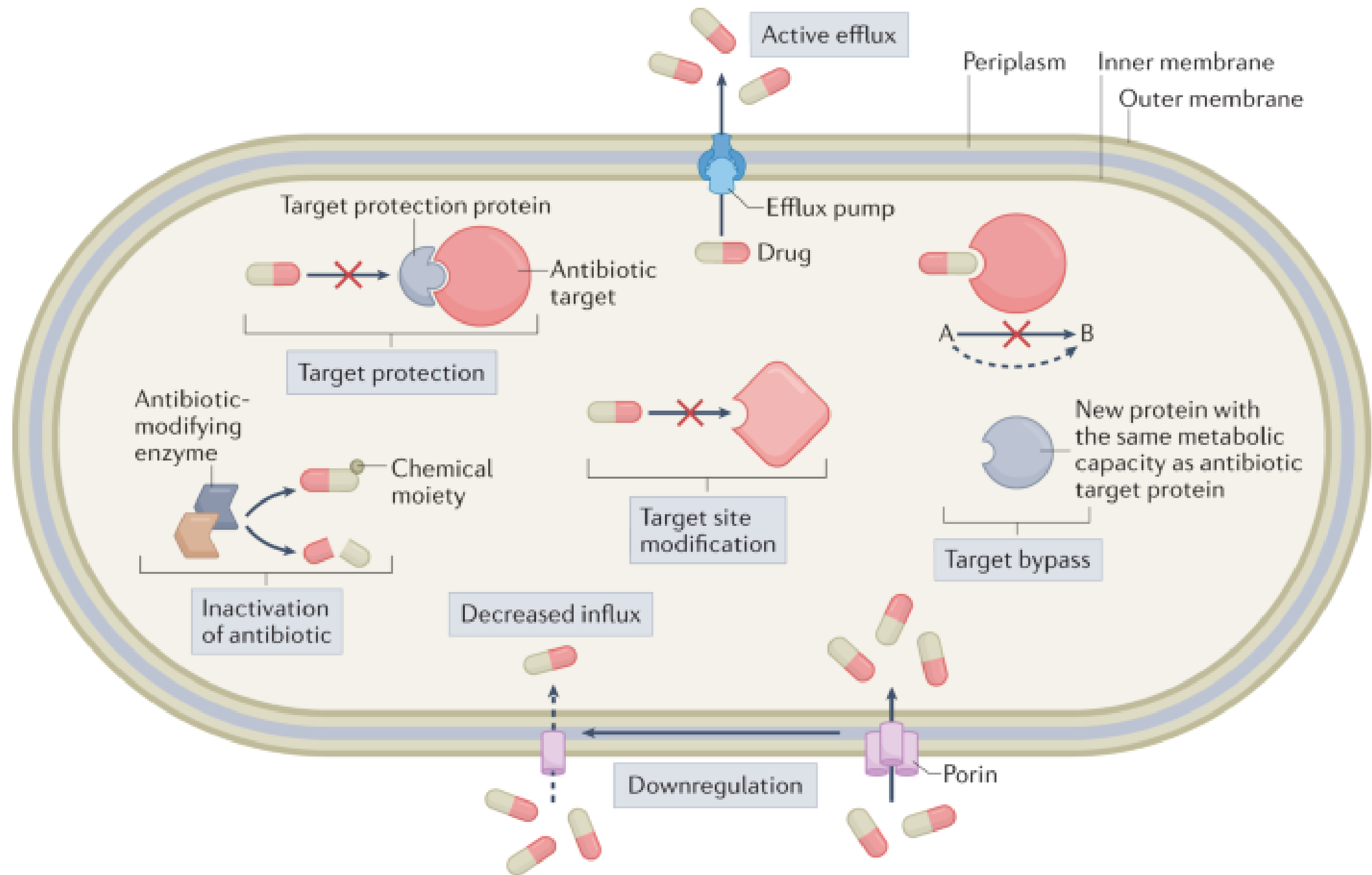


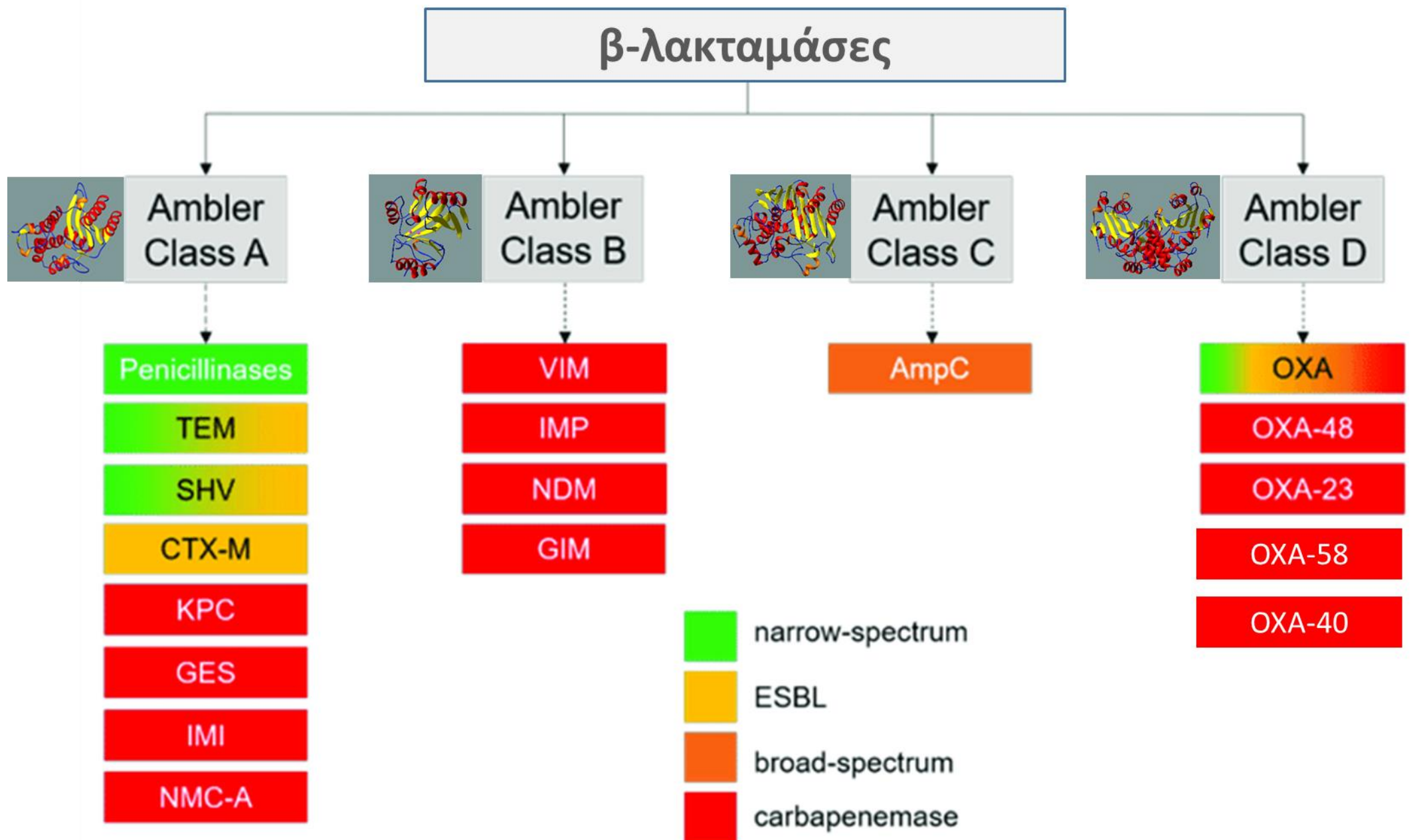
Πληροφορία



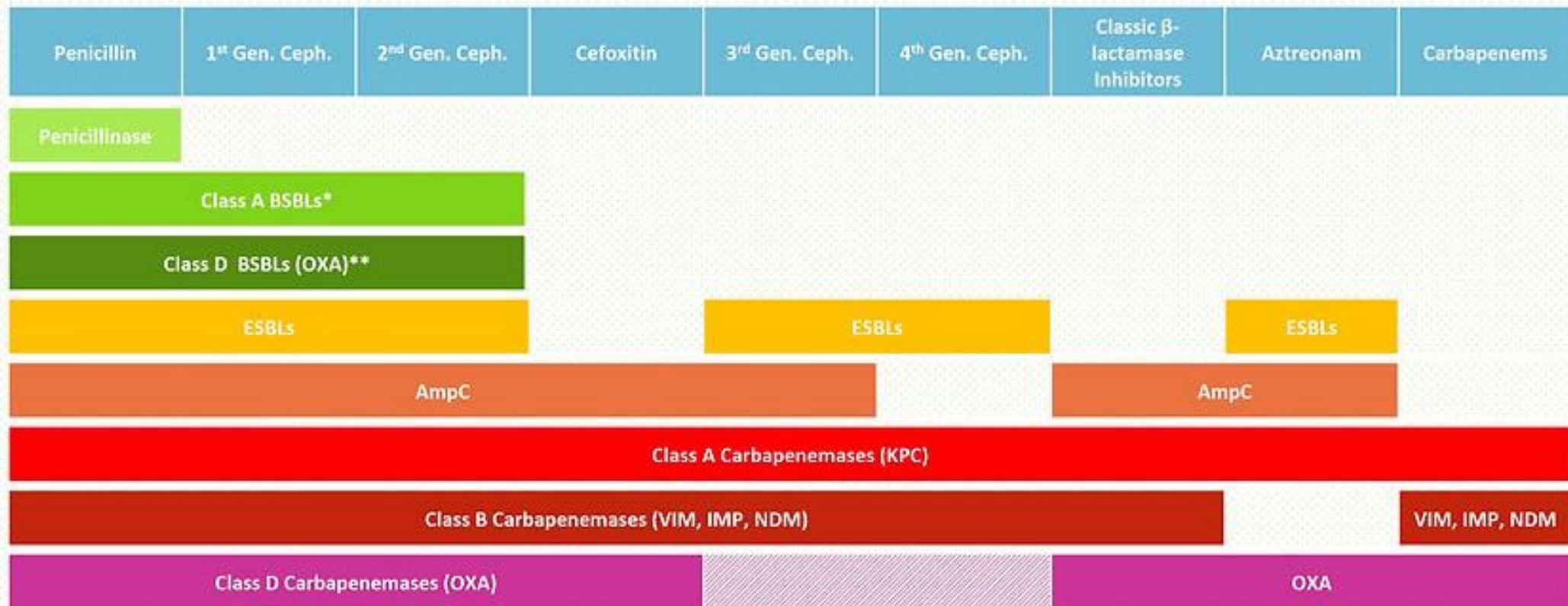
Παράδειγμα CASR: Τζάνειο, Κλινικά Τμήματα εκτός ΜΕΘ







β- λακταμάσες: υδρολυτικό φάσμα



*including benzylpenicillins, aminopenicillins, carboxypenicillins, ureidopenicillin, narrow spectrum cephalosporins (cefazolin and cefuroxime and others)

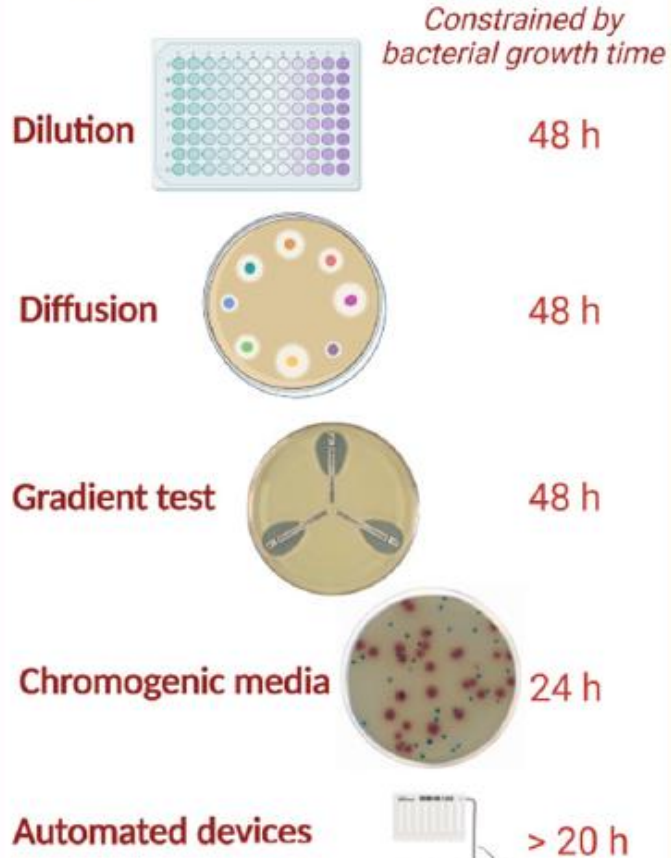
**BSBL substrates plus oxacillin, nafcillin, and dicloxacillin

β- λακταμάσες και νεότερα αντιβιοτικά

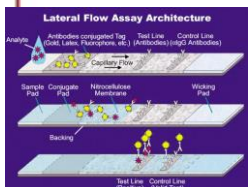
	ESBL	KPC	MBL	AmpC	OXA-48	<i>P. aeruginosa</i> (MDR/XDR)	<i>Acinetobacter</i> (MDR/XDR)	<i>S. maltophilia</i>
Aztreonam/avibactam								
Cefepime/enmetazobactam								
Cefepime/taniborbactam								
Cefepime/zidebactam								
Cefiderocol								
Ceftaroline/avibactam								
Ceftolozane/tazobactam								
Ceftazidime/avibactam								
Imipenem/relebactam								
Meropenem/nacubactam								
Meropenem/vaborbactam								

Μέθοδοι ανίχνευσης της αντοχής

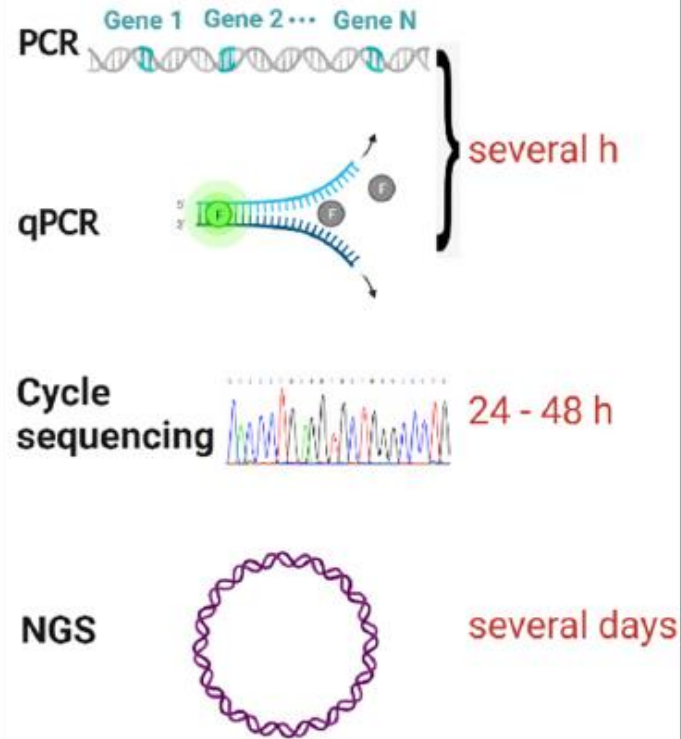
Phenotypic methods



LFIA



Molecular-based methods

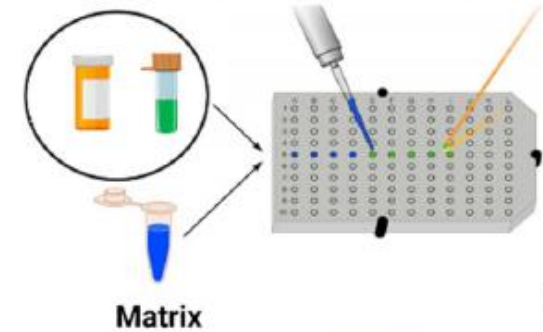


** estimated time depend on a sample analysed (clinical specimen vs. isolated bacterial culture)*

Mass spectrometry

MALDI-TOF MS

Incubation (antibiotic + sample)



several h

** estimated time depend on a sample analysed (clinical specimen vs. isolated bacterial culture)*

Table 1. Differences between phenotypic and genotypic resistance detection methods

Characteristic	Phenotypic methods	Genotypic methods
Question to be answered	Does the antibiotic inhibit bacterial growth at clinically relevant concentrations?	Is a gene or mutation associated with antibiotic resistance present?
Turn-around time	Slow	Fast 
Inoculum needed	High	Low
Provides information about resistance mechanism	No	Yes
Predicts antibiotic susceptibility and resistance	Yes	Sometimes. Only detects a gene or mutation associated with resistance; this may not correlate with phenotypic resistance in all isolates (e.g. if a gene is not expressed). If a singular genotypic resistance type is associated with resistance to a particular antibiotic in a particular bacterial species, its absence infers susceptibility. However, when there is more than one genotypic resistance type associated with resistance, such an inference may not always be correct.
Provides MIC	Yes	No
Cost	Moderate 	High

Banerjee R, Patel R. JAC Antimicrob Resist. 2023;5(1):dlad018.

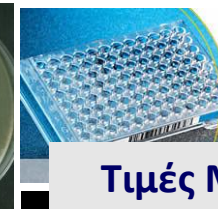


Κλινικό
δείγμα



Ζώνες
αναστολής

ID /AST



Τιμές MIC

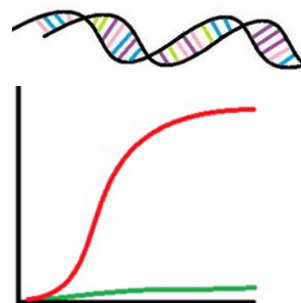


Pheno R Μηχανισμός;

TAT= 48-72 h



CIDT



TAT= 1-4 h

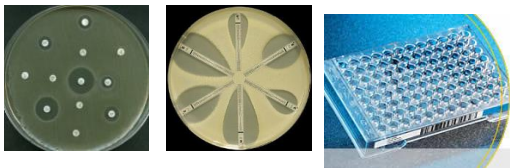


ID / Ανίχνευση γονιδίων αντοχής

Διάγνωση
Κατάλληλη
αγωγή



AST



Ταχύτητα

αξιοπιστία

επαναληψιμότητα



Προαγωγή της φροντίδας
του ασθενούς



Διασφάλιση συνταγογράφησης
του κατάλληλου ΑΒ

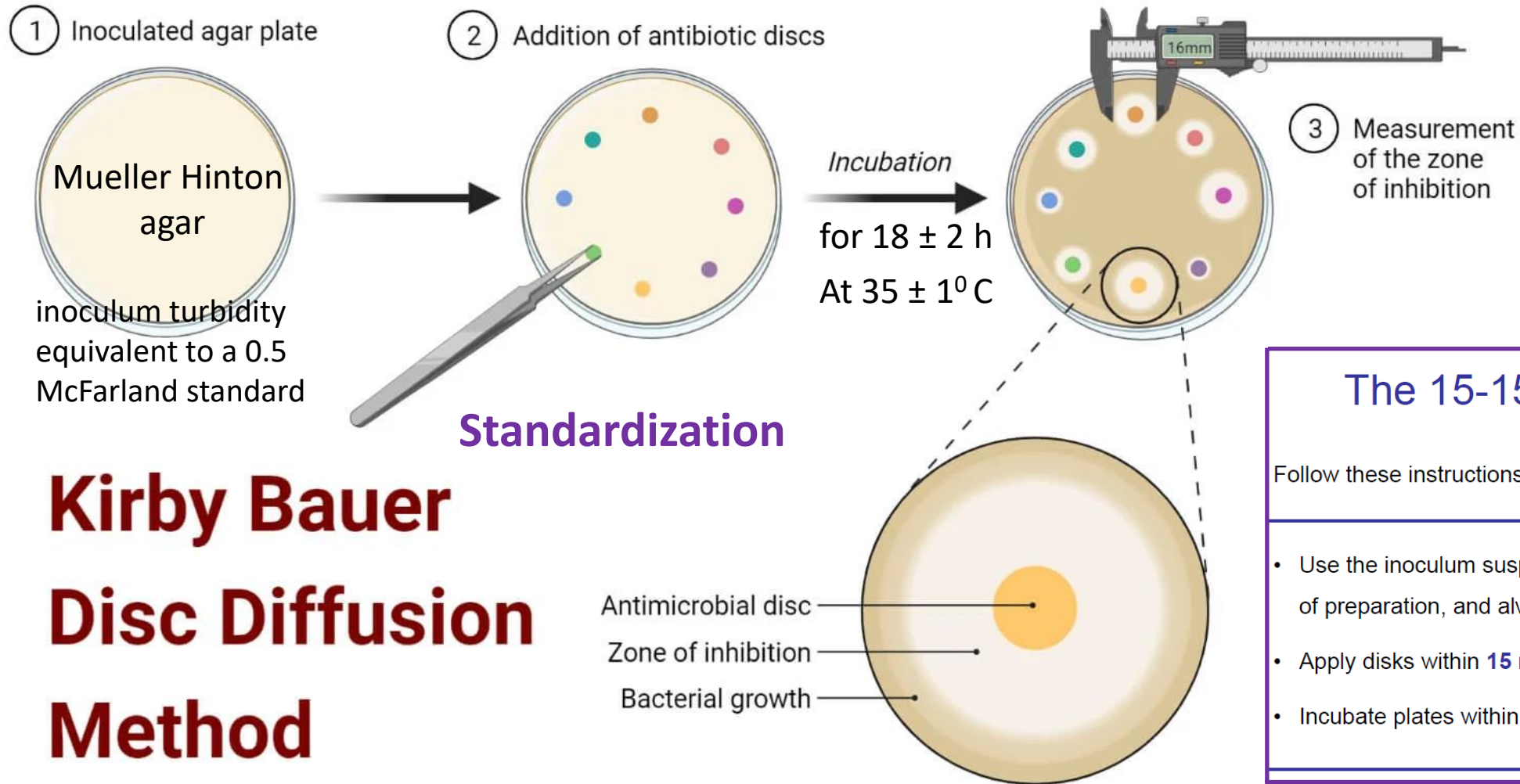


Προαγωγή της ορθολογικής
χρήσης ΑΒ



Ανίχνευση μηχανισμών αντοχής
Εφαρμογή μέτρων για περιορισμό
της διασποράς

Φαινοτυπικές μέθοδοι: Μέθοδος διάχυσης των δίσκων (Kirby-Bauer)

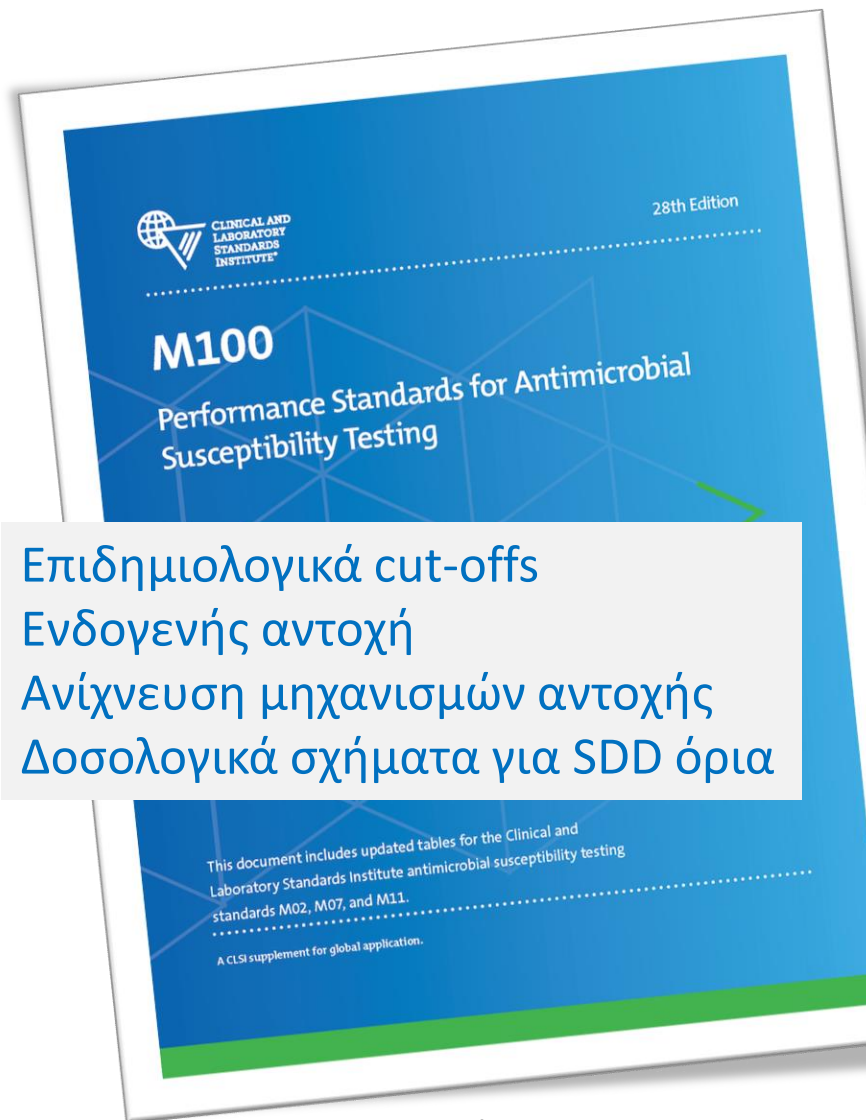


Bauer A.W., Kirby W.M.M., Sherris J.C. and Turck M. 1966.

Antibiotic susceptibility testing by a standardized single disk method. Am. J. Clin. Pathol. 45:493-496

EUCAST disk diffusion method for AST. Version 11.0, January 2023.

Έλεγχος ευαισθησίας και ερμηνεία των αποτελεσμάτων



Μη μέλη: \$200.00

Επιδημιολογικά cut-offs
Ενδογενής αντοχή
Ανίχνευση μηχανισμών αντοχής
Δοσολογικά σχήματα για SDD όρια

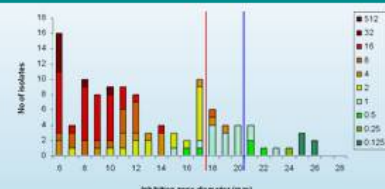
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 **EUCAST** EUROPEAN COMMITTEE ON ANTIMICROBIAL SUSCEPTIBILITY TESTING
European Society of Clinical Microbiology and Infectious Diseases

Clinical breakpoints and dosing of antibiotics

[Organization](#)
[Consultations](#)
[EUCAST News](#)
[New definitions of S, I and R](#)
[Clinical breakpoints and dosing](#)
[Rapid AST in blood cultures](#)
[Expert rules and expected phenotypes](#)
[Resistance mechanisms](#)
[Guidance documents](#)
[SOP](#)
[MIC and zone distributions and ECOFFs](#)
[AST of bacteria](#)
[AST of mycobacteria](#)
[AST of fungi](#)
[AST of veterinary pathogens](#)
[Frequently Asked Questions \(FAQ\)](#)
[Meetings](#)
[Rationale documents and publications](#)

E.coli with cefotaxime 5 ug
112 non-consecutive isolates chose because of resistance



Clinical breakpoints - breakpoints and guidance
January 2, 2023

- Clinical breakpoints (v 13.1) - new file for printing** (29 June, 2023) to make new breakpoints for anaerobic bacteria available (there are no changes beyond the tab for anaerobic bacteria)
- Clinical breakpoints (v 13.1) - new file for screen** (29 June, 2023) to make new breakpoints for anaerobic bacteria available (there are no changes beyond the tab for anaerobic bacteria)

Breakpoint tables

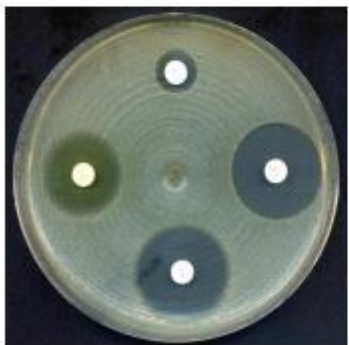
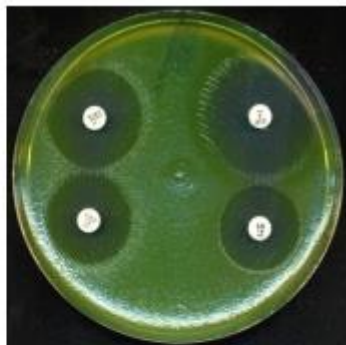
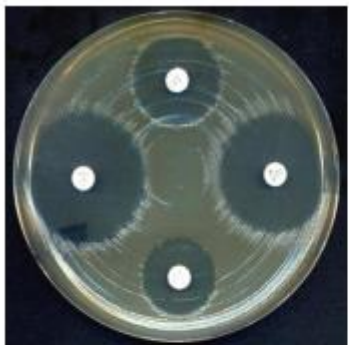
- Breakpoints bacteria (print)
- Breakpoints bacteria (screen)
- Breakpoints fungi
- Dosing table

Make sure the device you are using for the presentation of tables can correctly display

Κλινικά όρια ευαισθησίας
Δοσολογικά σχήματα
Επιδημιολογικά cut-offs (ECOFs)
Expert rules / Ενδογενής αντοχή
Ανίχνευση μηχανισμών αντοχής
Rational documents (breakpoint background)

Μέθοδος διάχυσης των δίσκων Kirby-Bauer

S, I, R



Enterobacterales*

Expert Rules and Expected Phenotypes

For abbreviations and explanations of breakpoints, see the Notes sheet

EUCAST Clinical Breakpoint Tables v. 13.1, valid from 2023-06-29

Carbapenems ¹	MIC breakpoints (mg/L)			Disk content (μg)	Zone diameter breakpoints (mm)			Notes
	S ≤	R >	ATU		S ≥	R <	ATU	
Doripenem	1	2		10	24	21		1. Some isolates that produce carbapenemase are categorised as susceptible with the current breakpoints and should be reported as tested, i.e. the presence or absence of a carbapenemase does not in itself influence the categorisation of susceptibility. Carbapenemase detection and characterisation are recommended for public health and infection control purposes. For carbapenemase screening, a meropenem screening cut-off of >0.125 mg/L (zone diameter <28 mm) is recommended.
Ertapenem	0.5	0.5		10	25	25		
Imipenem, Enterobacterales except Morganellaceae	2	4		10	22	19		2. The intrinsically low activity of imipenem against <i>Morganella morganii</i> , <i>Proteus</i> spp. and <i>Providencia</i> spp. requires the high exposure of imipenem.
Imipenem ² , Morganellaceae	0.001	4		10	50	19		
Imipenem-relebactam, Enterobacterales except Morganellaceae	2 ³	2 ³		10-25	22	22	20-22	3. For susceptibility testing purposes, the concentration of relebactam is fixed at 4 mg/L.
Meropenem (indications other than meningitis)	2	8		10	22	16		
Meropenem (meningitis)	2	2		10	22	22		4. For susceptibility testing purposes, the concentration of vaborbactam is fixed at 8 mg/L.
Meropenem-vaborbactam	8 ⁴	8 ⁴		20-10	20	20	15-19 ^A	

Monobactams	MIC breakpoints (mg/L)			Disk content (μg)	Zone diameter breakpoints (mm)			Notes
	S ≤	R >	ATU		S ≥	R <	ATU	
Aztreonam ¹	1	4		30	26	21		1. The aztreonam breakpoints for <i>Enterobacterales</i> will detect clinically important resistance mechanisms (including ESBL). Some isolates that produce beta-lactamases are susceptible to aztreonam with these breakpoints and should be reported as tested, i.e. the presence or absence of an ESBL does not in itself influence the categorisation of susceptibility. ESBL detection and characterisation are recommended for public health and infection control purposes.

Fluoroquinolones	MIC breakpoints (mg/L)			Disk content (μg)	Zone diameter breakpoints (mm)			Notes
	S ≤	R >	ATU		S ≥	R <	ATU	
Ciprofloxacin, <i>Salmonella</i> spp. ¹	0.06	0.06			Note ^A	Note ^A		1. There is clinical evidence for ciprofloxacin to indicate a poor response in systemic infections caused by <i>Salmonella</i> spp. with low-level ciprofloxacin resistance (MIC >0.06 mg/L). The available data relate mainly to <i>Salmonella</i> Typhi but there are also case reports of poor response with other <i>Salmonella</i> species.
Ciprofloxacin (indications other than meningitis)	0.25	0.5	0.5	5	25	22	22-24	
Ciprofloxacin (meningitis) ²	0.125	0.125			Note ^B	Note ^B		2/B. In meningitis, where low-level ciprofloxacin resistance must be excluded, either perform an MIC test, or infer susceptibility from the pefloxacin 5 μg screening test.
Pefloxacin (screen only)	NA	NA		5	24 ^{A,B,C}	24 ^{A,B,C}		
Delafloxacin, <i>E. coli</i>	0.125	0.125			Note ^D	Note ^D		A. Tests with a ciprofloxacin 5 μg disk will not reliably detect low-level resistance in <i>Salmonella</i> spp. Perform an MIC test, or infer susceptibility from the pefloxacin 5 μg screening test.
Levofloxacin	0.5	1		5	23	19		
Moxifloxacin	0.25	0.25		5	22	22		C. The pefloxacin screening test can also be used to detect fluoroquinolone resistance mechanisms in other <i>Enterobacterales</i> such as <i>E. coli</i> , <i>K. pneumoniae</i> and <i>Shigella</i> spp.
Nalidixic acid (screen only)	NA	NA			NA	NA		
Norfloxacin (uncomplicated UTI only)	0.5	0.5		10	24	24		D. A disk diffusion test awaits action from the responsible pharmaceutical company.
Ofloxacin	0.25	0.5		5	24	22		

The European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters. Version 13.1, 2023. <http://www.eucast.org>.

**Methodology - EUCAST rapid antimicrobial susceptibility testing (RAST)
directly from positive blood culture bottles.**

Version 4.0

April 2023

EUCAST RAST Breakpoint Tables version 6.1 (2023-06-07)

European Committee on Antimicrobial Susceptibility Testing

**Zone diameter breakpoint tables for rapid antimicrobial susceptibility testing (RAST)
directly from blood culture bottles**

Version 6.1, valid from 2023-06-07

**Screening for ESBL and carbapenemases in *E. coli* and
K. pneumoniae for epidemiological purposes as part of the
RAST procedure.**

EUCAST Guidelines for detection of resistance mechanisms and specific resistance of
clinical and/or epidemiological importance using EUCAST rapid antimicrobial
susceptibility testing (RAST) directly from positive blood culture bottles.

Version 2.0

April 2022

EUCAST, Rapid AST

- Χρήση με φιάλες BACTEC (BD), BacT/ALERT (bioMérieux) και VersaTREK (Thermo Fisher)
- 0 – 18 h μετά τη θετικοποίηση της φιάλης
- Απευθείας ενοφθαλμισμός (MH, MH-F) με 125±25 μl από τη θετική φιάλη
- Το μικροβιακό είδος **πρέπει να είναι γνωστό** πριν την ερμηνεία των αποτελεσμάτων

Organism	Incubation time	Medium
<i>Escherichia coli</i> <i>Klebsiella pneumoniae</i> <i>Staphylococcus aureus</i>	4, 6 and 8 hours 16-20 hours	MH
<i>Pseudomonas aeruginosa</i>	6 and 8 hours 16-20 hours	MH
<i>Acinetobacter baumannii</i> <i>Enterococcus faecalis</i> <i>Enterococcus faecium</i>	4, 6 and 8 hours	MH
<i>Streptococcus pneumoniae</i>	4, 6 and 8 hours 16-20 hours	MH-F

Ταινίες διαβαθμισμένης συγκέντρωσης αντιβιοτικών

1



Prepare inoculum to
0.5 – 1 McFarland

2



Inoculate plate,
allowing drying time of
15 – 20 minutes

3



Position ETEST
strip and incubate
overnight

4



Read and interpret
results

The MIC value is read from the scale in terms of $\mu\text{g/mL}$
at complete inhibition of bacterial growth, where the
pointed end of the ellipse intersects the strip.

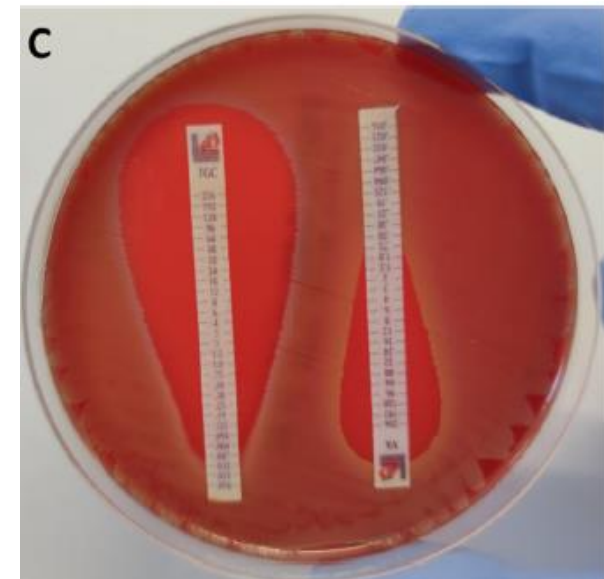
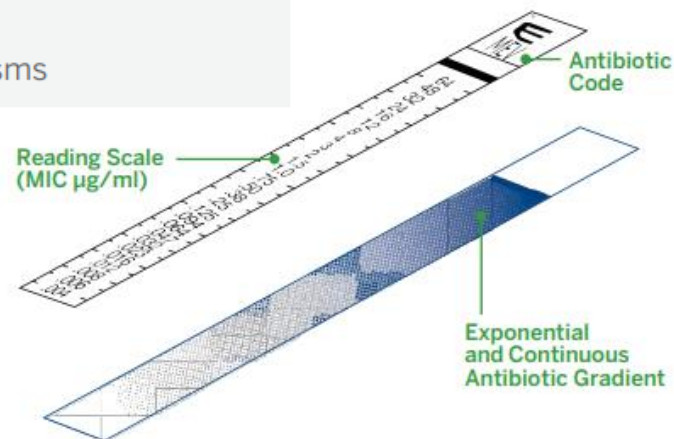
SET UP TIME:

< 5 Minutes

TIME TO RESULT:

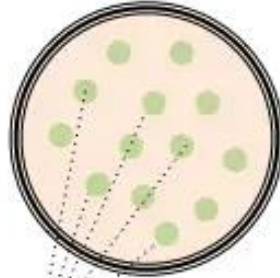
16 – 20 hours for most organisms

Τιμή MIC



Μέθοδος μακροαραιώσεων σε ζωμό

1. Obtain isolated colonies of bacterial strain to test.

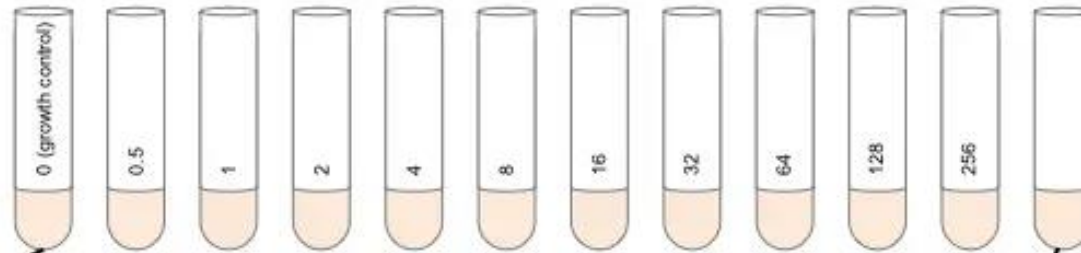


2. Combine 4-5 colonies and culture overnight in rich media broth.



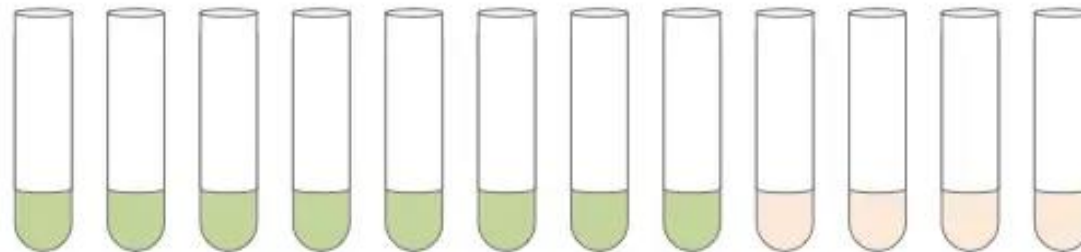
Broth dilution method for measuring minimum inhibitory concentration of antibiotics

3. After overnight incubation shown at left, add rich broth with appropriate dilution series of test antibiotic to test tubes. Example concentrations (mg/L) are shown below. Inoculate bacteria to a final density of 5×10^5 cfu/ml.



No bacteria; broth control

4. Plate aliquot of growth control (i.e., no antibiotic added) to verify cfu/ml counts of viable bacteria. Incubate overnight and count colonies.

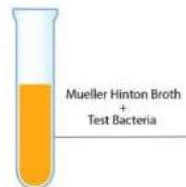


5. After overnight incubation, check cultures for growth. The MIC is the lowest concentration of antibiotic that prevents visible growth. In this example, the MIC is 64 mg/L.

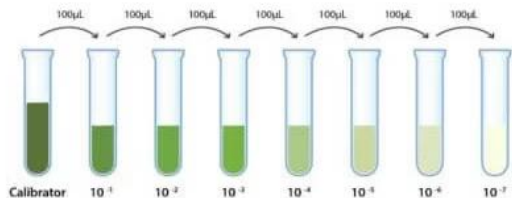
Correct inoculum
 5×10^5 CFU/ml

Μέθοδος μικροαραίωσης σε ζωμό (BMD)

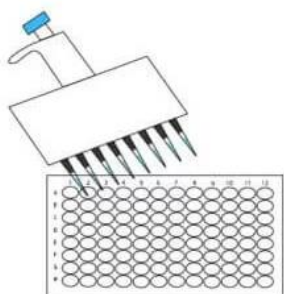
1. Preparation of test inoculum



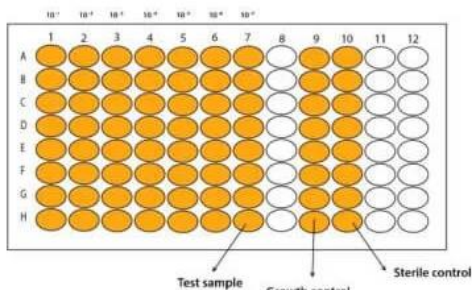
2. Preparation of different dilutions of antimicrobial agent



3. Inoculation on 96 well plate



(Antimicrobial agents are transferred into a 96-well microtiter plate and inoculated with bacterial suspension)

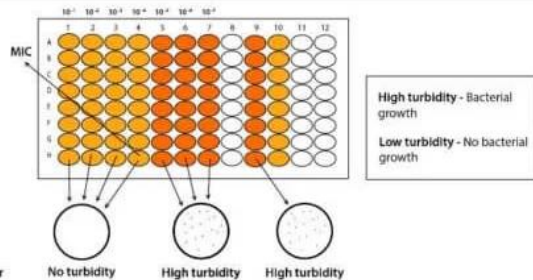


Kept for incubation at 37 °C for 18 hours

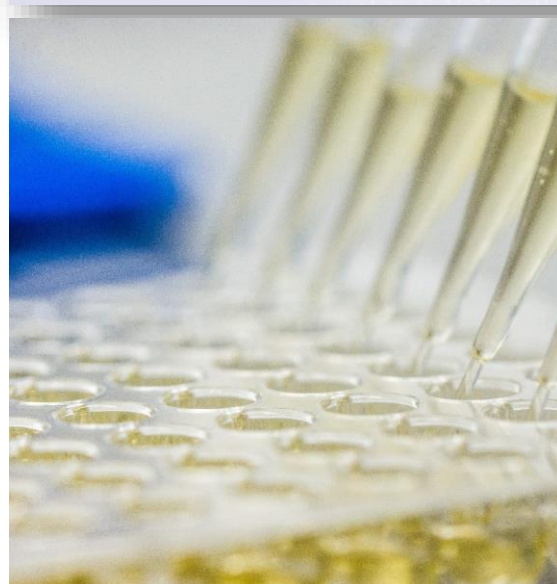
4. Results - Determination of Minimum Inhibitory Concentration (MIC)



(Turbidity of the sample is determined)



The lowest concentration of antimicrobial agent that is capable of inhibiting bacterial growth is called MIC.



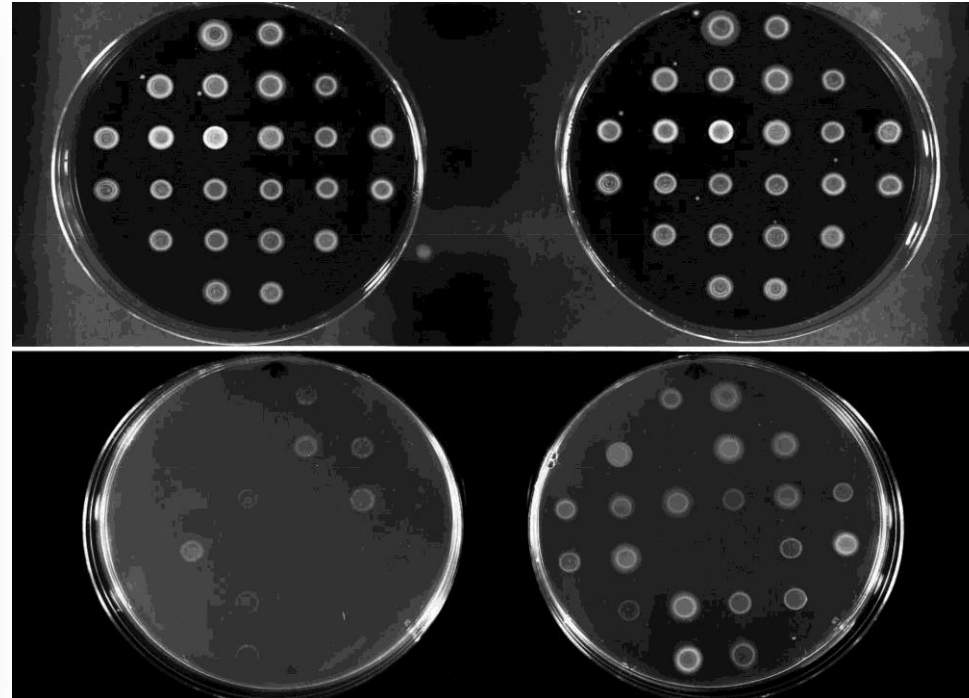
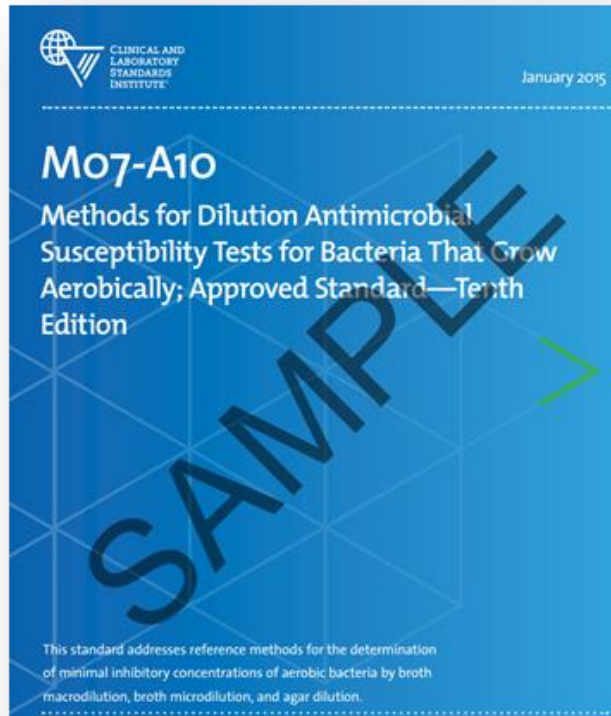
EUCAST EUROPEAN COMMITTEE
ON ANTIMICROBIAL
SUSCEPTIBILITY TESTING
European Society of Clinical Microbiology and Infectious Diseases

Version 4.0, January 2022

EUCAST reading guide for broth microdilution

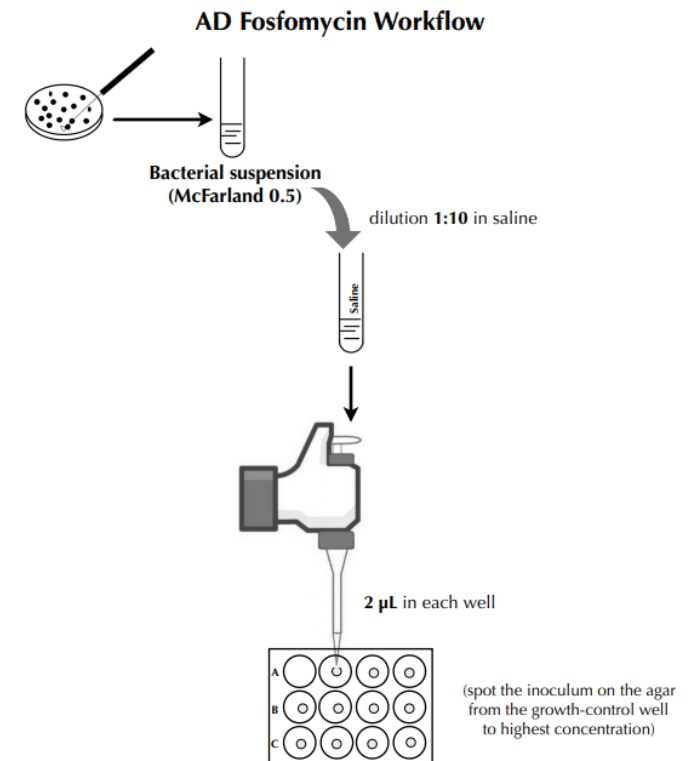
- BMD is the reference method for AST of rapidly growing aerobic bacteria, except for mecillinam and fosfomycin, where agar dilution is the reference method.
- EUCAST recommends testing according to the International Standard ISO 20776-1 (with the use of MH-F broth for fastidious organisms).

Μέθοδος αραίωσης σε άγαρ (agar dilution)

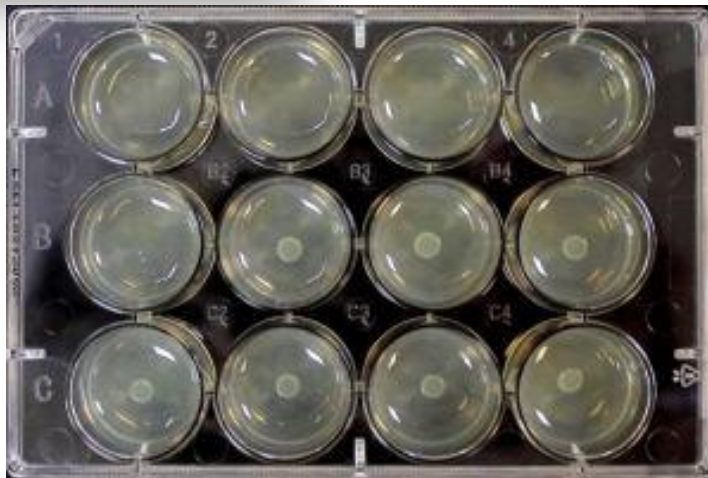


mecillinam and fosfomycin:
agar dilution is
the reference method

Fosfomycin Agar Dilution Panel, Liofilchem



Fosfomycin MIC range: 0.25 - 256 µg/mL



A	256	128	64	32
B	16	8	4	2
C	1	0.5	0.25	Control

Growth-control: No antimicrobial agent in the well.

Αυτοματοποιημένα συστήματα



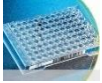
Detection of AMR facilitated by Advanced Expert System (AES).

The instruments read the kinetics of the growth using an optical system with photometer/multichannel fluorimeter readings to assess fluorescence / turbidity /colorimetric signals



- Each of the systems has inherent advantages and limitations
- Results can vary widely by antimicrobial drugs, software versions, and cards
- Some of the systems are not reliable for correct categorization of susceptibility for certain drugs, leading to wrong classifications.
- Software updates and synchronization of breakpoints according to the current standards are mandatory.
- Panels usually contain only several concentrations of each antimicrobial agent, and the resulting MIC is not always given as an exact value.

Advantages and disadvantages of the common methods of antimicrobial susceptibility testing

Method	Advantage	Disadvantage	Comments
Broth dilution 	Well-standardised Harmonised Commercially available tests are easy to perform	Time-consuming Individual mistakes	Quantitative **
Agar Dilution	Well-standardised Suitable for testing a large number of isolates	Time-consuming Limited concentration of antimicrobial agents	Quantitative Possible automation in part
Disk diffusion 	Simple to perform Low cost Simple and fast interpretation The high number of test antibiotics per test High flexibility in antibiotic selection Detection of resistance patterns Mass use and the possibility of automatisisation A number of a different use (AST, identification, screening, etc.) Detection of heteroresistant population or contamination	Time-consuming No MIC value The inability for some antibiotics to be tested	Qualitative *
Gradient test	Convenient and flexible Simple to perform Does not require expertise Detection of resistance patterns	Relatively expensive Relatively long incubation	Quantitative
Automated systems	Simple to perform	Relatively expensive	Semi-quantitative ***

Όχι τιμές MIC

Antibiotic resistance threats

Carbapenem-resistant
Enterobacterales (CRE)

Extended-spectrum
cephalosporin resistance in
Enterobacterales suggestive of
extended-spectrum β -lactamase
(ESBL) production

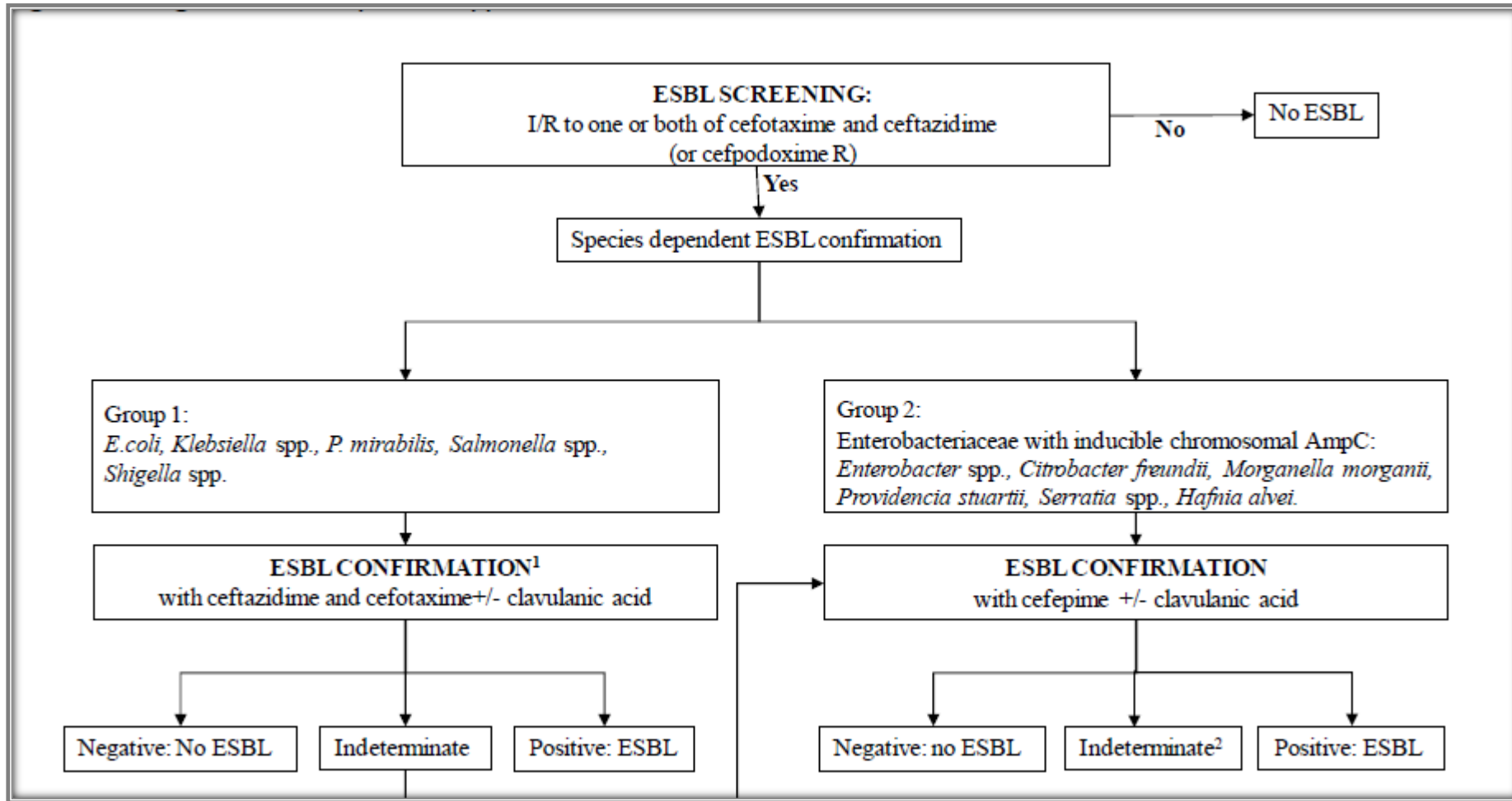
Multidrug-resistant (MDR)
Pseudomonas aeruginosa

Carbapenem-resistant
Acinetobacter species

Methicillin-resistant
Staphylococcus aureus
(MRSA)

Vancomycin-resistant
Enterococcus (VRE)

Αλγόριθμος για φαινοτυπική ανίχνευση ESBLs σε *Enterobacterales*



¹If cefoxitin has an MIC >8 mg/L, perform cefepime +/- clavulanic acid confirmation test

If confirmation with cefepime +/- clavulanic acid is still indeterminate, genotypic testing is required.

Φαινοτυπική ανίχνευση ESBL

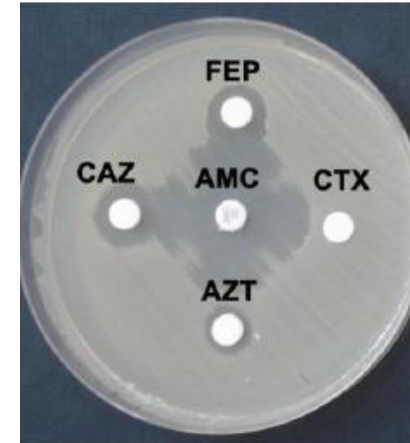
Bionumber: 0405610450106601

Organism Quantity:

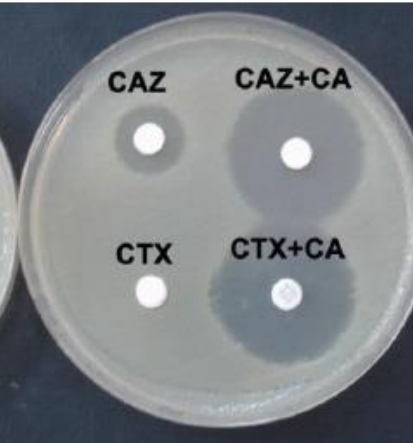
Selected Organism: Escherichia coli

Antimicrobial	MIC	Interpretation	Antimicrobial	MIC	Interpretation
ESBL	POS	+	Ertapenem	≤ 0.12	S
Temocillin			Imipenem	≤ 0.25	S
Urine	≤ 4	I	Meropenem		
Other	≤ 4		Meningitis	≤ 0.25	S
Ampicillin	≥ 32	R	Other	≤ 0.25	S
Amoxicillin/Clavulanic Acid	8	S	Amikacin	2	S
Piperacillin/Tazobactam	≤ 4	S	Gentamicin	≤ 1	S
Cefuroxime	≥ 64	R	Tobramycin	≤ 1	S
Cefuroxime Axetil	≥ 64	R	Ciprofloxacin		
Cefoxitin	≤ 4	IE	Meningitis	0.5	
Cefditoren			Other	0.5	I
Cefixime	≥ 4	R	Levofloxacin	1	I
Cefotaxime			Moxifloxacin	0.5	R
Meningitis	≥ 64	R	Minocycline	(-)	(-)
Other	≥ 64	R	Tetracycline	(-)	(-)
Ceftazidime	8	R	Tigecycline	≤ 0.5	S
Ceftriaxone			Fosfomycin		
Meningitis	≥ 64	R	Oral	≤ 16	
Other	≥ 64	R	Other	≤ 16	S
Ceftazidime/Avibactam	≤ 0.12	S	Nitrofurantoin	≤ 16	S
Ceftolozane/Tazobactam	≤ 0.25	S	Chloramphenicol	4	IE
Cefepime	2	I	Colistin	≤ 0.5	S
Aztreonam	16	R	Trimethoprim/ Sulfamethoxazole	≤ 20	S

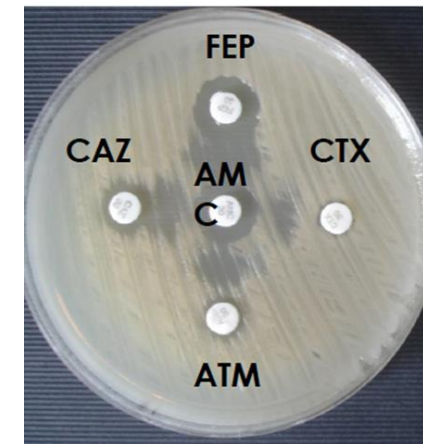
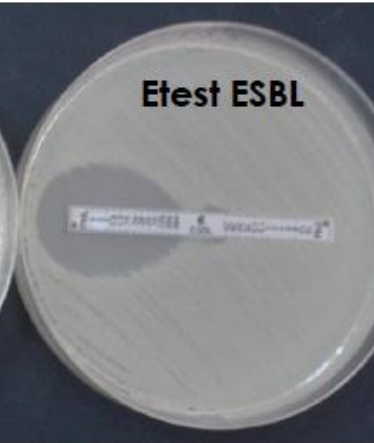
DDST



CDT



Etest ESBL

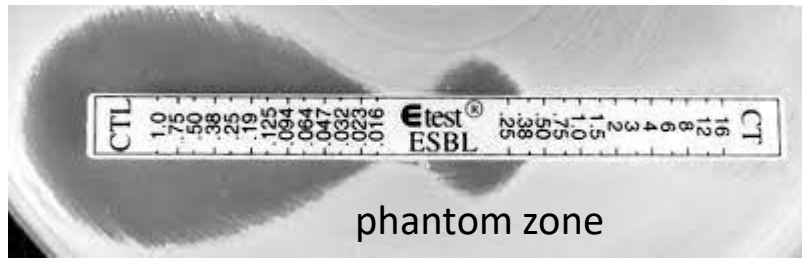
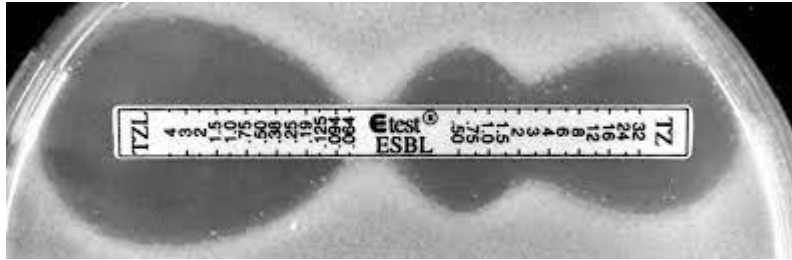


≥ 5 mm increase
in inhibition zone

MIC ratio ≥ 8
or if a
phantom zone
or deformed
ellipse is
present

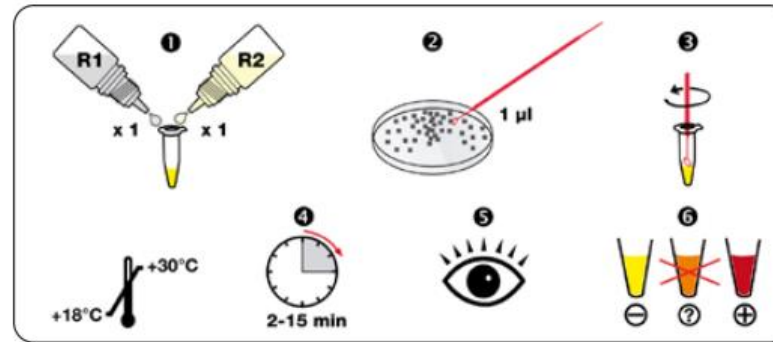
Φαινοτυπική ανίχνευση ESBL

Etest ESBL



Χρωματομετρικές μέθοδοι

β LACTA™ test

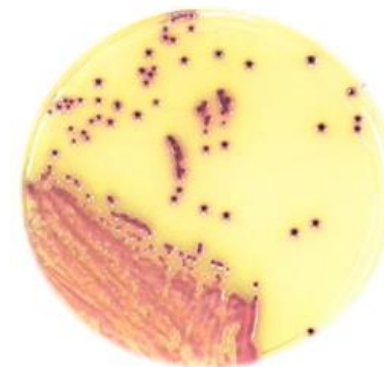


directly with isolated colonies or with bacterial pellets from positive blood cultures or urines

Se: 98%
Sp: 100%

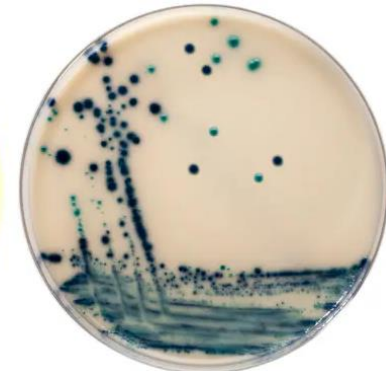
Χρωμογόνα υλικά

Liofilchem®
Chromatic ESBL



Escherichia coli DSM 22311

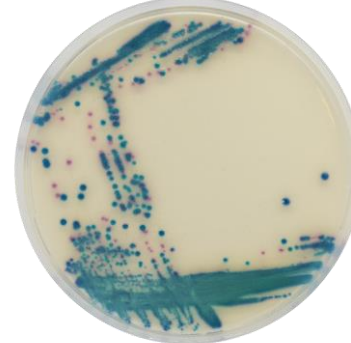
Brilliance™
ESBL Agar



CHROMID® ESBL



CHROMagar™
ESBL



Vitek reports

Organism Quantity:

Selected Organism: Klebsiella pneumoniae

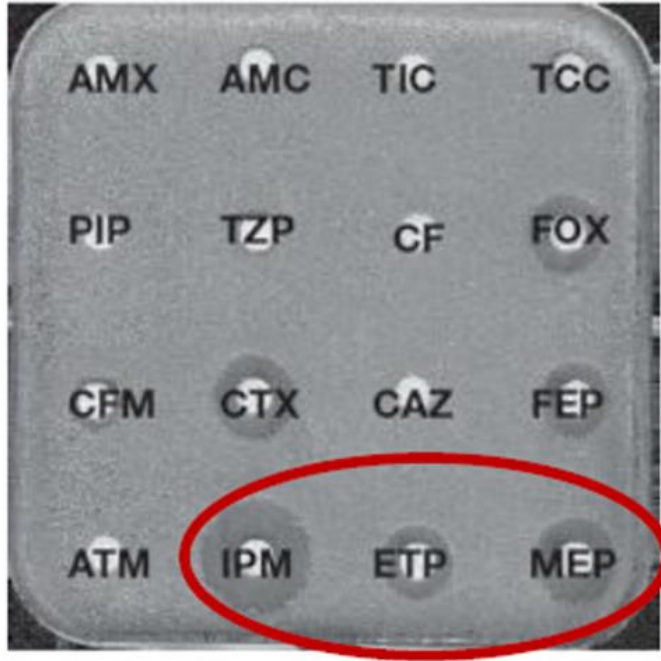
Antimicrobial	MIC	Interpretation	Antimicrobial	MIC	Interpretation
ESBL	NEG	-	Ertapenem	≥ 8	R
Temocillin			Imipenem	≥ 16	R
Urine	≥ 32	R	Meropenem		
Other	≥ 32		Meningitis	≥ 16	R
Ampicillin	≥ 32	R	Other	≥ 16	R
Amoxicillin/Clavulanic Acid	≥ 32	R	Amikacin	16	R
Piperacillin/Tazobactam	≥ 128	R	Gentamicin	≥ 16	R
Cefuroxime	≥ 64	R	Tobramycin	≥ 16	R
Cefuroxime Axetil	≥ 64	R	Ciprofloxacin		
Cefoxitin	≥ 64	IE	Meningitis	≥ 4	
Cefditoren			Other	≥ 4	R
Cefixime	≥ 4	R	Levofloxacin	≥ 8	R
Cefotaxime			Moxifloxacin	≥ 8	R
Meningitis	≥ 64	R	Minocycline	(-)	(-)
Other	≥ 64	R	Tetracycline	(-)	(-)
Ceftazidime	≥ 64	R	Tigecycline	2	R
Ceftriaxone			Fosfomycin		
Meningitis	≥ 64	R	Oral	128	R
Other	≥ 64	R	Other	128	R
Ceftazidime/Avibactam	4	(S)	Nitrofurantoin		
Ceftolozane/Tazobactam	≥ 32	R	Chloramphenicol	≥ 64	IE
Cefepime	≥ 32	R	Colistin	≥ 16	(R)
Aztreonam	≥ 64	R	Trimethoprim/ Sulfamethoxazole	≥ 320	R

Organism Quantity:

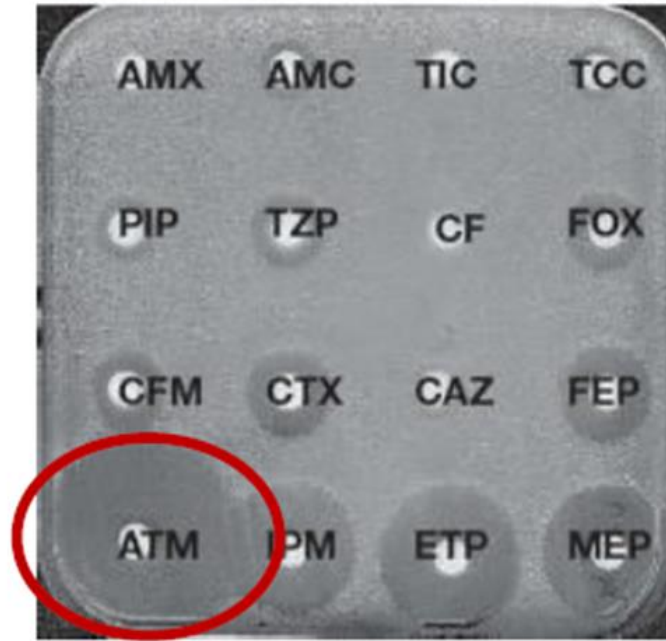
Selected Organism: Klebsiella pneumoniae

Antimicrobial	MIC	Interpretation	Antimicrobial	MIC	Interpretation
ESBL	NEG	-	Ertapenem	≥ 8	R
Temocillin			Imipenem	≥ 16	R
Urine	≥ 32	R	Meropenem		
Other	≥ 32		Meningitis	≥ 16	R
Ampicillin	≥ 32	R	Other	≥ 16	R
Amoxicillin/Clavulanic Acid	≥ 32	R	Amikacin	4	(S)
Piperacillin/Tazobactam	≥ 128	R	Gentamicin	≥ 16	R
Cefuroxime	≥ 64	R	Tobramycin	8	R
Cefuroxime Axetil	≥ 64	R	Ciprofloxacin		
Cefoxitin	≥ 64	IE	Meningitis	≥ 4	
Cefditoren			Other	≥ 4	R
Cefixime	≥ 4	R	Levofloxacin	≥ 8	R
Cefotaxime			Moxifloxacin	≥ 8	R
Meningitis	≥ 64	R	Minocycline	(-)	(-)
Other	≥ 64	R	Tetracycline	(-)	(-)
Ceftazidime	≥ 64	R	Tigecycline	1	R
Ceftriaxone			Fosfomycin		
Meningitis	≥ 64	R	Oral	≤ 16	
Other	≥ 64	R	Other	≤ 16	(S)
Ceftazidime/Avibactam	≥ 16	R	Nitrofurantoin		
Ceftolozane/Tazobactam	≥ 32	R	Chloramphenicol	16	IE
Cefepime	≥ 32	R	Colistin	≤ 0.5	(S)
Aztreonam	≥ 64	R	Trimethoprim/ Sulfamethoxazole	≤ 20	(S)

Ανίχνευση παραγωγής καρβαπενεμασών στα Εντεροβακτηριακά: πληροφορίες από το αντιβιογράμμα



KPC / ESBL +/- [Caz-avi S]
MBL / ESBL + [Caz-avi R]
OXA-48 / ESBL +
Co-production of ...



MBL
ESBL negative



OXA-48
ESBL negative

Ανίχνευση μηχανισμών αντοχής: CRE

Clinical breakpoints and **screening cut-off values** for carbapenemase-producing Enterobacteriaceae (according to EUCAST methodology)

Carbapenem	MIC (mg/L)		Disk diffusion zone diameter (mm) with 10 µg disks	
	S/I breakpoint	Screening cut-off	S/I breakpoint	Screening cut-off
Meropenem ¹	≤2	>0.125	≥22	<28 ²
Ertapenem ³	≤0.5	>0.125	≥25	<25

¹Best balance of sensitivity and specificity

²Isolates with **25-27 mm only need to be investigated for carbapenemase-production if they are resistant to piperacillin-tazobactam and/or temocillin** (temocillin contributes more to the specificity).

Investigation for carbapenemases is always warranted if zone diameter of meropenem is <25 mm.

³High sensitivity but low specificity. Can be used as an alternative screening agent, but isolates with ESBL and AmpC may be resistant without having carbapenemases.

Phenotypic methods

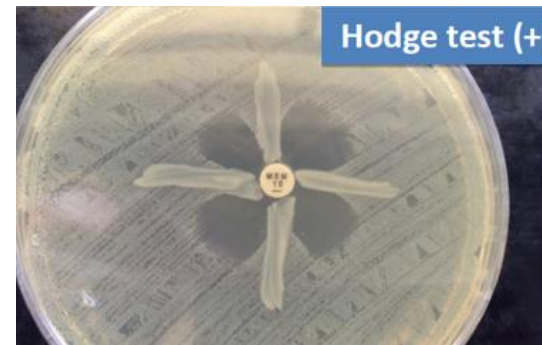
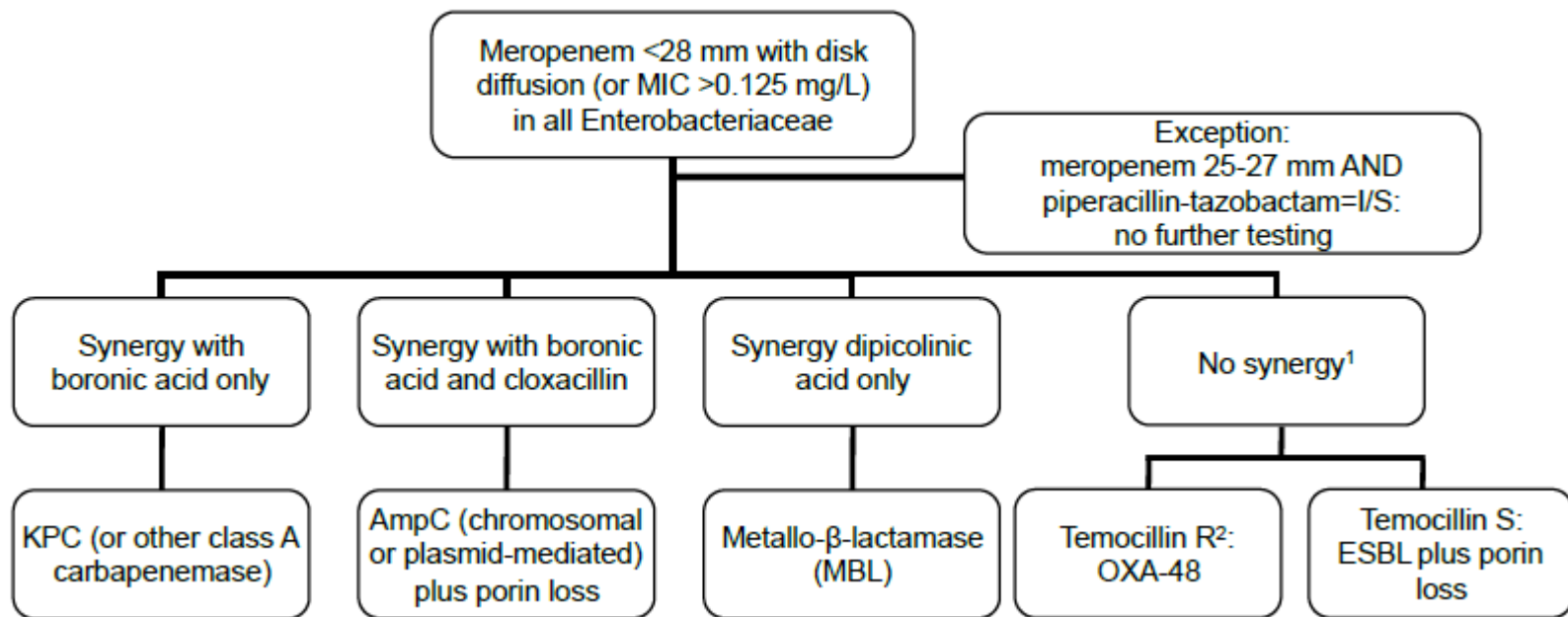
- combination disk test methods
- colorimetric assays based on hydrolysis of carbapenems



- other methods detecting carbapenem hydrolysis
- lateral flow assays

Ανίχνευση μηχανισμών αντοχής: CRE

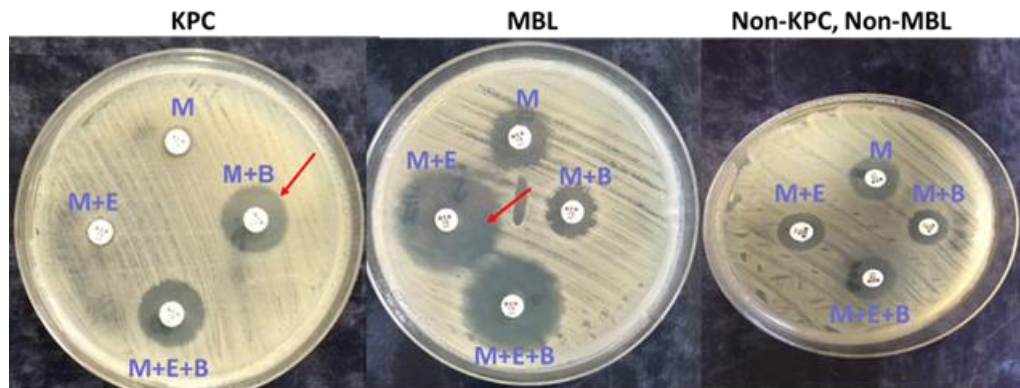
Φαινοτυπικός αλγόριθμος για ανίχνευση παραγωγής καρβαπενεμασών στα Εντεροβακτηριακά



Use of the modified cloverleaf (Hodge) test is **not currently recommended** as results are difficult to interpret, the specificity is poor, and in some cases the sensitivity is also suboptimal

¹Combination of several carbapenemases can also contribute to no synergy – e.g. MBL and KPC in combination. Molecular testing is usually necessary in such cases

²High-level temocillin resistance (>128 mg/L, zone diameter <11 mm) is a phenotypic marker of OXA-48



Journal of Antimicrobial Chemotherapy (2008) **62**, 1257–1260
doi:10.1093/jac/dkn364
Advance Access publication 4 September 2008

JAC

First occurrence of KPC-2-possessing *Klebsiella pneumoniae* in a Greek hospital and recommendation for detection with boronic acid disc tests

Athanassios Tsakris^{1*}, Ioulia Kristo², Aggeliki Poulou³, Fani Markou³,
Alexandros Ikonomidis² and Spyros Pournaras²

JOURNAL OF CLINICAL MICROBIOLOGY, Feb. 2009, p. 362–367
0095-1137/09/\$08.00+0 doi:10.1128/JCM.01922-08
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Vol. 47, No. 2

Evaluation of Boronic Acid Disk Tests for Differentiating KPC-Possessing *Klebsiella pneumoniae* Isolates in the Clinical Laboratory[▽]

Athanassios Tsakris,^{1*} Ioulia Kristo,² Aggeliki Poulou,³ Katerina Themeli-Digalaki,⁴
Alexandros Ikonomidis,² Dimitra Petropoulou,⁵ Spyros Pournaras,² and Danai Sofianou⁶

JOURNAL OF CLINICAL MICROBIOLOGY, Nov. 2009, p. 3420–3426
0095-1137/09/\$12.00 doi:10.1128/JCM.01314-09
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Vol. 47, No. 11

Use of Boronic Acid Disk Tests To Detect Extended-Spectrum β -Lactamases in Clinical Isolates of KPC Carbapenemase-Possessing *Enterobacteriaceae*[▽]

Athanassios Tsakris,^{1*} Aggeliki Poulou,² Katerina Themeli-Digalaki,³ Evangelia Voulgari,¹
Theodore Pittaras,¹ Danai Sofianou,⁴ Spyros Pournaras,⁵ and Dimitra Petropoulou⁶

Journal of
Antimicrobial
Chemotherapy

J Antimicrob Chemother 2010; **65**: 1319–1321
doi:10.1093/jac/dkq124 Advance Access publication 15 April 2010

Inhibitor-based methods for the detection of KPC carbapenemase-producing *Enterobacteriaceae* in clinical practice by using boronic acid compounds

Spyros Pournaras¹, Aggeliki Poulou² and Athanassios Tsakris^{3*}

JOURNAL OF CLINICAL MICROBIOLOGY, Aug. 2011, p. 2804–2809
0095-1137/11/\$12.00 doi:10.1128/JCM.00666-11
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2011;49(8):2804-9.

Vol. 49, No. 8

Comparative Evaluation of Combined-Disk Tests Using Different Boronic Acid Compounds for Detection of *Klebsiella pneumoniae* Carbapenemase-Producing *Enterobacteriaceae* Clinical Isolates[▽]

Athanassios Tsakris,^{1*} Katerina Themeli-Digalaki,² Aggeliki Poulou,^{1,3} Georgia Vrioni,¹
Evangelia Voulgari,¹ Vasiliki Koumaki,¹ Antonella Agodi,⁴
Spyros Pournaras,⁵ and Danai Sofianou⁶



2014;52(5):1483-9.

Modified CLSI Extended-Spectrum β -Lactamase (ESBL) Confirmatory Test for Phenotypic Detection of ESBLs among *Enterobacteriaceae* Producing Various β -Lactamases

Aggeliki Poulou,^{1,2} Evgenia Grivakou,² Georgia Vrioni,² Vassiliki Koumaki,² Theodoros Pittaras,² Spyros Pournaras,²
Athanassios Tsakris²

Department of Microbiology, Medical School, University of Athens, Athens, Greece^a; Department of Microbiology, General Hospital of Serres, Serres, Greece^b



2013;51(9):2986-90.

A Combined Disk Test for Direct Differentiation of Carbapenemase-Producing *Enterobacteriaceae* in Surveillance Rectal Swabs

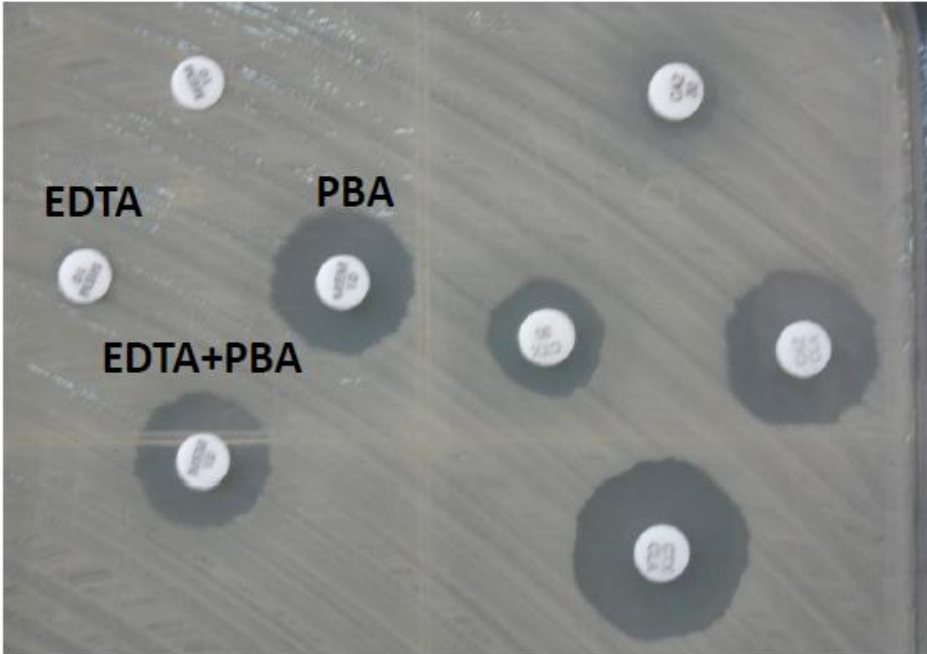
Spyros Pournaras,² Olympia Zarkotou,^{2,3} Aggeliki Poulou,^{2,4} Ioulia Kristo,⁴ Georgia Vrioni,² Katerina Themeli-Digalaki,²
Athanassios Tsakris²

Department of Microbiology, Medical School, University of Athens, Athens, Greece^a; Department of Microbiology, Tzaneio General Hospital, Piraeus, Greece^b; Department of Microbiology, Serres General Hospital, Serres, Greece^c; Department of Microbiology, Faculty of Medicine, University of Thessaly, Larissa, Greece^d

Φαινοτυπική ανίχνευση καρβαπενεμασών στα Εντεροβακτηριακά

CDT

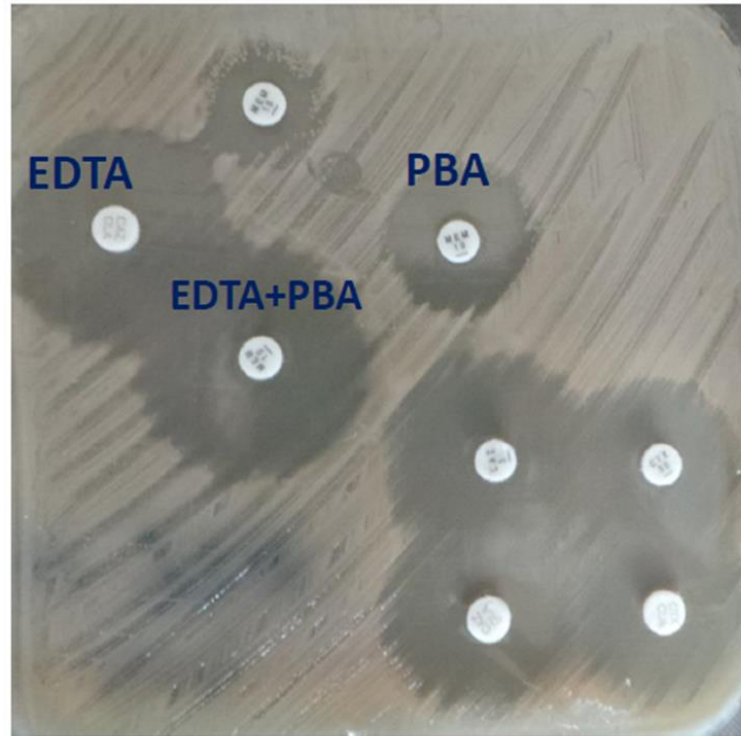
Modified
ESBL test



KPC

CDT

Modified
ESBL test



MBL



KPC+MBL

Φαινοτυπική ανίχνευση καρβαπενεμασών στα Εντεροβακτηριακά: MBL

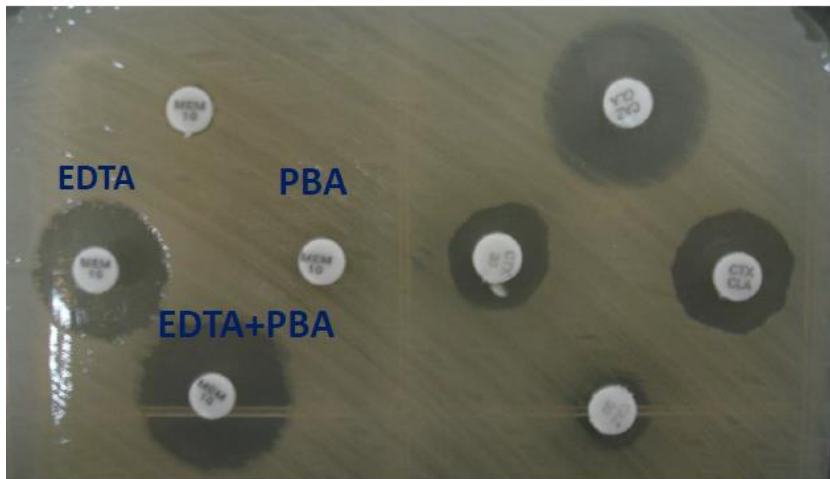
CDT

DDST

Etest MBL

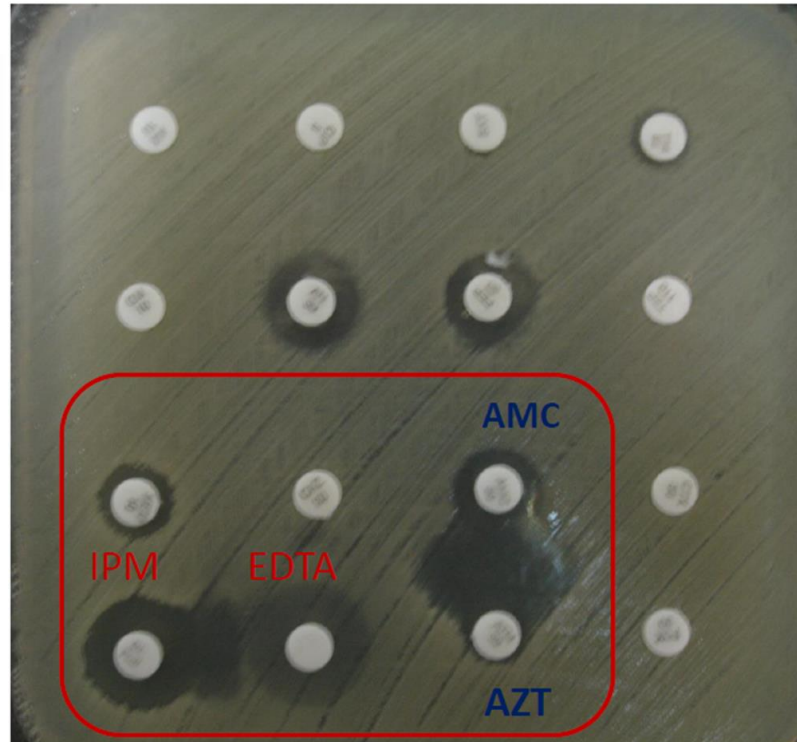
Αλγόριθμος
καρβαπεμενασών

Modified CLSI
ESBL test



MBL positive (NDM-1)

ESBL positive



MBL positive

ESBL positive

Different growth-inhibition patterns:

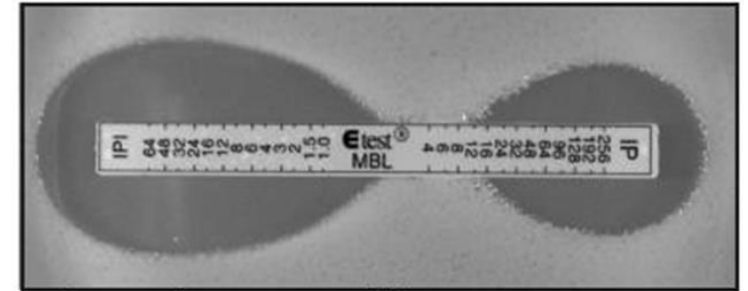


Figure 2. Clear cut MBL positive: $IP/IPIC = 16 / <1 = >16$



Figure 3. Phantom zone between IP/IPI is indicative of MBL

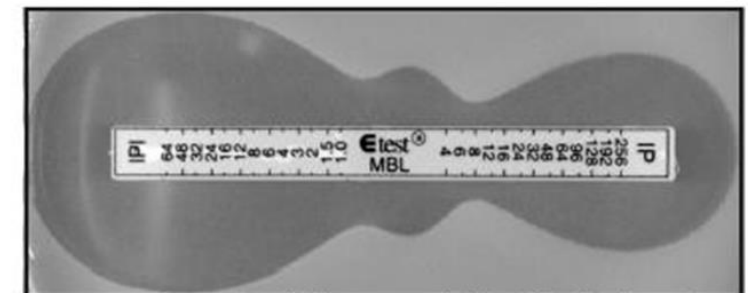
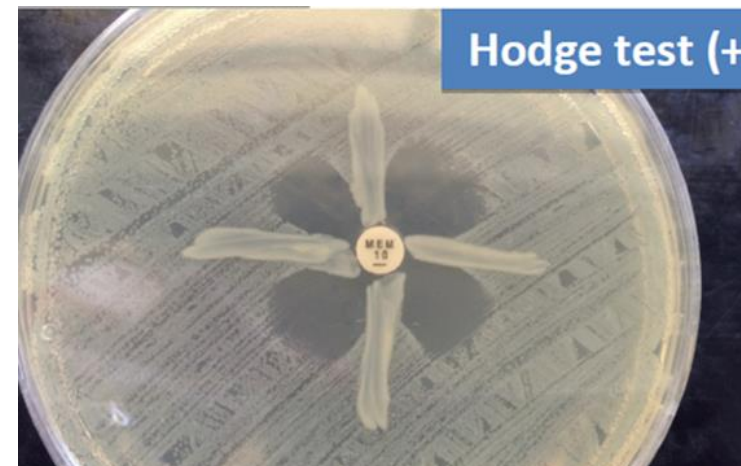
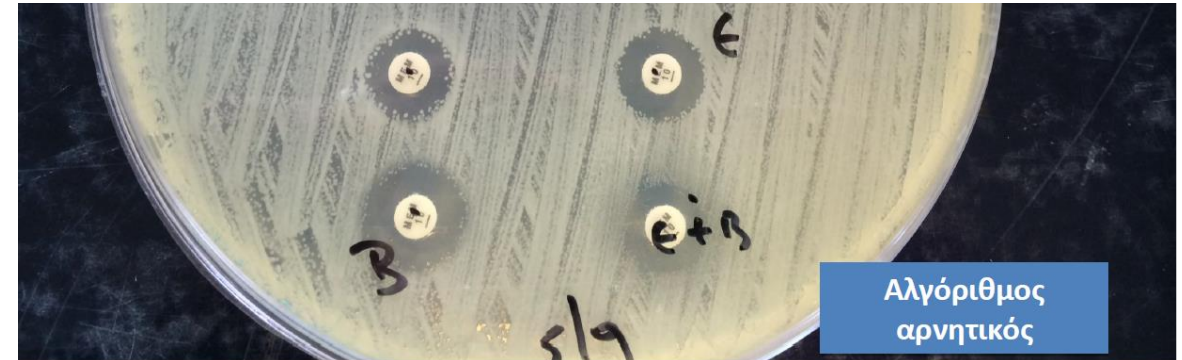
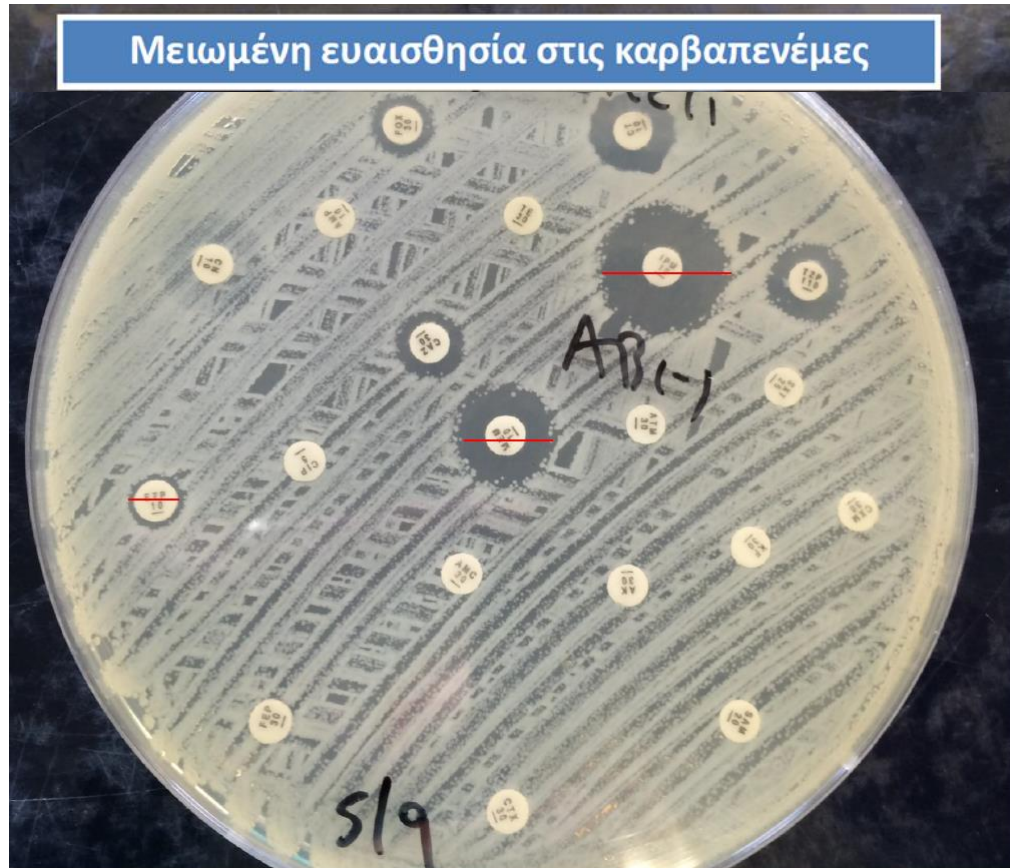


Figure 4. Deformation of the IP or IPI ellipse is indicative of MBL

Φαινοτυπική ανίχνευση καρβαπενεμασών στα Εντεροβακτηριακά



VIM (-)	KPC (-)	OXA-48 (+)	CTX-M (+)	TEM (+)	NDM (-)	SHV (+)
---------	---------	-------------------	------------------	----------------	---------	----------------

E. coli

Antimicrobial	MIC	Interpretation	Antimicrobial	MIC	Interpretation
ESBL	POS	+	Ertapenem	0.25	S
Temocillin			Imipenem	≤ 0.25	S
Urine	≥ 32	R	Meropenem		
Other	≥ 32		Meningitis	≤ 0.25	S
Ampicillin	≥ 32	R	Other	≤ 0.25	S
Amoxicillin/Clavulanic Acid	≥ 32	R	Amikacin	2	S
Piperacillin/Tazobactam	≥ 128	R	Gentamicin	≤ 1	S
Cefuroxime	≥ 64	R	Tobramycin	≤ 1	S
Cefuroxime Axetil	≥ 64	R	Ciprofloxacin		
Cefoxitin	≤ 4	IE	Meningitis	0.5	
Cefditoren			Other	0.5	I
Cefixime	≥ 4	R	Levofloxacin	1	I
Cefotaxime			Moxifloxacin	0.5	R
Meningitis	≥ 64	R	Minocycline	(-)	(-)
Other	≥ 64	R	Tetracycline	(-)	(-)
Ceftazidime	32	R	Tigecycline	≤ 0.5	S
Ceftriaxone			Fosfomycin		
Meningitis	≥ 64	R	Oral	≤ 16	
Other	≥ 64	R	Other	≤ 16	S
Ceftazidime/Avibactam	≤ 0.12	S	Nitrofurantoin		
Ceftolozane/Tazobactam	8	R	Chloramphenicol	16	IE
Cefepime	≥ 32	R	Colistin	≤ 0.5	S
Aztreonam	≥ 64	R	Trimethoprim/ Sulfamethoxazole	≥ 320	R

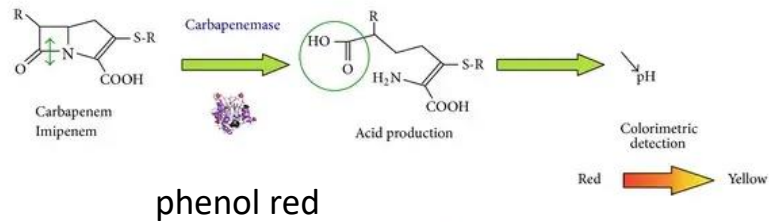
(-)= Susceptibility testing not recommended; species is a poor target for therapy IE= Insufficient Evidence that species is good target for therapy; MIC may be reported without interpretation



Φαινοτυπική ανίχνευση καρβαπενεμασών στα Εντεροβακτηριακά

Χρωματομετρικές μέθοδοι (colorimetric tests)

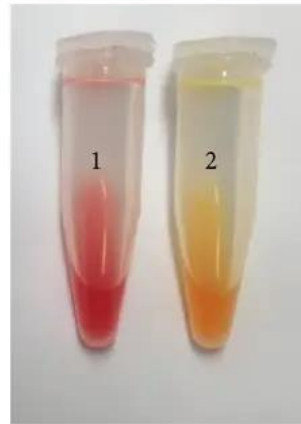
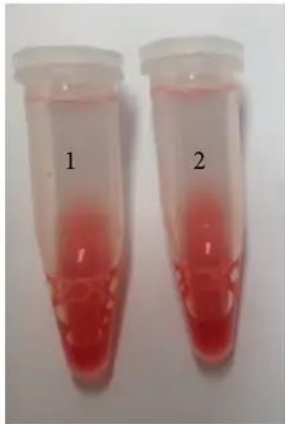
Carba NP Test



(a)

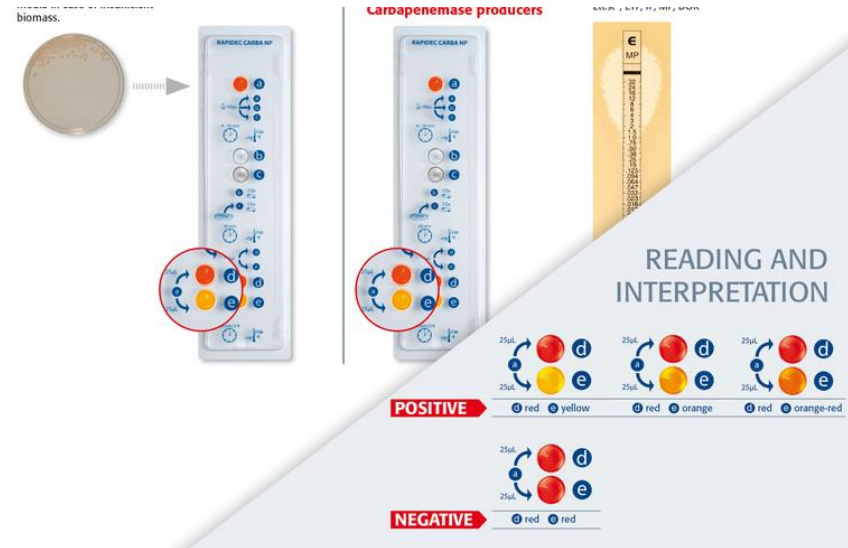
No carbapenemase

Carbapenemase production



- (1) Revealing solution
(internal negative control)
(2) Revealing solution + imipenem

(b)



Sensitivity: 73–100% for most carbapenemases
Performed poorly for OXA-48 enzyme

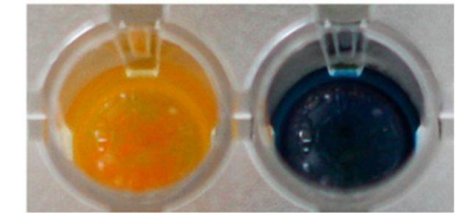
Nordmann P, Poirel L, Dortet L. Emerg Infect Dis. 2012;18:1503-7.
Dortet L, Poirel L, Nordmann P. Antimicrob Agents Chemother. 2012;56:6437-40.
Dortet L et al. J Antimicrob Chemother. 2015 ; 70):3014-22.
Kabir MH et al. J Antimicrob Chemother. 2016 ;71:1213-6.
Noël A et al. J Clin Microbiol. 2017;55(2):510-518
Pasteran F et al. J Clin Microbiol. 2015 ;53(6):1996-8.

Blue-Carba Test

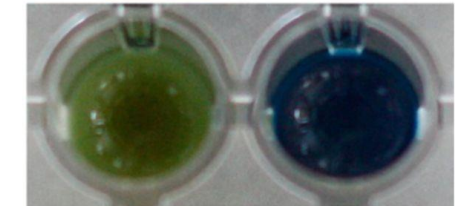
indicator bromothymol blue

Test Solution Negative Control

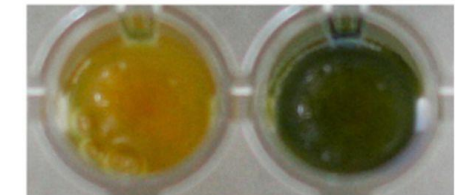
A



B



C

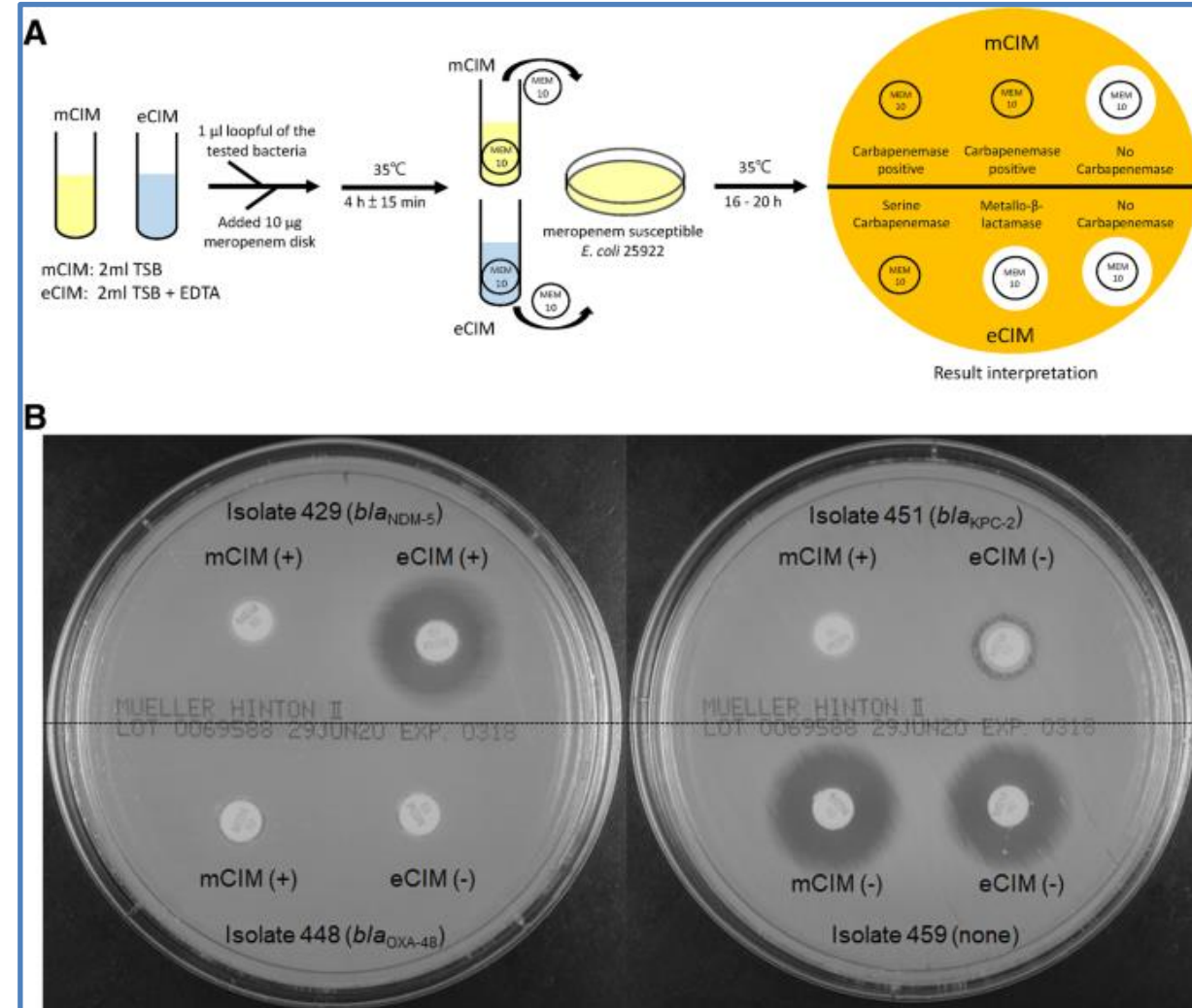
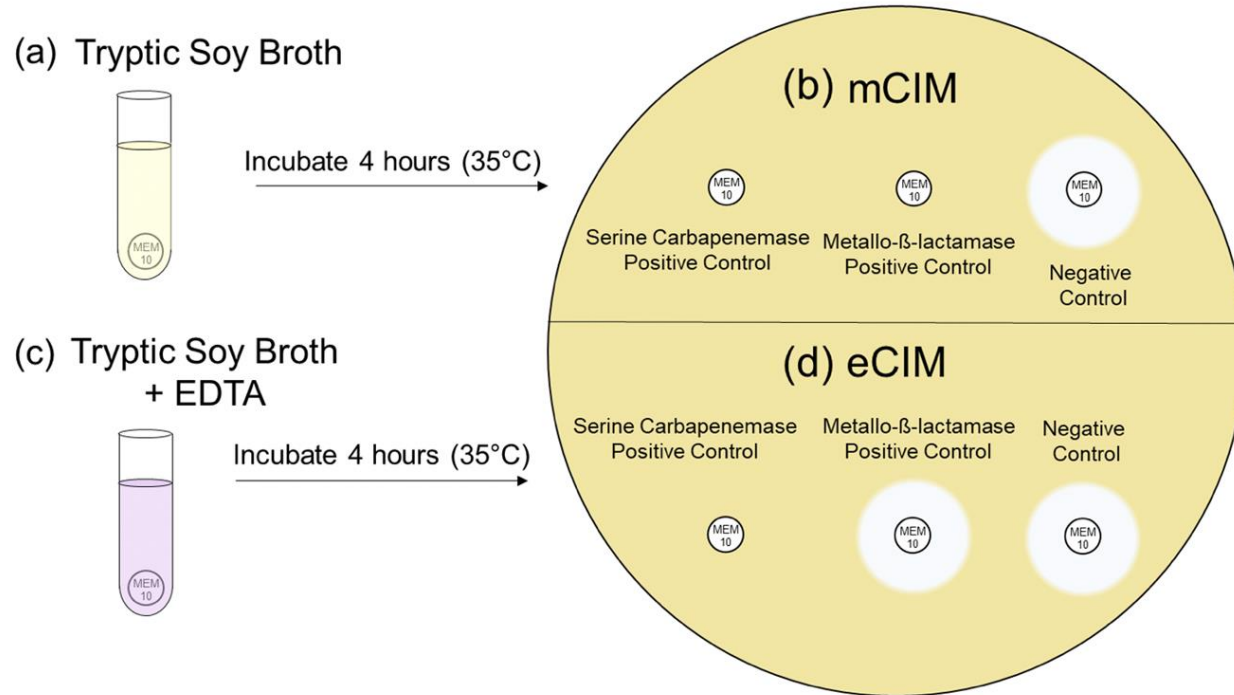


D



Screening test: Carbapenemase Inactivation Methods (mCIM/eCIM)

The Carbapenemase Inactivation Methods (mCIM/eCIM) are Currently Recommended by CLSI



Tamma PD, Simner PJ. J Clin Microbiol. 2018;56(11):e01140–18.

<https://asm.org/Articles/2019/May/Opening-the-Black-Box-Phenotypic-Carbapenemase-Det>

van der Zwaluw K et al. PLoS One. 2015;10(3):e0123690

Yamada K et al. J Microbiol Methods. 2016;128:48-51.

Tijet N, Patel SN, Melano RG. J Antimicrob Chemother. 2016 ;71(1):274-6.

Vitek reports

Bionumber: 0201010103500210

Organism Quantity:

Selected Organism: Acinetobacter baumannii complex

Antimicrobial	MIC	Interpretation	Antimicrobial	MIC	Interpretation
ESBL			Imipenem	≥ 16	R
Temocillin			Meropenem		
Ampicillin	(-)	(-)	Meningitis	≥ 16	R
Amoxicillin/Clavulanic Acid			Other	≥ 16	R
Piperacillin/Tazobactam	≥ 128	IE	Amikacin	≥ 64	R
Cefuroxime	(-)	(-)	Gentamicin	≥ 16	R
Cefuroxime Axetil	(-)	(-)	Tobramycin	≥ 16	R
Cefoxitin	(-)	(-)	Ciprofloxacin	≥ 4	R
Cefditoren			Levofloxacin	4	R
Cefixime	(-)	(-)	Moxifloxacin	(-)	(-)
Cefotaxime	(-)	(-)	Minocycline	2	IE
Ceftazidime	(-)	(-)	Tetracycline	(-)	(-)
Ceftriaxone	(-)	(-)	Tigecycline	4	IE
Ceftazidime/Avibactam			Fosfomycin	(-)	(-)
Ceftolozane/Tazobactam			Nitrofurantoin	(-)	(-)
Cefepime	(-)	(-)	Chloramphenicol	(-)	(-)
Aztreonam	(-)	(-)	Colistin	≤ 0.5	S
Ertapenem			Trimethoprim/ Sulfamethoxazole	≥ 320	R

4 Result reading

Turbid = growth = positive Clear = absence of growth = negative

The growth control must be positive

MIC = FIRST CLEAR WELL

Easy reading of the MIC with the UMIC Box

Report on the provided results sheet

Organism Quantity:

Selected Organism: Pseudomonas aeruginosa

Comments:	Colistin S results / Gram negative bacilli: perform an alternative method of testing prior to reporting results.

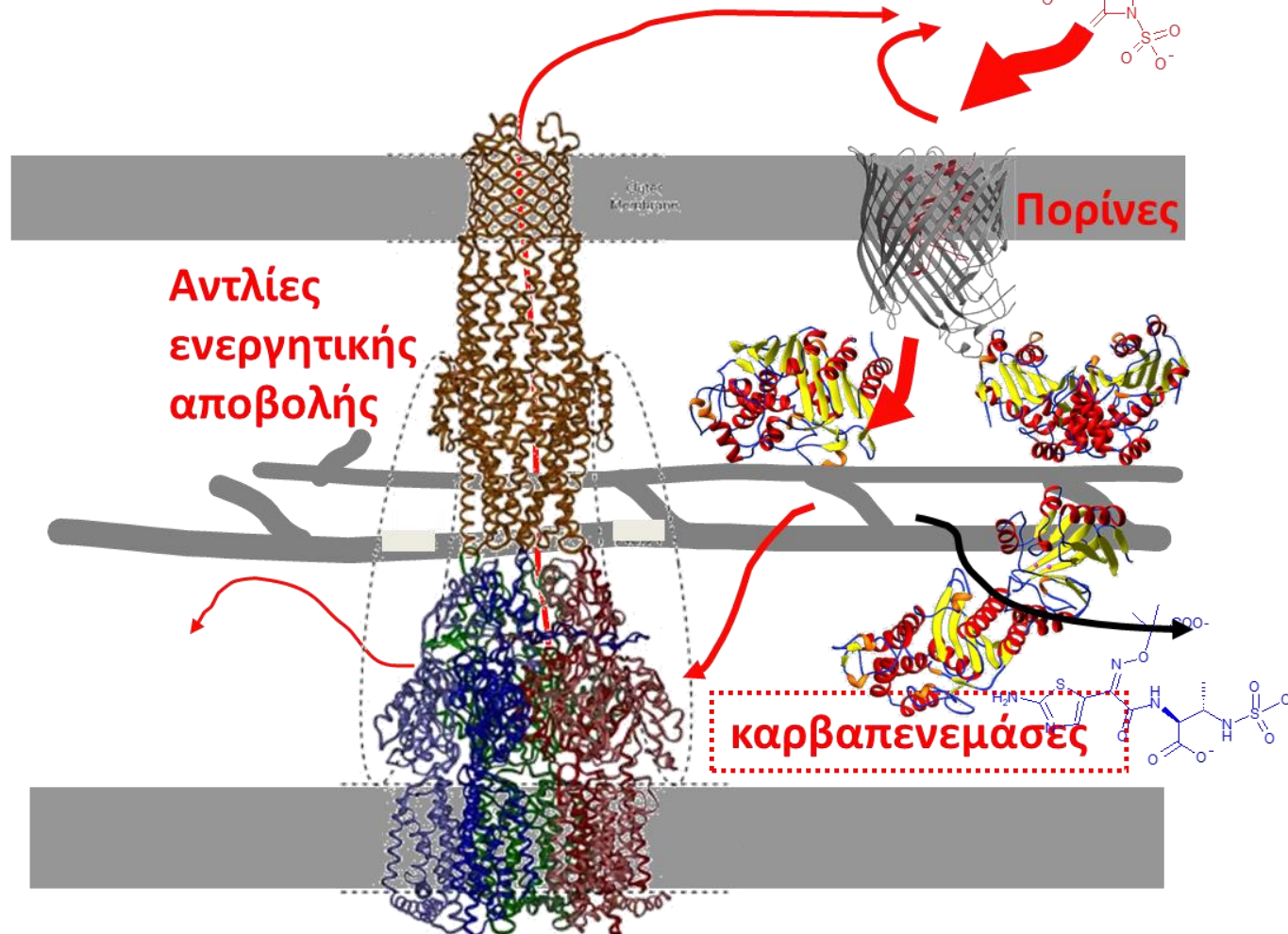
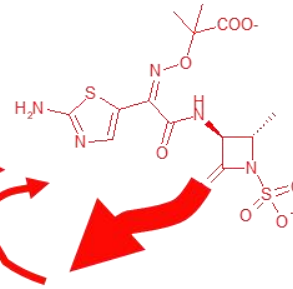
Identification Information	
Organism Origin	Technologist
Selected Organism	Pseudomonas aeruginosa
	Entered: Sep 15, 2023 13:42 EEST By: LabTech
Analysis Messages:	

Susceptibility Information	Card: AST-N318	Lot Number: 7582293203	Expires: Mar 7, 2024 12:00 EEST
	Status: Final	Analysis Time: 9.27 hours	Completed: Sep 15, 2023 22:42 EEST

Antimicrobial	MIC	Interpretation	Antimicrobial	MIC	Interpretation
+Ampicillin		(-)	Meropenem		
Ampicillin/Sulbactam			Meningitis	≥ 16	R
Piperacillin/Tazobactam	≥ 128	R	Other	≥ 16	R
Cefoxitin			Amikacin	≥ 64	R
+Cefotaxime		(-)	Gentamicin	≥ 16	IE
Ceftazidime	≥ 64	R	Ciprofloxacin	≥ 4	R
Ceftriaxone	(-)	(-)	Levofloxacin	≥ 8	R
+Ceftolozane/Tazobactam			Tigecycline	(-)	(-)
Cefepime	≥ 64	R	Fosfomycin	(-)	(-)
Aztreonam	≥ 64	R	Colistin	2	S
+Ertapenem		(-)	Trimethoprim/ Sulfamethoxazole	(-)	(-)
Imipenem	≥ 16	R			

P. aeruginosa και αντοχή στις καρβαπενέμες

Multidrug-resistant (MDR)
Pseudomonas aeruginosa



Τάξη	Ένζυμα
A	KPC GES
D	OXA (OXA-40, -50 -198)
B	VIM IMP NDM SPM GIM, AIM, FIM

P. aeruginosa και φαινοτυπική ανίχνευση MBL

Δεν υπάρχουν προτυποποιημένες δοκιμασίες



Οι δοκιμασίες στηρίζονται στη
χρήση χηλικών παραγόντων
EDTA, MPA, DPA

**All phenotypic combined disk
tests lacked either sensitivity
or specificity for the detection
of MBL in *P. aeruginosa***

μεθοδολογία
(DDST, CDT, Etest MBL)
το β-λακταμικό υπόστρωμα
το είδος του αναστολέα
ποσότητα του αναστολέα
χαρακτηριστικά των στελεχών

Συνήθως

- υψηλού επιπέδου αντοχή σε IMP και MEM
- υψηλού επιπέδου αντοχή στην CAZ
- ευαισθησία στην αζτρεονάμη και ουρεϊδοπενικιλίνες ποικίλλει
- συνυπάρχει αντοχή στις αμινογλυκοσίδες

FP: επίδραση του EDTA

στη διαπερατότητα της μεμβράνης

επίδραση του Zn

στην έκφραση της OprD

Συγκέντρωση ιόντων Zn στο MH

FN: χαμηλού επιπέδου αντοχή

CDT



EDTA 292 μg (10 μl EDTA 0.1 M)

$\Delta d > 4 \text{ mm}$

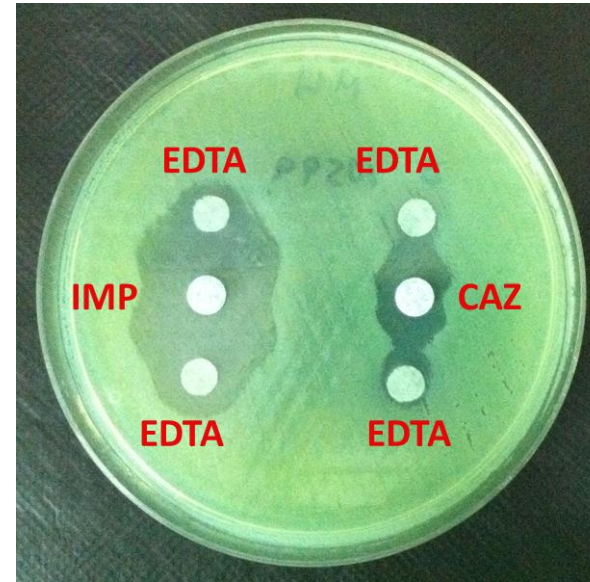
EDTA 750 μg , $\Delta d \geq 6 \text{ mm}$

100% SN και SP

DPA 835 μg , $\Delta d \geq 5 \text{ mm}$

SN=93,2% SP= 97,1%

DDST

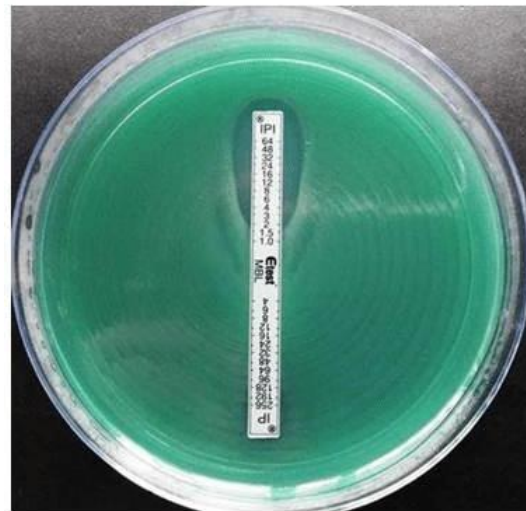
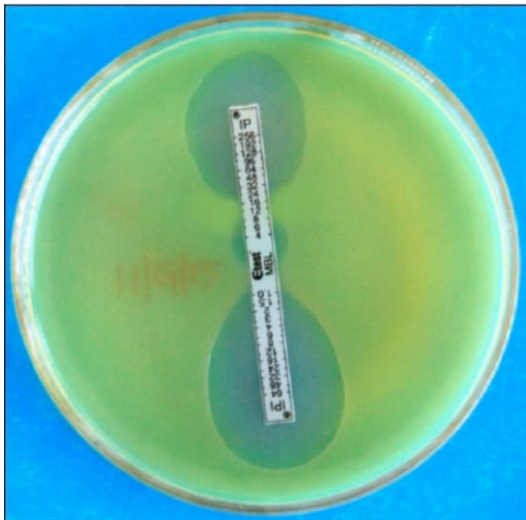


EDTA 292 μg

5 μl MPA 1.4 Mm

DPA 250 μg

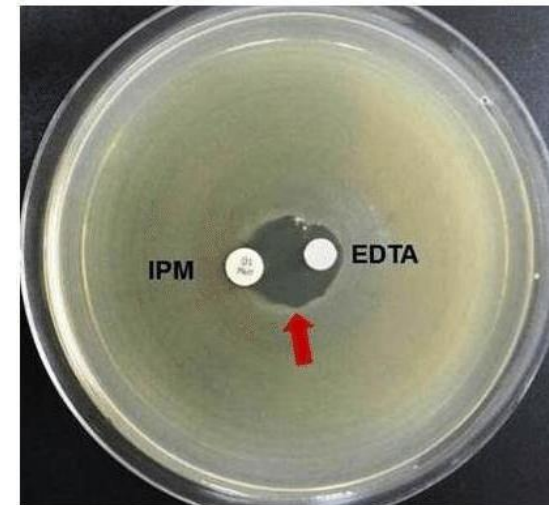
Etest MBL, bioMerieux



IP/ IPI ≥ 8

παρουσία ζώνης-φάντασμα

παραμόρφωση των ελλείψεων IP ή IPI



For *P. aeruginosa* the MBL Etest® as well as disk-based assays have been used for several decades, but are hampered **by poor specificity.**

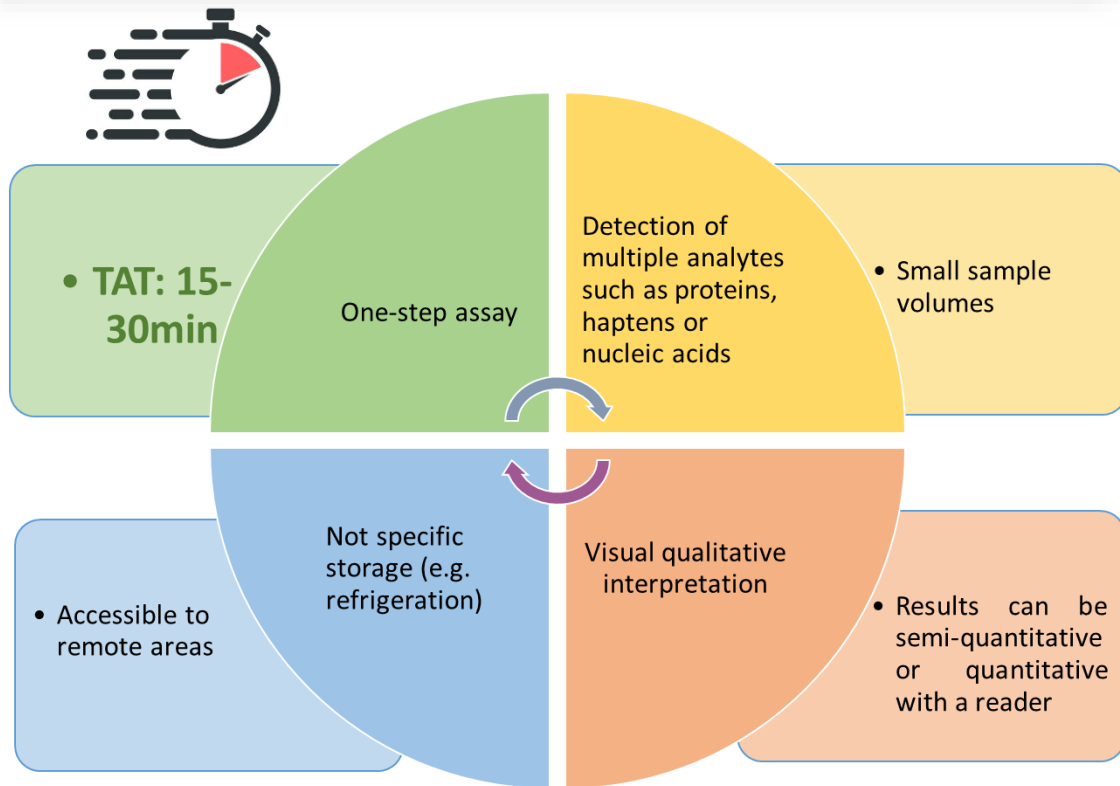
EUCAST guidelines for detection of resistance mechanisms and specific resistances of clinical and/or epidemiological importance



Review

The Revolution of Lateral Flow Assay in the Field of AMR Detection

Hervé Boutil¹, Christian Moguet¹, Lilas Pommès¹, Stéphanie Simon¹, Thierry Naas^{2,3,4} and Hervé Volland^{1,*}

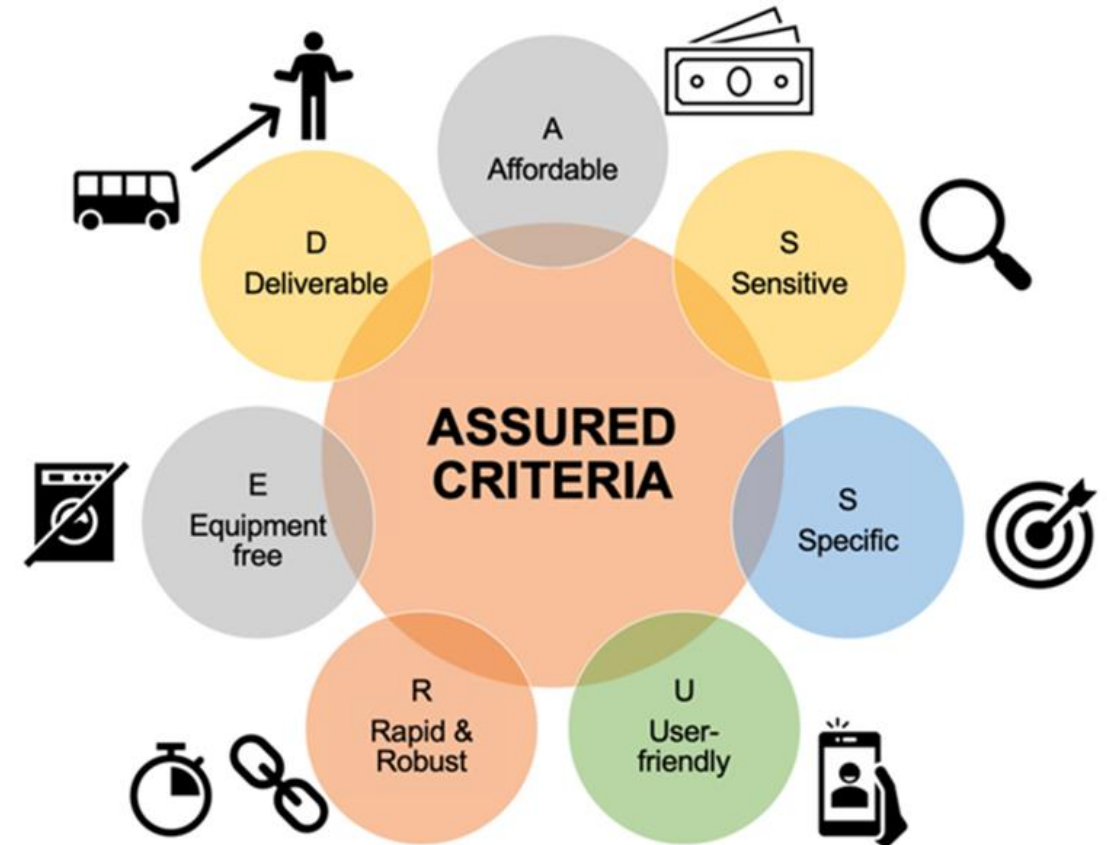


LFIA performance relies mostly on antibody affinity and specificity



World Health Organization

an ideal PoC test

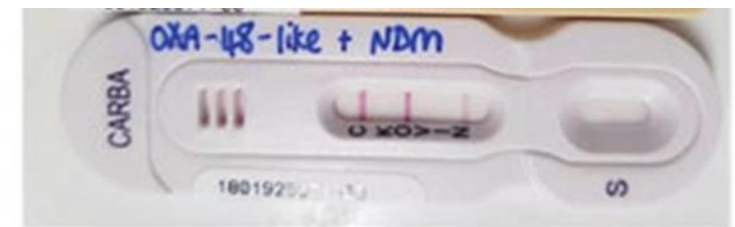
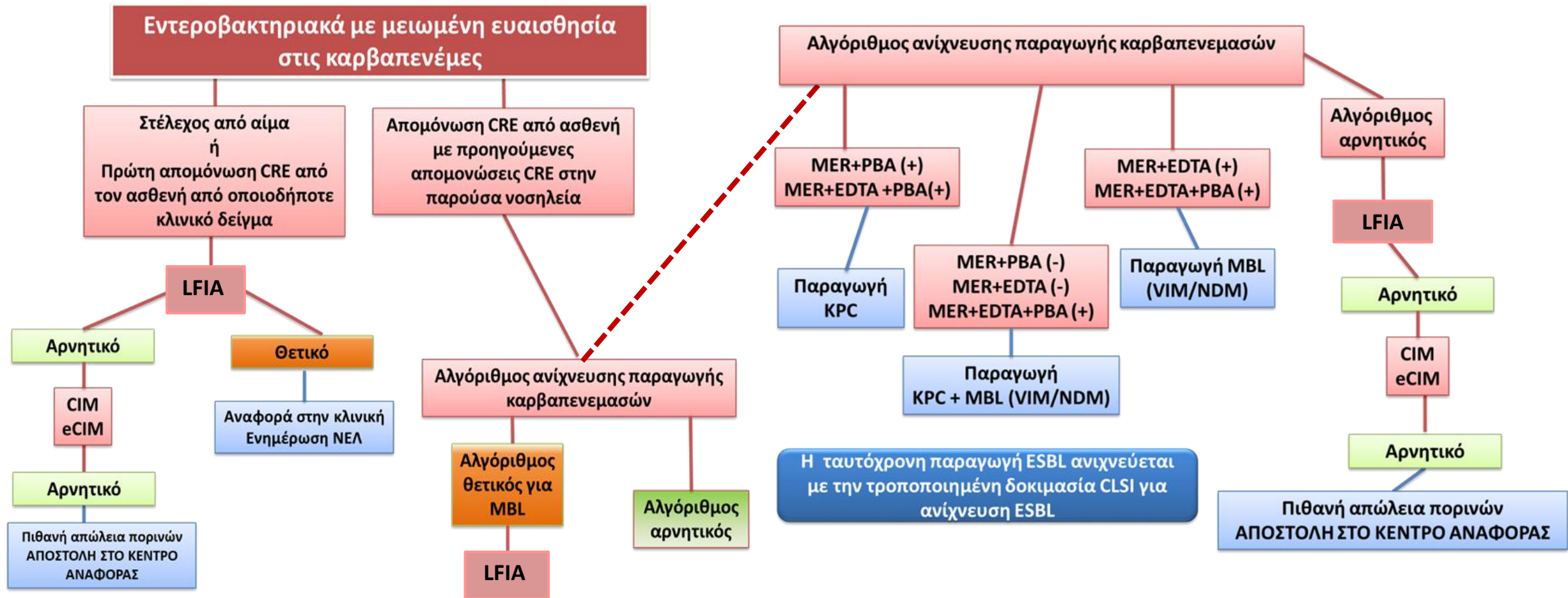


PBC

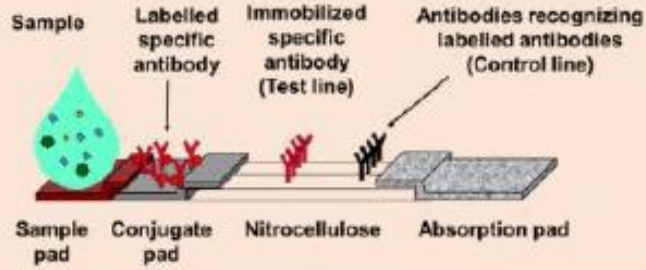
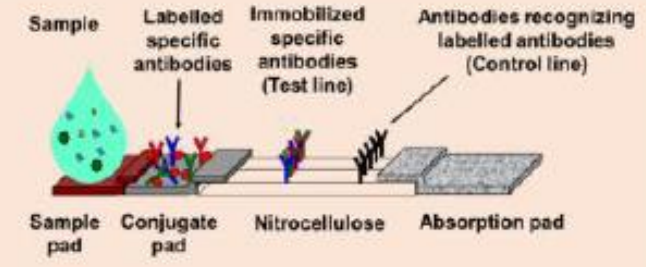
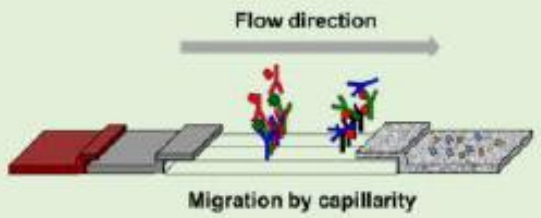
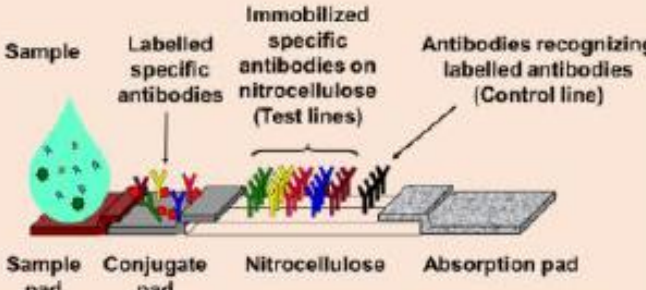
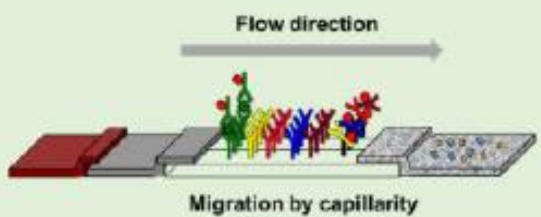
Se: 98%

Sp: 100%

Del Corpo O et al. Clin Microbiol Infect. 2023 Sep 16:S1198-743X(23)00425-1.



Lateral flow assay formats

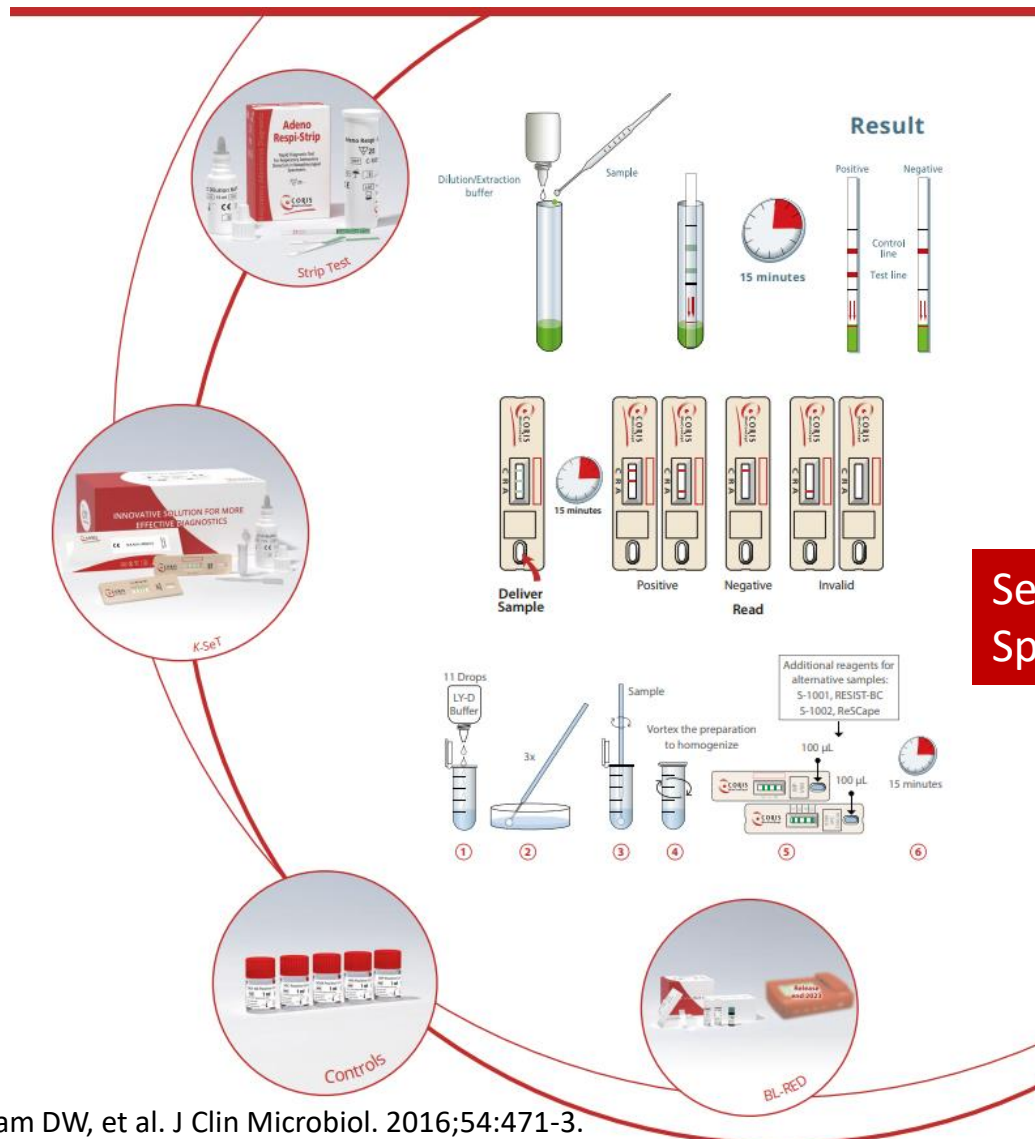
Monoplex format	Strip structure	Immunological detection	Results
	 Sample pad Conjugate pad Nitrocellulose Absorption pad	 Flow direction Migration by capillarity	✓ The control line appears: the test is correct ✓ The test line appears: positive test ✓ No test line: negative test ✓ No control line: invalid result 
Multiplex format (one test line)	 Sample pad Conjugate pad Nitrocellulose Absorption pad	 Flow direction Migration by capillarity	✓ The control line appears: the test is correct ✓ The test line appears: positive test for at least one target ✓ No test line: negative test ✓ No control line: invalid result 
Multiplex format (Five test lines)	 Sample pad Conjugate pad Nitrocellulose Absorption pad	 Flow direction Migration by capillarity	✓ The control line appears: the test is correct ✓ One or several test lines appear: positive test for the corresponding target ✓ No test line appears: negative test for any target ✓ No control line: invalid result 



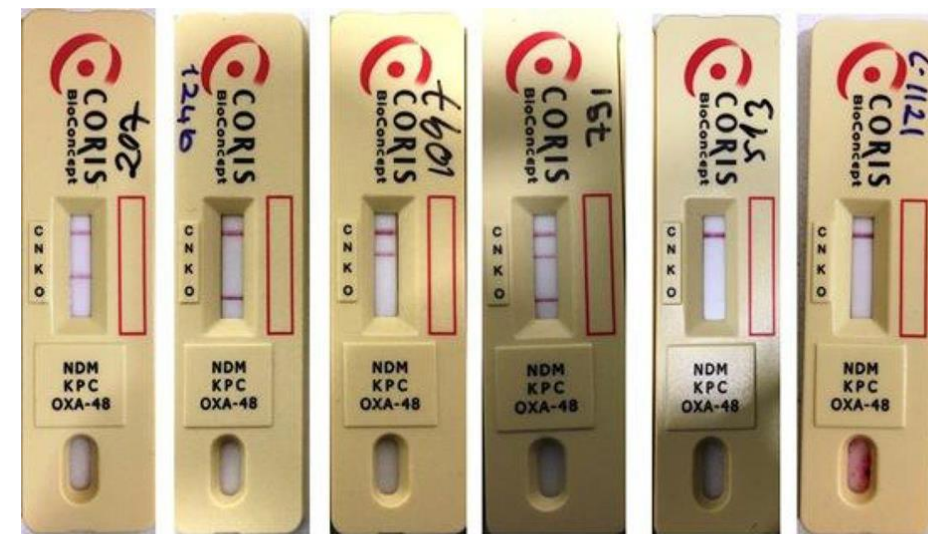
Targets	Technology	Product	Other identifications
Beta Lactamases	Electrochemistry	BL-RED 25	
OXA-48-like	Lateral flow Lateral flow Lateral flow Lateral flow Reagents Reagents	OXA-48 K-Set O.K.N.V.I. RESIST-5 RESIST-3 O.O.K. K-Set RESIST-3 O.K.N. K-Set RESIST-BC ReSCape	KPC, NDM, VIM, IMP OXA-163-like, KPC KPC, NDM Carbapenemases: KPC, NDM, VIM, IMP, OXA-40/58 Carbapenemases: KPC, NDM, VIM, IMP, OXA-40/58
KPC	Lateral flow Lateral flow Lateral flow Reagents Reagents	O.K.N.V.I. RESIST-5 RESIST-3 O.O.K. K-Set RESIST-3 O.K.N. K-Set RESIST-BC ReSCape	OXA-48-like, NDM, VIM, IMP OXA-48-like, OXA-163-like OXA-48-like, NDM Carbapenemases: OXA-48-like, NDM, VIM, IMP, OXA-40/58 Carbapenemases: OXA-48-like, NDM, VIM, IMP, OXA-40/58
OXA-163-like	Lateral flow	RESIST-3 O.O.K. K-Set	OXA-48-like, KPC
NDM	Lateral flow Lateral flow Reagents Reagents Lateral flow	O.K.N.V.I. RESIST-5 RESIST-3 O.K.N. K-Set RESIST-BC ReSCape RESIST ACINETO	OXA-48-like, KPC, VIM, IMP OXA-48-like, KPC Carbapenemases: OXA-48-like, KPC, VIM, IMP, OXA-40/58 Carbapenemases: OXA-48-like, KPC, VIM, IMP, OXA-40/58 OXA-23-like, OXA-40/58
OXA-23-like	Lateral flow Lateral flow	OXA-23 K-Set RESIST ACINETO	OXA-40/58, NDM
VIM	Lateral flow Reagents Reagents	O.K.N.V.I. RESIST-5 RESIST-BC ReSCape	OXA-48-like, KPC, NDM, IMP Carbapenemases: OXA-48-like, KPC, NDM, IMP, OXA-40/58 Carbapenemases: OXA-48-like, KPC, NDM, IMP, OXA-40/58
IMP	Lateral flow Lateral flow Reagents Reagents	O.K.N.V.I. RESIST-5 IMP K-Set RESIST-BC ReSCape	OXA-48-like, KPC, NDM, VIM Carbapenemases: OXA-48-like, KPC, NDM, VIM, OXA-40/58 Carbapenemases: OXA-48-like, KPC, NDM, VIM, OXA-40/58
Carbapenemases	Reagents Reagents	RESIST-BC ReSCape	Carbapenemases: OXA-48-like, KPC, NDM, VIM, IMP, OXA-40/58 Carbapenemases: OXA-48-like, KPC, NDM, VIM, IMP, OXA-40/58
OXA-40/58	Reagents Reagents Lateral flow	RESIST-BC ReSCape RESIST ACINETO	Carbapenemases: OXA-48-like, KPC, NDM, VIM, IMP Carbapenemases: OXA-48-like, KPC, NDM, VIM, IMP OXA-23-like, NDM
Group 1 (CTX-M-15,...)	Lateral flow	RESIST CTX-M	Group 9 (CTX-M-14,...)
Group 9 (CTX-M-14,...)	Lateral flow	RESIST CTX-M	Group 1 (CTX-M-15,...)

ACCESSORIES TO ACCELERATE RESIST RANGE RESULTS

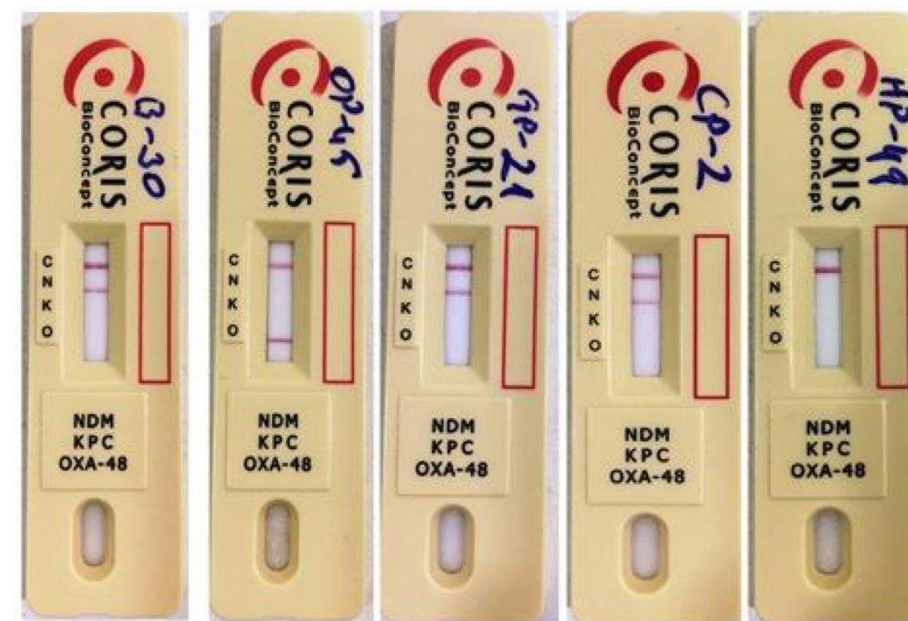
PRODUCT NAME	DESCRIPTION
RESIST-BC	Reagents kit for blood culture 1 RBCL solution, 1 MS buffer, 1 Washing buffer
ReSCape	Culture reagent kit on rectal swab 20 CProBE MEDIUM tubes, 2 Selective Mix vials, 2 Water vials



Se: 98 -100%
Sp: 100%



1 2 3 4 5 6



7 8 9 10 11

Wareham DW, et al. J Clin Microbiol. 2016;54:471-3.
 Pasteran F, et al. J Clin Microbiol. 2016;54:2832-6.
 Glupczynski Y et al. J Antimicrob Chemother. 2017;72(7):1955-1960.
 Greissl C et al. Eur J Clin Microbiol Infect Dis. 2019;38(2):331-335.
 Song W et al. Ann Lab Med. 2020;40(3):259-263.
 Sadek M et al. Diagn Microbiol Infect Dis. 2022;104(4):115761.
 Bouvier M et al. Diagn Microbiol Infect Dis. 2023;107(3):116043.

Ανίχνευση καρβαπενεμασών σε στελέχη *Acinetobacter*

N = 174 well-characterized carbapenemase and non-carbapenemase producing isolates belonging to various *Acinetobacter* species

149 *A. baumannii*, 11 *A. pittii*, four *A. radioresistens*, three *A. ursingii*, two *A. calcoaceticus*, two *A. nosocomialis*, one *A. bereziniae*, one *A. lwoffii* and one *A. junii*.



Negative test
A. baumannii



OXA-40/58 +
A. baumannii



OXA-23 +
A. baumannii



NDM +
A. baumannii



OXA-40/58 +
OXA-23 +
A. baumannii



OXA-40/58 +
NDM +
A. baumannii



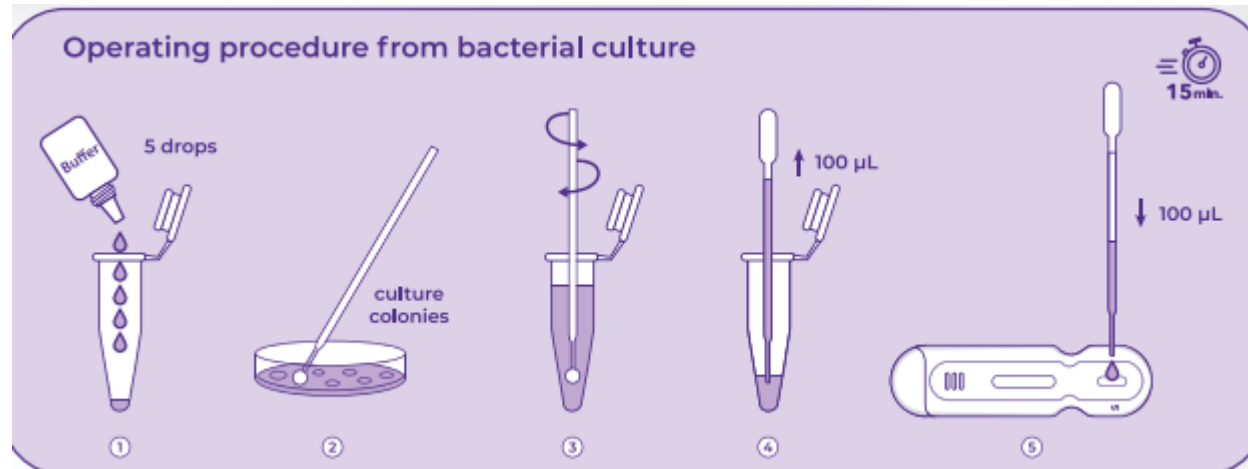
OXA-40/58 +
A. pittii



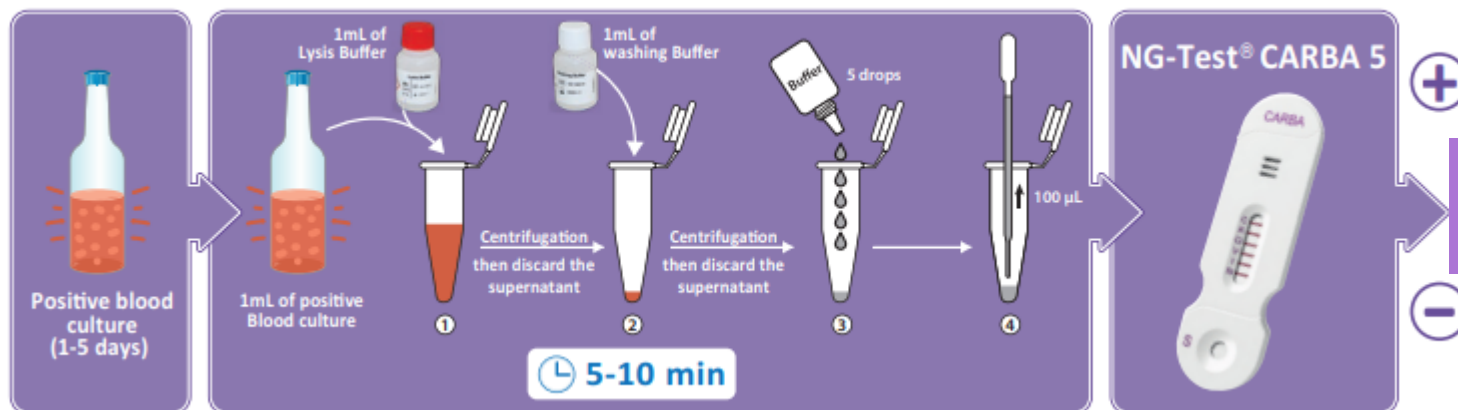
OXA-23 +
A. radioresistens

Overall sensitivity and specificity: 100% and 96%
PPA and NPA 91% and 100%.

After exclusion of *A. pittii* and *A. radioresistens* results, sensitivity, specificity, PPA and NPA were 100%.



IDENTIFICATION PROCESS FROM DIRECT BLOOD CULTURE: 20–40 min protocol



Bodendoerfer E et al. J Antimicrob Chemother. 2019;74(6):1749-1751.

NG•TEST®/CARBA-5



Boutal H et al. J Antimicrob Chemother. 2018;73:909-15.

Han R et al. Front Microbiol. 2021;11:609856.

Volland H, et al. Antimicrob Agents Chemother. 2019;64(1):e01940-19.

NG•TEST®/CTX-M Multi

Rapid detection of Extended Spectrum Beta-Lactamase (ESBL)



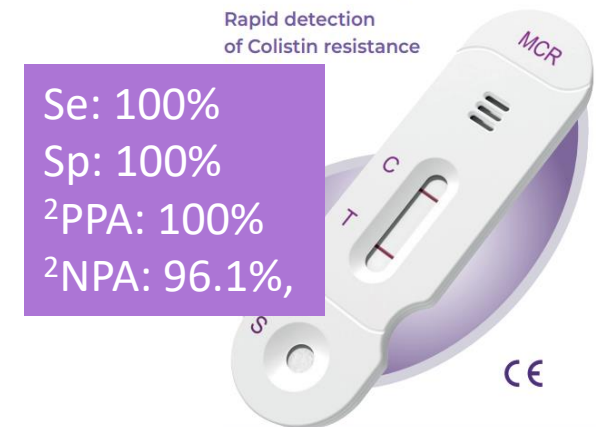
Bernabeu S et al. Diagnostics (Basel). 2020;10(10):764.

Bianco G, et al. J Hosp Infect. 2020;105(2):341-343.

Boattini M et al. Eur J Clin Microbiol Infect Dis. 2021;40(7):1495-1501.

NG•TEST®/MCR-1

Rapid detection of Colistin resistance



Volland H et al. J Clin Microbiol. 2019;57:e01454-18.

Fenwick AJ et al. J Clin Microbiol. 2020;58(4):e01823-19.

LFIA και ανίχνευση φορέων απευθείας από το δείγμα

J Antimicrob Chemother. 2019 Feb 1;74(2):357-359. doi: 10.1093/jac/dky429.

Lateral flow immunochromatographic assay for rapid screening of faecal carriage of carbapenemase-producing Enterobacteriaceae

Charlotte Fauconnier¹, Magali Dodemont², Angélique Depouhon¹, Ahalieyah Anantharajah¹, Alexia Verroken¹, Hector Rodriguez-Villalobos¹

Residual rectal swabs ($n=149$), routinely screened for CPE through selective culture and confirmed by PCR

Protocol: 2.5h incubation of the swab in an enrichment medium containing meropenem followed by **O.K.N. K-SeT** testing after centrifugation.

Strains: 82 OXA-48, 22 NDM, 5 KPC, 1 IMP, 7 VIM, 3 OXA-23

Detected: 76 OXA-48, 19 NDM, 5 KPC

Sensitivity: 96%

Specificity: 100%

LoD: 10^4 - 10^5 cfu/ml

Microorganisms. 2021 Apr 27;9(5):942. doi: 10.3390/microorganisms9050942.

Detection of KPC, NDM and VIM-Producing Organisms Directly from Rectal Swabs by a Multiplex Lateral Flow Immunoassay

Alexandra Vasilakopoulou¹, Polyxeni Karakosta¹, Sophia Vourli¹, Eleni Kalogeropoulou¹, Spyros Pournaras¹

Fecal swabs ($n=30$)

- Culture
- PCR
- **NG-Test® CARBA-5**

Sensitivity:

80% (direct testing)
88% (1h incubation)
92% (2h incubation)
100% (3h incubation)

Specificity: 100%

Each sample was tested by CARBSA-5 five times:

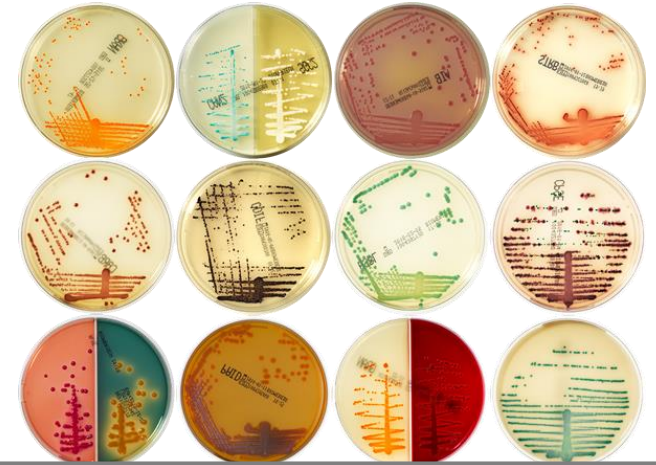
- Direct testing
- Incubation in TSB for 1h, 2h, 3h, 4h

TSB enriched with 0.25mg/l meropenem

Concordant results from LFIA and PCR: 11 KPC, 8 VIM, 6 NDM. [In five cases, co-carriage of KPC and VIM]

Χρωμογόνα υλικά

- Purpose: more rapid detection and identification of MDRO
- The target organisms are characterized by specific enzyme systems that metabolize the substrates to release the chromogen.
- The chromogen can then be visually detected by direct observation of a distinct colour change in the medium (coloured colonies).
- Compared with the use of conventional culture media, the use of chromogenic agar often reduces the costs and labor time.
- Their primary use is for screening of patients colonized with various pathogens.
- The sensitivity and specificity depend on the manufacturer and the type of microorganism detected
- Additional confirmation of the resistant bacteria is sometimes needed.



Advantage

Mass use and the possibility of automatisisation
Simple to perform
Simple and fast interpretation

Disadvantage

Not completely susceptible and specific
Time-consuming
Limited spectra or single antibiotic
Relatively expensive
Screening only or required confirmatory identification
No MIC value

Comments

Qualitative with no interpretation criteria (S, I, R)

Χρωμογόνα εκλεκτικά υλικά για CRE

CHROMagar KPC, Colorex KPC (CHROMagar)



Klebsiella, Enterobacter, Citrobacter CR
→ **Metallic blue**
Pseudomonas CR → Cream, translucent
E. coli CR → **dark pink to reddish**

- Μειονέκτημα: έλλειψη ευαισθησίας
- Δυσκολία ανάπτυξης στελεχών με $MIC \leq 4 \mu g/ml$
- Προβληματική η ανίχνευση στελεχών MBL και OXA-48

ChromID Carba (bioMérieux)

E. coli:
pink
to burgundy

1 - *E. coli*
(NDM-1)
2 - *P. aeruginosa*
(IMP)

KESC group:
bluish-green
to bluish-grey

K. pneumoniae
(KPC)
Proteae: dark brown to brown

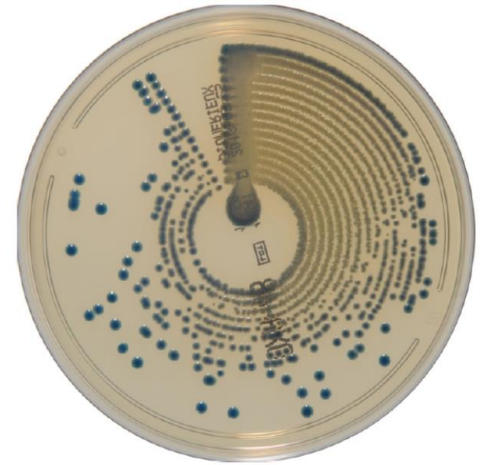
- SN: 92,4%, SP: 96, 9%
- Ικανότητα ανίχνευσης στελεχών με χαμηλές MIC^1
- Υψηλότερη ικανότητα ανίχνευσης για KPC-στελέχη vs Brilliance CRE /Colorex KPC (σε υψηλό και σε χαμηλό inoculum)²
- Υψηλότερη ικανότητα ανίχνευσης NDM-στελεχών³

1. Vriani G, et al. J Clin Microbiol. 2012; 50(6):1841-6.

2. Wilkinson KM, et al. J Clin Microbiol. 2012; 50(9):3102-4.

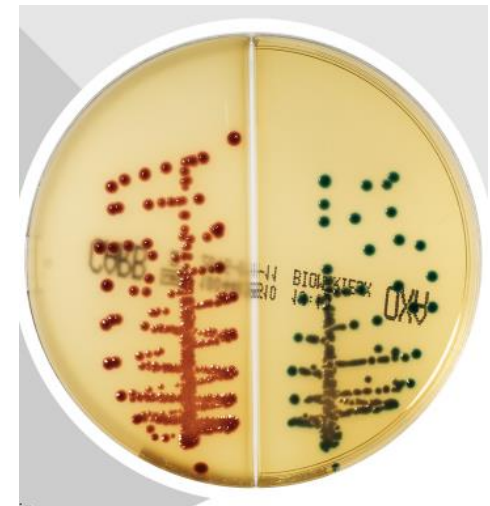
3. Perry JD, et al. J Antimicrob Chemother. 2011; 66(10):2288-94.

ChromID OXA-48 (bioMérieux)



Incubation : 18 hours
K. pneumoniae OXA-48

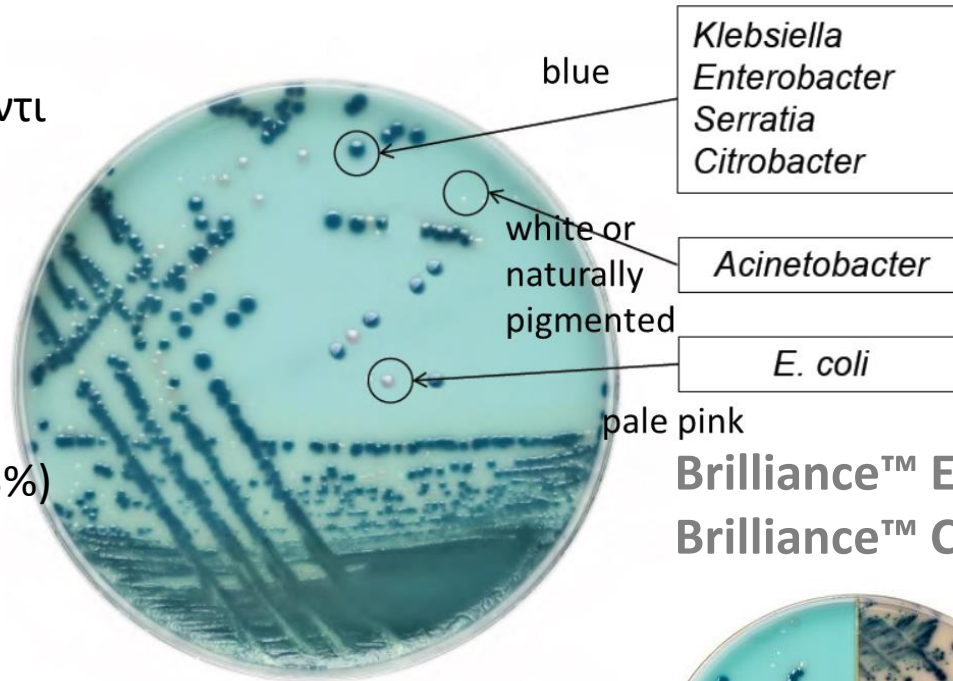
CHROMID® CARBA SMART Agar



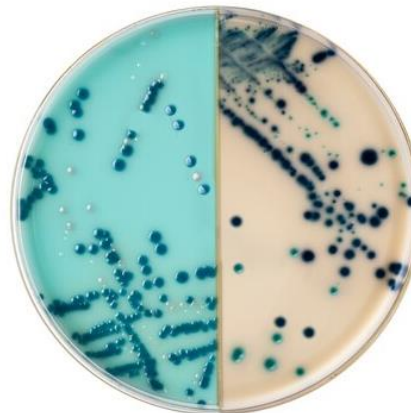
Χρωμογόνα εκλεκτικά υλικά για CRE

Brilliance™ CRE (Oxoid)

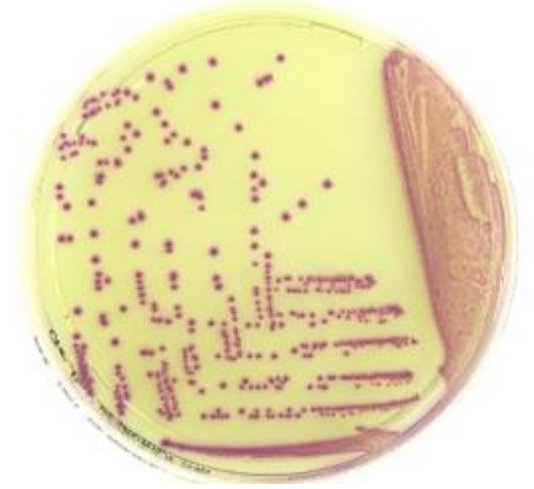
- Χαμηλότερη SN έναντι του ID Carba για την ανίχνευση NDM-στελεχών
- Υπεροχή έναντι του CHROMagar KPC (43%)
- Παρόμοια SP



Brilliance™ ESBL Agar/
Brilliance™ CRE Agar



Liofilchem® Chromatic CRE



Liofilchem® Chromatic OXA-48



OXA-48 positive *Enterobacter cloacae*

Έλεγχος φορέας για CRE: καλλιεργητικές μέθοδοι

Home-made τρυβλία MacConkey agar:

- Με ενσωματωμένη καρβαπενέμη
 - ικανοποιητική ευαισθησία
- Με δίσκους καρβαπενεμών:
 - SN/SP εξαρτώνται από το όριο
- Με δίσκους + αναστολείς:
 - **ανίχνευση + διαφοροποίηση**

Χρωμογόνα υλικά

- **chromID CARBA / chromID OXA-48, bioMérieux**
 - Υψηλή ευαισθησία και ειδικότητα
- **Brilliance CRE, Oxoid**
 - Ικανοποιητική ευαισθησία και ειδικότητα
- **CHROMagar KPC/ Colorex KPC, CHROMagar**
 - Ανιχνεύει στελέχη με υψηλού επιπέδου αντοχή

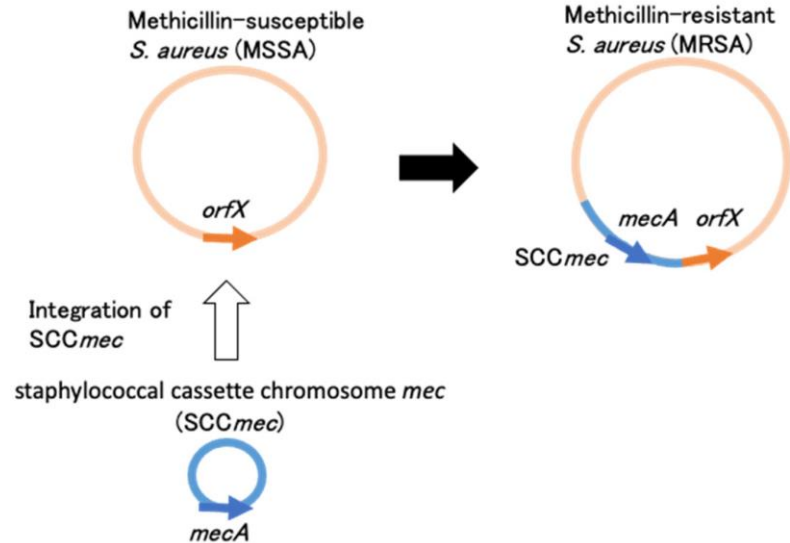
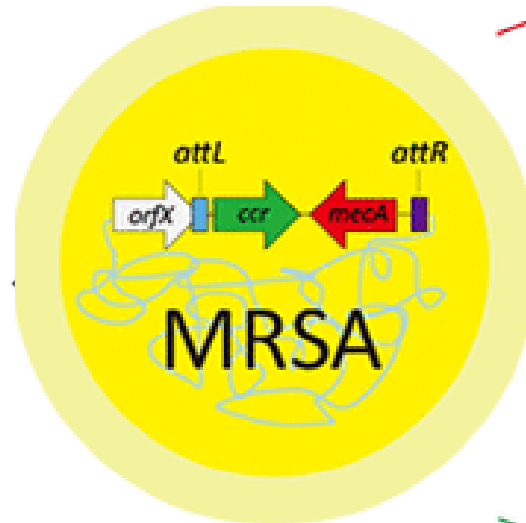
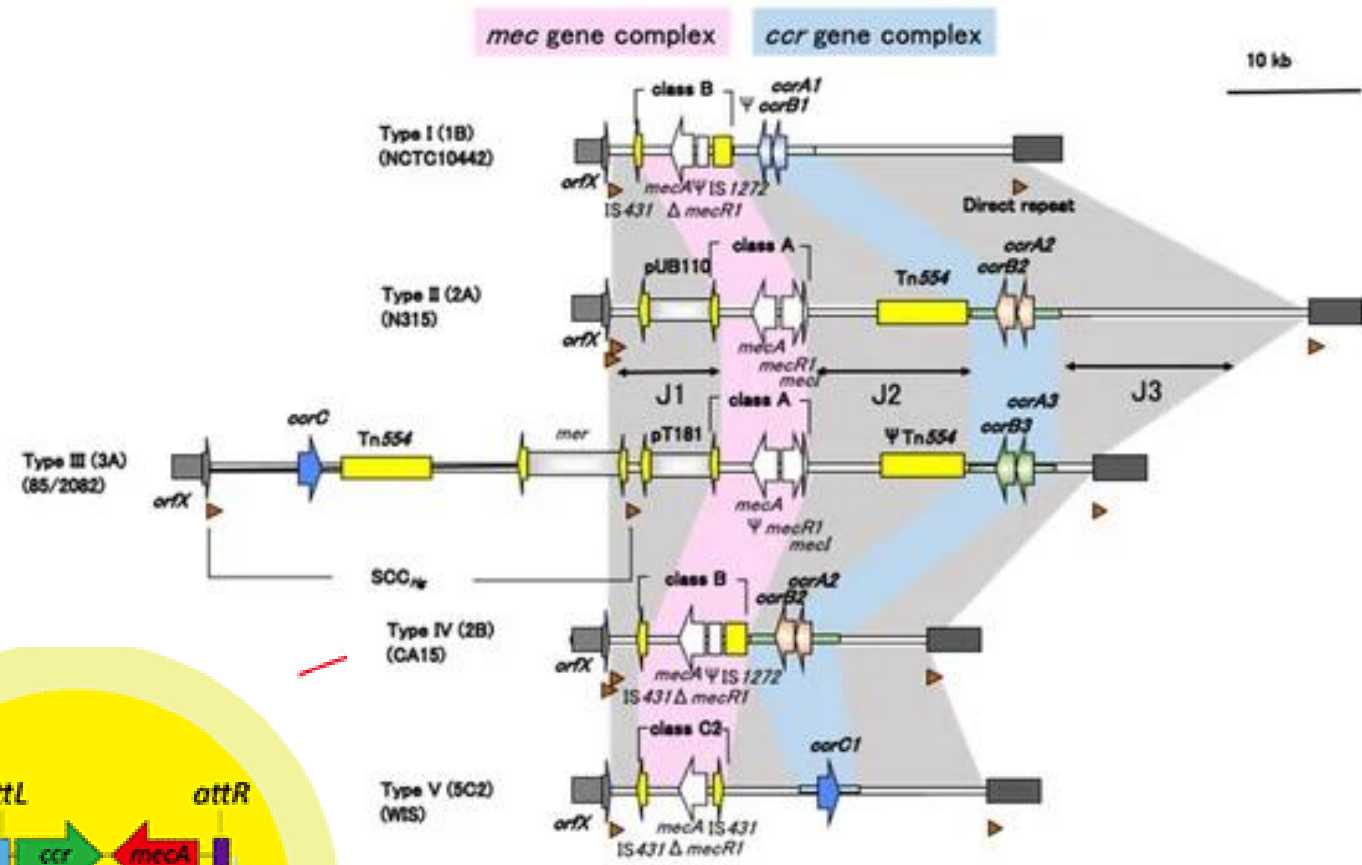
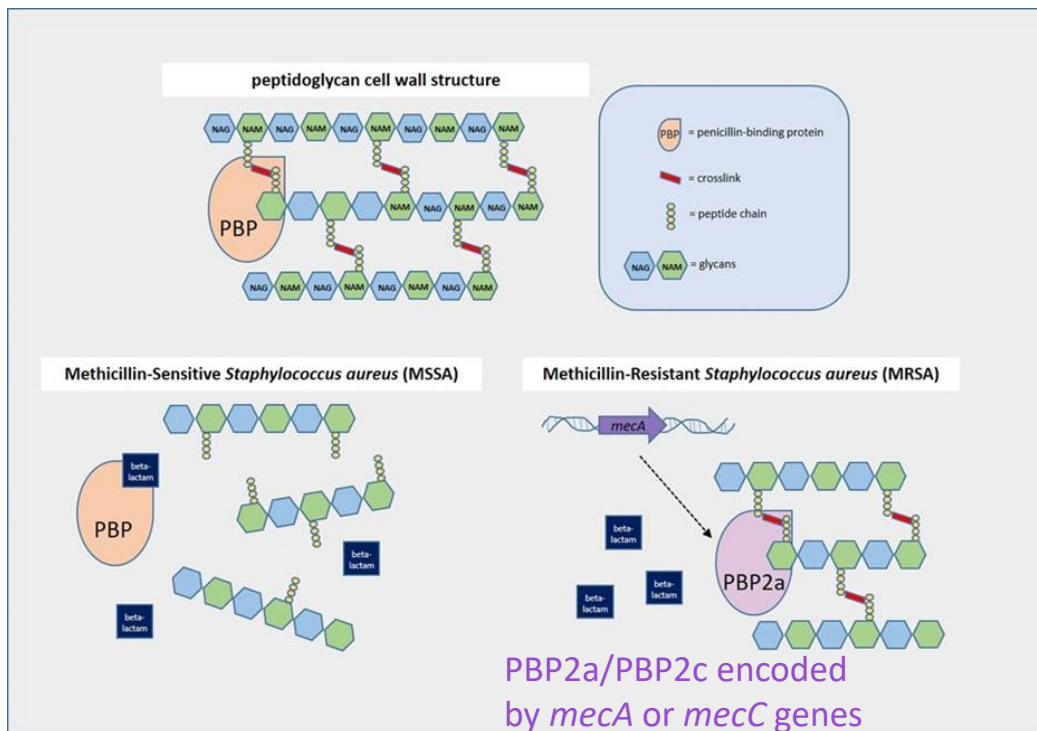


Pournaras S, et al. J Clin Microbiol. 2013; 51(9):2986-90.

Η επίδοση της κάθε μεθόδου εξαρτάται:

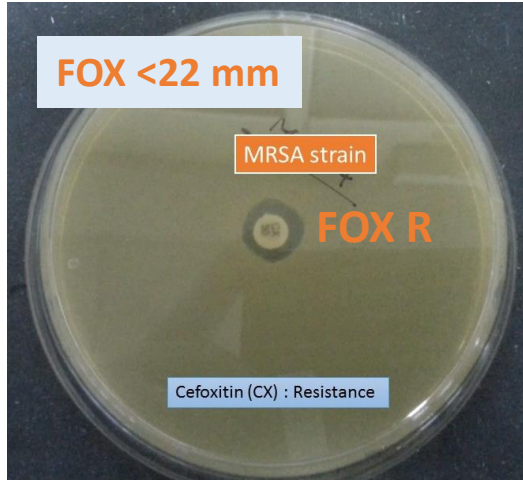
- επίπεδο αντοχής στις καρβαπενέμες
- υδρόλυση άλλων β-λακταμών (φάσμα)
- παρουσία άλλων μηχανισμών αντοχής
- γεωγραφική περιοχή, βακτηριακό είδος, δείγμα, χρόνο επώασης, μέθοδο αναφοράς, ορισμό αληθώς θετικών, ...

Σύγκριση μελετών-μεθόδων
δύσκολη



Φαινοτυπικές μέθοδοι

Disk diffusion



CEFOXITIN: a very **sensitive and specific marker** of *mecA/mecC*-mediated methicillin resistance including in heterogeneous expressing strains and is the **agent of choice**.

Broth microdilution



Organism Quantity:

Selected Organism: Staphylococcus aureus

Comments:	S.aureus - Mupirocin: ECOFF

Identification Information	
Organism Origin	Technologist
Selected Organism	Staphylococcus aureus
	Entered: Oct 7, 2023 17:56 EEST By: LabTech
Analysis Messages:	

Susceptibility Information	Card:	AST-P659	Lot Number:	8092252103	Expires:	Jan 26, 2024 12:00 EET
	Status:	Final	Analysis Time:	8.48 hours	Completed:	Oct 8, 2023 02:22 EEST

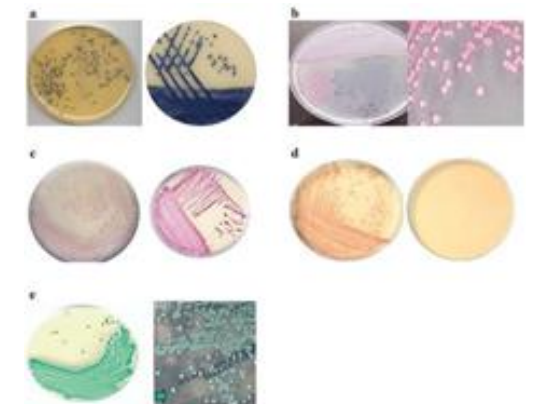
Antimicrobial	MIC	Interpretation	Antimicrobial	MIC	Interpretation
Cefoxitin Screen	POS	+	Linezolid	2	S
Benzylpenicillin	>= 0.5	R	Daptomycin	0.25	S
Oxacillin	>= 4	R	Teicoplanin	<= 0.5	S
Ceftaroline			Vancomycin	<= 0.5	S
Pneumonia	0.25	S	Tetracycline	<= 1	S
Other	0.25	S	Tigecycline	<= 0.12	S
Gentamicin	<= 0.5	S	Fusidic Acid	8	R
Levofloxacin	>= 8	R	Mupirocin	<= 1	S
Inducible Clindamycin Resistance	NEG	-	Rifampicin	<= 0.03	S
Erythromycin	0.5	S	Trimethoprim/ Sulfamethoxazole	<= 10	S
Clindamycin	0.25	S			

Detection of PBP2a with latex agglutination kits



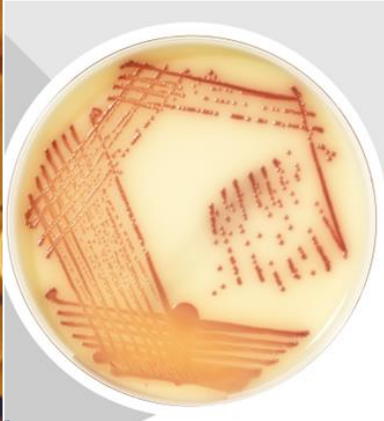
*PBP2c is not detected by the majority of commercial assays

Chromogenic media



Χρωμογόνα εκλεκτικά υλικά για MRSA

CHROMID® MRSA
SMART, bioMerieux



ChromID *S. aureus*/
ChromID MRSA, bioMerieux



Brilliance™ MRSA 2
Agar, Oxoid



Brilliance™ MRSA 2 Agar/
Brilliance™ Staph 24 Agar



MRSA Select,
Bio-Rad



ChromAgar MRSA



Variable	CHROMagar [43]	BBL-CHROMagar [43]	CHROMagar MRSA [35]
Sensitivity (%)	82–93	83–94	96–100
Specificity (%)	97–99	98–99	95–97
Turnaround time (h)	24–48	24–48	24–48
Storage for incubation	2–8 °C; dark	2–8 °C; dark	2–8 °C; dark

Variable	MRSASelect [35]	Brilliance MRSA agar [43]	ChromID [43]
Sensitivity (%)	81–93	90–96	83–94
Specificity (%)	92–97	69–87	90–96
Turnaround time (h)	18–24	18–24	
Storage for incubation	Room temperature	2–10 °C; dark	Room temperature

Harbarth S et al. Int J Antimicrob Agents. 2011;37(2):110-7.

Nonhoff C et al. Eur J Clin Microbiol Infect Dis. 2009 Apr;28(4):363-9.

Malhotra-Kumar S et al. J Clin Microbiol. 2010 Apr;48(4):1040-6.

Φαινοτυπικές μέθοδοι για ανίχνευση του MRSA

Phenotypic Method Used	Principle of the Method	On Culture/on Clinical Sample	TAT ¹	Brief Advan- tages/Disadvantages +/-	No. and Type of <i>S. aureus</i> Analyzed ²	Major Diagnostic Performance ³	Reference
Chromogenic media	Selective media, colorimetric colony detection	Nasal swabs, positive blood culture	24 h	+ Easy to perform and interpret, low-cost – No MSSA identification	72 MRSA 28 MSSA	Sensitivity 100% Specificity 78.6%	[28]
					19 MRSA 29 MSSA	Sensitivity 78.9% Specificity 41.3%	[29]
Rapid BMD	Colorimetric assay, MTT dye as oxidation-reactions indicator	Culture	7 h	+ Easy to perform, rapid, low-cost – Larger validations needed	21 MRSA 19 MSSA	Sensitivity 100% Specificity 94.7%	[30]
Disk diffusion on Mueller-Hinton rapid agar MHR-SIR	Disk diffusion with shorter incubation	Positive blood culture	6-8 h	+ Easy to perform, rapid, low-cost – Larger validations needed	23 <i>S. aureus</i> tested for several antibiotics	CA 97.8% mEs 1.9% VME 0.3%	[31]
					29 MRSA 50 MSSA	TAT 17h shorter, better therapy de-escalation	[32]
Rapid AST	Disk diffusion with shorter incubation	Positive blood culture	4-6 h	+ Easy to perform, rapid, low-cost – Low growth, diameters margins less demarcated	24 MRSA 313 MSSA	Sensitivity 100% Specificity 100%	[33]
					14 MRSA 183 MSSA	Sensitivity 100% Specificity 99.4% ATU 0.5%	[34]
					9 MRSA 212 MSSA	Sensitivity 100% Specificity 99.1% ATU 0.4% ME 0.5%	[35]

Φαινοτυπικές μέθοδοι για ανίχνευση του MRSA

Phenotypic Method Used	Principle of the Method	On Culture/on Clinical Sample	TAT ¹	Brief Advantages/Disadvantages +/–	No. and Type of <i>S. aureus</i> Analyzed ²	Major Diagnostic Performance ³	Reference
VITEK 2	Turbidimetric monitoring of bacterial growth	Culture	4–11 h	+ Automated, rapid, standardized interpretation – Fixed range of testable antibiotics, less accurate for linezolid/vancomycin	72 MRSA 28 MSSA	Sensitivity 97.2% Specificity 100%	[28]
					27 MRSA resistant to linezolid	MEs 85.2%	[36]
					28 MRSA with a vancomycin MIC ≥ 2	MEs 46.4%	[37]
BD Phoenix	Turbidimetric and colorimetric growth detection	Culture	6–16 h	+ Automated, rapid, standardized interpretation – Less accurate for SXT	642 <i>S. aureus</i> analyzed for SXT resistance	CA 91.9% MEs 7.9% mEs 0.3%	[38]
Copan WAsP Colibri	Automated disk diffusion	Culture	16 h	+ Automated, flexible, detect heteroresistance and mixed cultures – Should be compared with standard disk diffusion	107 <i>S. aureus</i> tested for several antibiotics	CA 99.9%	[39]
Accelerate PhenoTest™ BC	Single-cell analysis, fluorescent in situ hybridization	Positive blood culture	7 h	+ Automated, monomicrobial call, rapid, identification and AST in the same platform	98 MRSA 86 MSSA	Sensitivity 100% Specificity 98.8% CA 99.5%	[17]
					22 MRSA 2 MSSA	Sensitivity 100% Specificity 100%	[40]
PBP2a latex agglutination assay	Particles with monoclonal antibodies, agglutination reaction	Culture	5 m	+ Rapid, easy to perform and interpret – no PBP2c detection	95 MRSA 10 MSSA	Sensitivity 98.95% Specificity 77.8%	[41]

Μοριακές μέθοδοι για ανίχνευση του MRSA απευθείας από το δείγμα: παραδείγματα

Molecular Method Used	Principle of the Method	On Culture/ on Clinical Sample	TAT ¹
Xpert [®] SA Nasal Complete (Cepheid)	Real-time PCR for <i>mecA/C</i> , <i>spa</i> and <i>SCCmec-orfX</i>	Nasal samples	1 h
Xpert [®] MRSA/SA BC Assay (Cepheid)	Real-time PCR for <i>mecA/C</i> , <i>spa</i> and <i>SCCmec-orfX</i>	Positive blood cultures	1.7 h
Xpert [®] MRSA/SA SSTI (Cepheid)	Real-time PCR for <i>mecA/C</i> , <i>spa</i> and <i>SCCmec-orfX</i>	BAL samples	68 m



Molecular Method Used	Principle of the Method	On Culture/ on Clinical Sample	TAT ¹	Brief Advan- tages/Disadvantages +/-
Hologic Panther Fusion [®] MRSA	PCR and Invader chemistries for <i>mecA/C</i> , <i>gap</i> and <i>SCCmec-orfX</i>	Nasal samples	<3 h	+ Can analyze 350 samples in 8 h – Need comparison with a similar method
MRSA/SA ELITe MGB assay (ELITech-Group)	Real time PCR for <i>mecA/C</i> and a <i>S. aureus</i> specific sequence	Sputum, tracheal aspirate, BAL	<3 h	+ Accurate – Do not target <i>SCCmec-orfX</i> junction
Eazyplex [®] MRSA	LAMP targeting <i>S. aureus</i> , <i>S. epidermidis</i> , <i>mecA/C</i>	Positive blood cultures	1 h	+ Portable; faster than similar methods – Need optimization for CONS

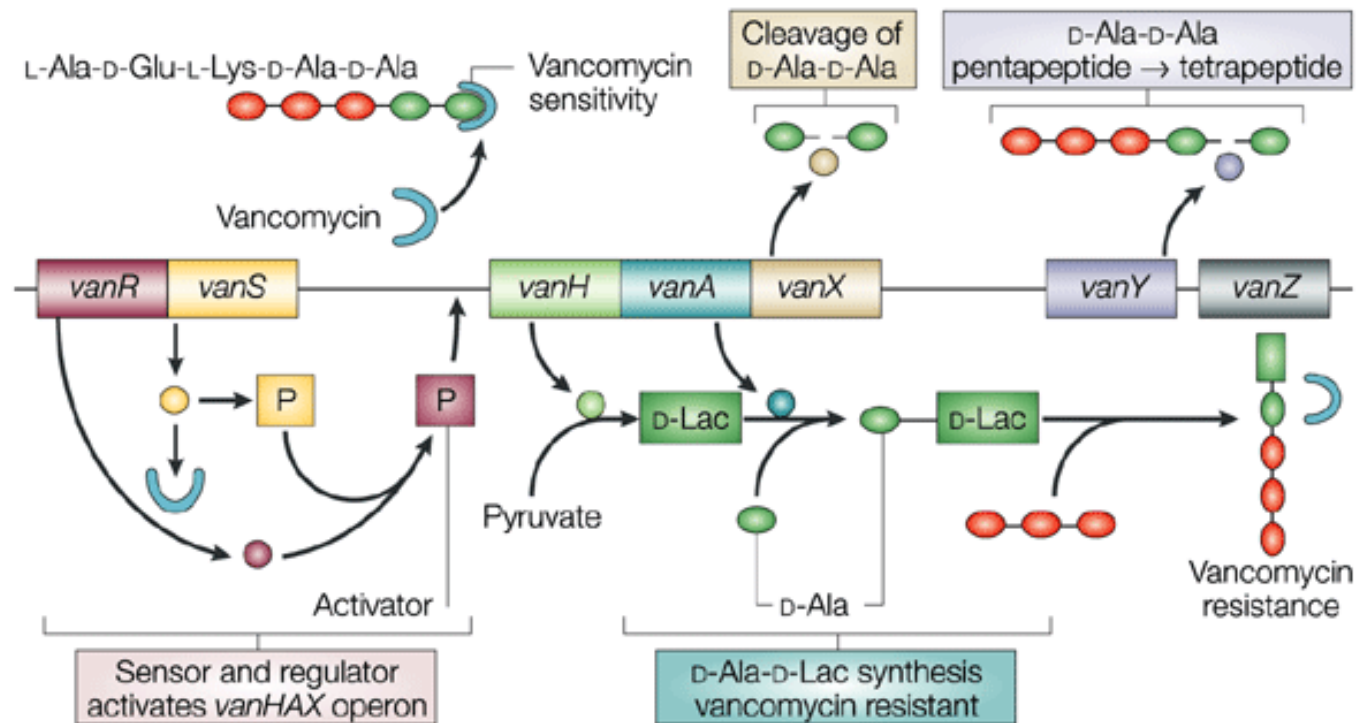
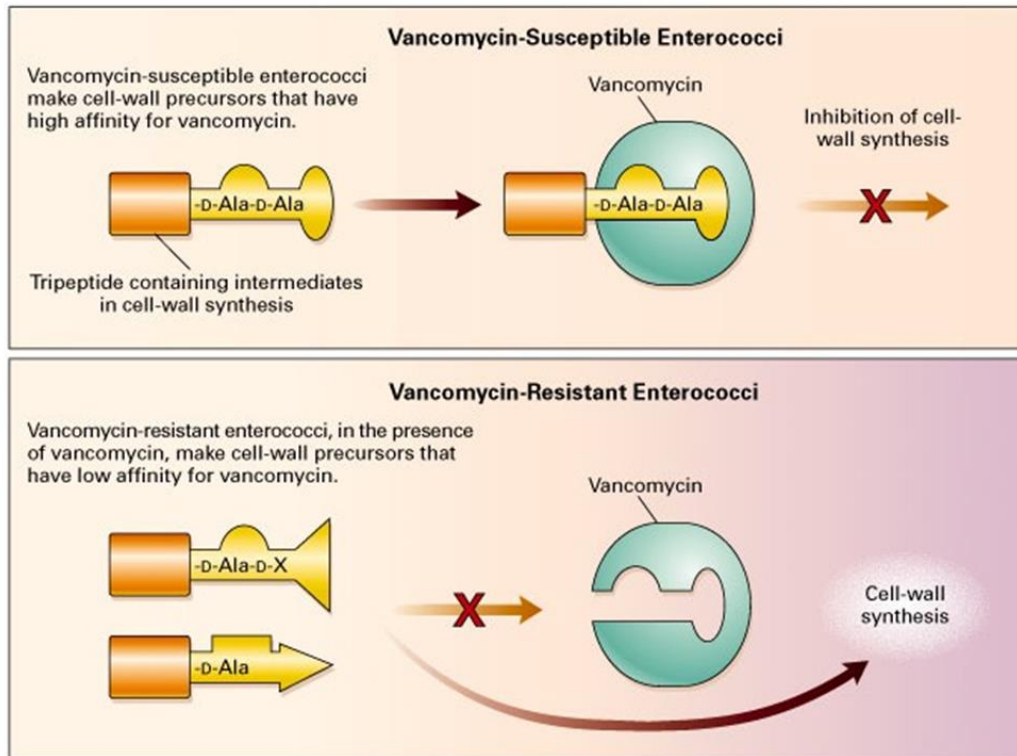


VRE, μηχανισμός αντοχής: τροποποίηση στόχου δράσης

Σύνθεση πεπτιδογλυκάνης που φέρει
πλευρικές πενταπεπτιδικές
αλυσίδες που καταλήγουν σε

D-ala-D-lactate
(VanA, VanB, VanD)

D-ala-D-serine
(VanC, VanE,
VanG, VanL, VanN)

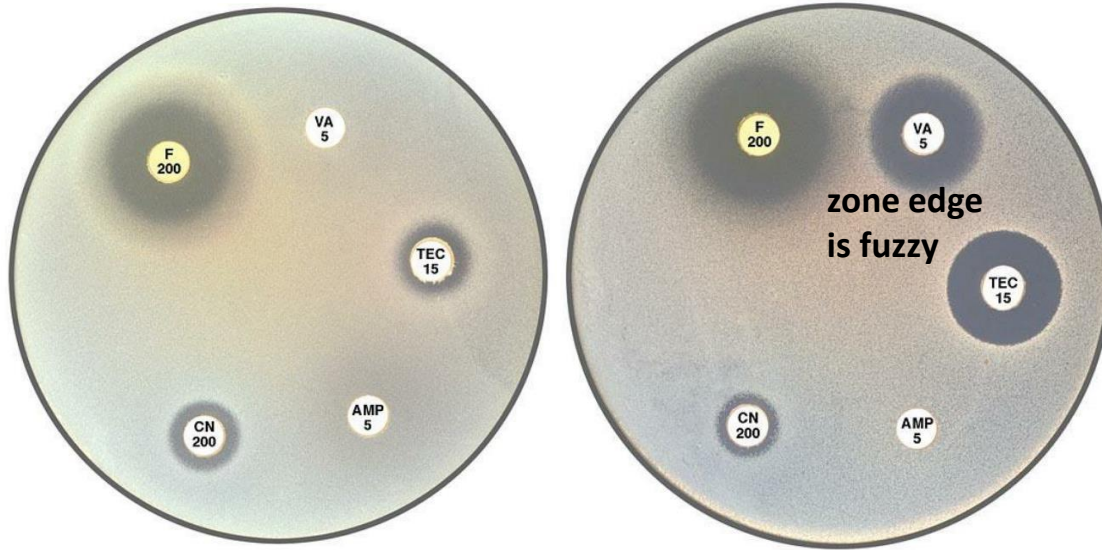


Glycopeptide resistance due to *van*-type gene clusters

Resistance level	Acquired							Intrinsic
	High	Variable	Moderate	Low				Low
Type	VanA	VanB	VanD	VanE	VanG	VanL	VanN	VanC
MIC in mg/L:								
Vancomycin	≥16	≥4	≥64	6–32	12–16	8	8	2–32
Teicoplanin	>8	0,5–1	4–64	0,5	0,5			0,5–1
Expression	Inducible	Inducible	Constitutive/ Inducible (<i>vanD2</i>)	Inducible/ (Constitutive)	Inducible			Constitutive/ Inducible
<i>van</i> ligase gene	<i>vanA</i>	<i>vanB1–B3</i>	<i>vanD1–5</i>	<i>vanE</i>	<i>vanG1–2</i>	<i>vanL</i>	<i>vanN</i>	<i>vanC1–C3</i>
Modified target	D-alanine-D-lactate	D-alanine-D-lactate	D-alanine-D-lactate	D-alanine-D-serine	D-alanine-D-serine	D-alanine-D-serine	D-alanine-D-serine?	D-alanine-D-serine
Conjugative transfer	Yes	Yes	No	No	Yes	No	Yes	No
Location	Plasmid/chromosome on transposon(s)	Plasmid/chromosome ± transposon/ICE ^a	Chromosome	Chromosome	Chromosome on possible ICE	Chromosome?	Plasmid	Chromosome
Distribution	<i>Enterococcus faecium</i> <i>Enterococcus faecalis</i> <i>Enterococcus avium</i> <i>Enterococcus casseliflavus</i> <i>Enterococcus durans</i> <i>Enterococcus gallinarum</i> <i>Enterococcus hirae</i> <i>Enterococcus mundtii</i> <i>Enterococcus raffinosus</i> <i>Staphylococcus aureus</i> <i>Bacillus circulans</i> <i>Oerskovia turbata</i> <i>Arcanobacterium haemolyticum</i> <i>Paenibacillus</i> <i>Rhodococcus</i>	<i>Enterococcus faecium</i> <i>Enterococcus faecalis</i> <i>Enterococcus casseliflavus</i> <i>Enterococcus durans</i> <i>Enterococcus gallinarum</i> <i>Enterococcus hirae</i> <i>Staphylococcus epidermidis</i> <i>Streptococcus</i> <i>Clostridium</i> <i>Ruminococcus</i> <i>Eggerthella</i>	<i>Enterococcus faecium</i> <i>Enterococcus faecalis</i> <i>Enterococcus avium</i> <i>Enterococcus gallinarum</i> <i>Enterococcus raffinosus</i> Non-enterococcal faecal flora	<i>Enterococcus faecalis</i>	<i>Enterococcus faecalis</i> Non-enterococcal faecal flora	<i>Enterococcus faecalis</i>	<i>Enterococcus faecium</i>	<i>Enterococcus gallinarum</i> – <i>vanC1</i> <i>Enterococcus casseliflavus</i> – <i>vanC2/3</i>

Ανίχνευση VRE: παραδείγματα

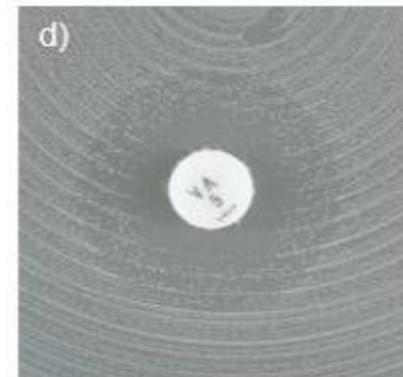
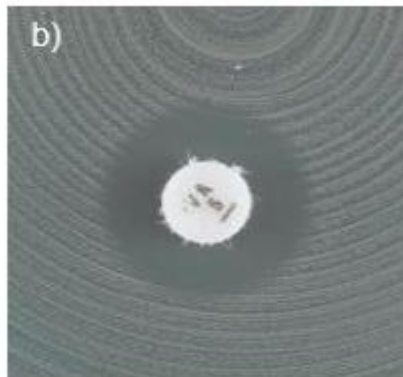
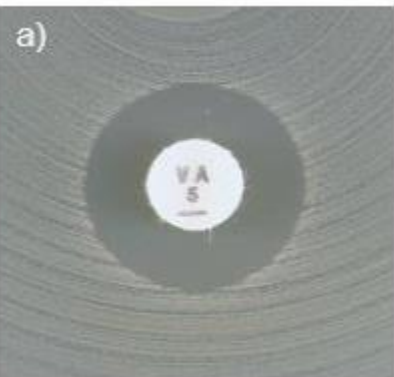
Μέθοδος διάχυσης των δίσκων



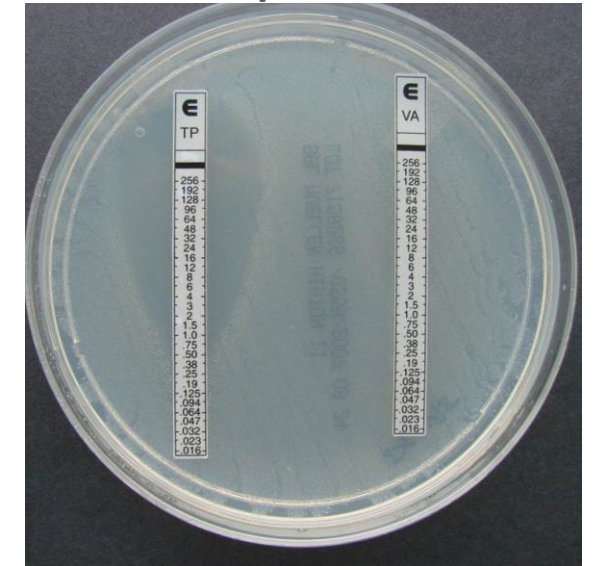
Isolates must not be reported susceptible before 24 h incubation

sharp zone edges

Confirmatory testing with PCR or report **resistant** even if the zone diameter is ≥ 12 mm



Ταινίες διαβαθμισμένης συγκέντρωσης αντιβιοτικών



Ανίχνευση VRE: παραδείγματα

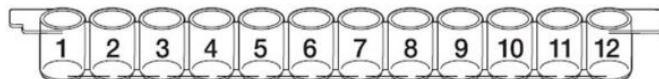
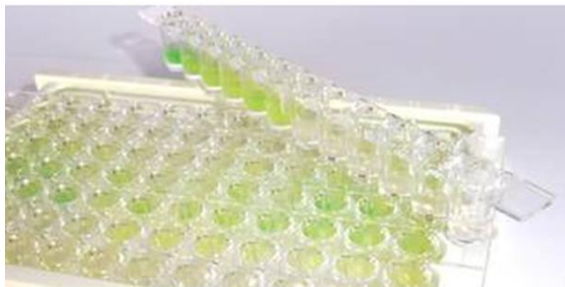
Αυτοματοποιημένα συστήματα

Bionumber: 532047265773771
Organism Quantity:

Selected Organism: Enterococcus faecium

Antimicrobial	MIC	Interpretation	Antimicrobial	MIC	Interpretation
Cefoxitin Screen			Moxifloxacin	(-)	(-)
Benzylpenicillin	(-)	(-)	Inducible Clindamycin Resistance		
Ampicillin	≥ 32	R	Erythromycin	(-)	(-)
Oxacillin			Clindamycin	(-)	(-)
Imipenem	≥ 16	R	Quinupristin/Dalfopristin	4	R
Gentamicin High Level (synergy)	SYN-S	S	Linezolid	1	S
Streptomycin High Level (synergy)	SYN-R	R	Daptomycin		
Gentamicin			Teicoplanin	≥ 32	R
Ciprofloxacin			Vancomycin	≥ 32	R
Urine	≥ 8	R	Tigecycline	≤ 0.12	S
Other	≥ 8		Trimethoprim/Sulfamethoxazole		

Broth microdilution



Μοριακές μέθοδοι απευθείας από το δείγμα

VanA or VanB resistance in 20 min, from rectal swabs, blood culture or as culture confirmation



VRE Cepheid Xpert® vanA/vanB



VanA SN= 87.1%, SP=99.7%, PPV=98.0%, NPV=97.7%

VanB SN=77.6%

The saving of time to results corresponded to 141 saved isolation days and 292 saved transmission risk days.



Development and validation of a lateral flow immunoassay for rapid detection of VanA-producing enterococci

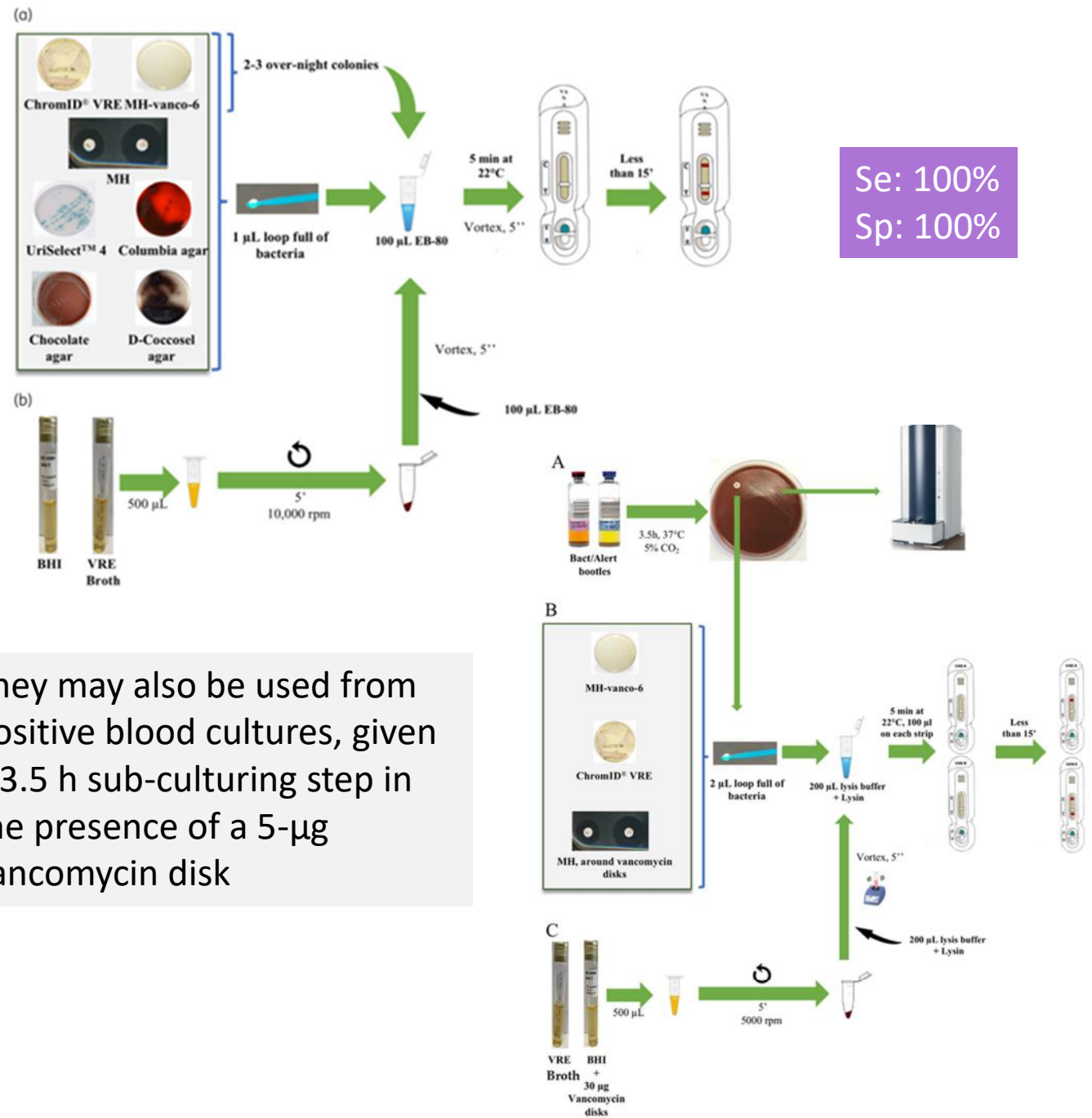
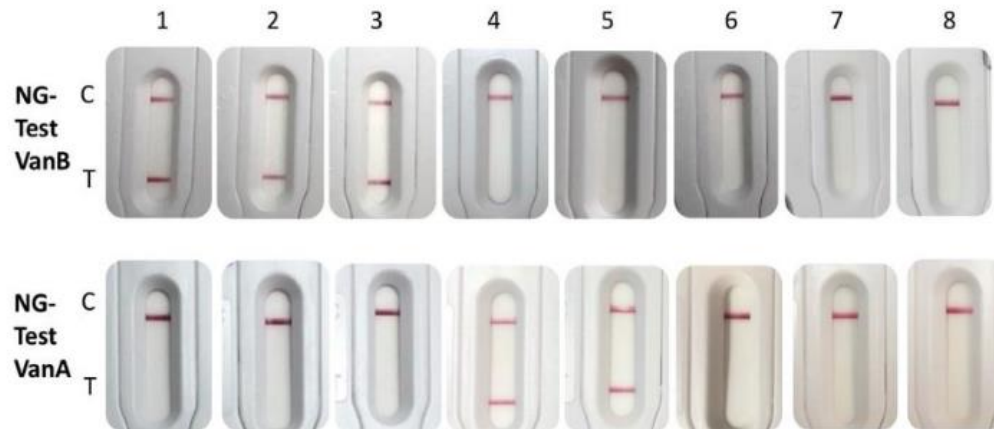
Saoussen Oueslati^{1,2,†}, Hervé Volland^{3,†}, Vincent Cattoir⁴, Sandrine Bernabeu^{1,2}, Delphine Girlich¹, Duncan Dulac³, Marc Plaisance³, Maxime Laroche⁵, Laurent Dortet^{1,2,6}, Stéphanie Simon³ and Thierry Naas^{1,2,6*}



Article

Rapid Detection of VanA/B-Producing Vancomycin-Resistant Enterococci Using Lateral Flow Immunoassay

Saoussen Oueslati^{1,2,†}, Camille Gonzalez^{1,2,†}, Hervé Volland^{3,*,†}, Vincent Cattoir⁴, Sandrine Bernabeu^{1,2}, Delphine Girlich^{1,2}, Duncan Dulac³, Marc Plaisance³, Laure Boutigny⁵, Laurent Dortet^{1,2}, Stéphanie Simon³ and Thierry Naas^{1,*,†}

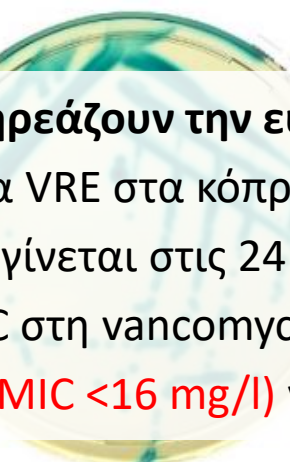


Χρωμογόνα εκλεκτικά υλικά για VRE

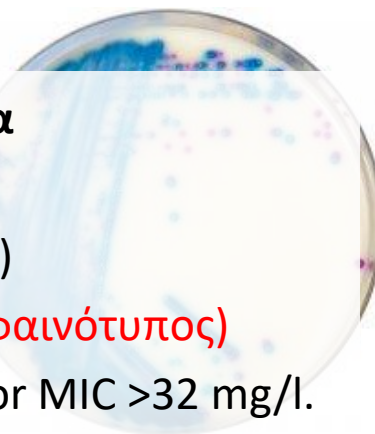
Brilliance VRE®, Oxoid



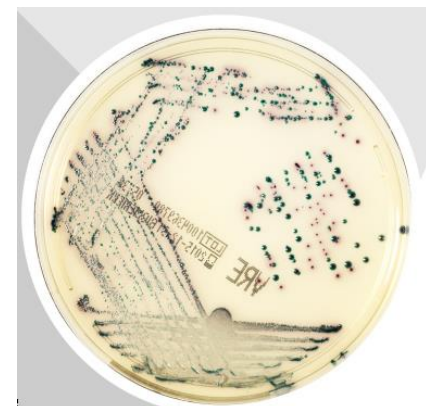
Chromatic VRE,
Liofilchem



ChromAgar® VRE



chromID® VRE,
bioMerieux



Bile-esculin-azide agar +
6 mg/L vancomycin



Παράγοντες που επηρεάζουν την ευαισθησία

- χαμηλή πυκνότητα VRE στα κόπρανα
- όταν η ανάγνωση γίνεται στις 24 h (vs 48 h)
- χαμηλές τιμές MIC στη vancomycin (**VanB φαινότυπος**)
 - **SN 25-75% (MIC <16 mg/l) vs 100% for MIC >32 mg/l.**

TABLE 2 Comparison 396 specimens (combined data)

Medium	Performance (% [95% CI]) ^b			
	Sensitivity	Specificity	PPV	NPV
BEAV broth	98.0 (94.7–98.0)	100 (98.9–100)	100 (96.7–100)	99.3 (98.3–99.3)
Enterococcosel (BEAV)	84.8 (80.9–84.8)	100 (98.7–100)	100 (95.3–100)	95.2 (93.9–95.2)
InTray Colorex VRE	91.9 (86.9–94.8)	98.3 (96.6–99.3)	94.8 (89.6–97.9)	97.3 (95.7–98.3)
chromID	94.9 (91.0–95.9)	99.7 (98.3–100)	98.9 (94.8–99.9)	98.3 (97.0–98.7)
VRESelect	91.9 (87.8–92.9)	99.7 (98.3–100)	98.9 (94.4–99.9)	97.4 (96.0–97.7)
HardyCHROM VRE	89.9 (85.6–90.9)	99.7 (98.2–100)	98.9 (94.2–99.9)	96.7 (95.4–97.0)
Spectra VRE	93.9 (89.9–94.9)	99.7 (98.3–100)	98.9 (94.7–99.9)	98.0 (96.7–98.3)

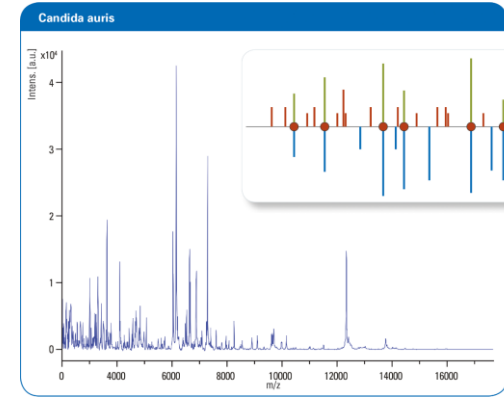
Τα χρωμογόνα υπερέχουν έναντι του BEAV: SN 89.9-93.9% vs 84.8%
Αποτελέσματα διαθέσιμα 24-48 h νωρίτερα

Επιλογή σύμφωνα με:

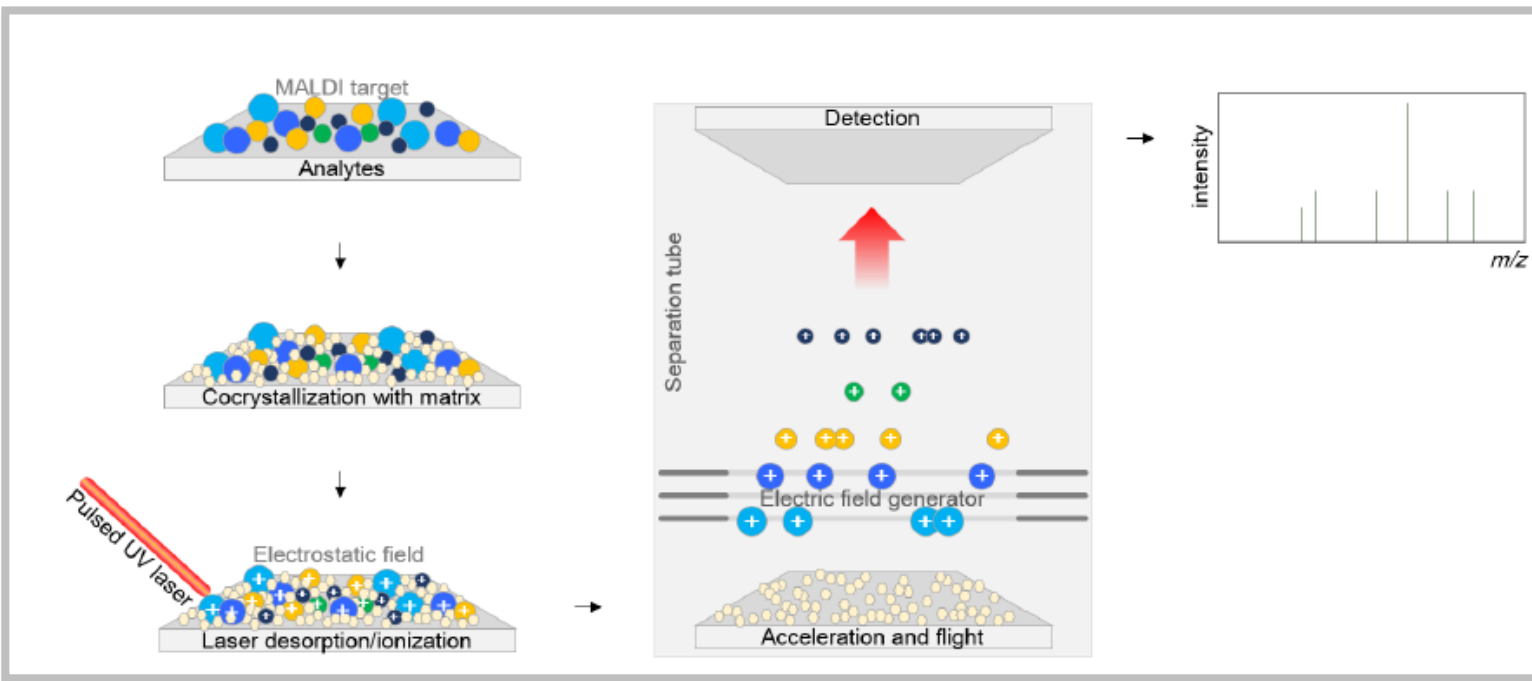
ευκολία διάκρισης αποικιών,
αξιοπιστία, κόστος, ανάγκη
επιβεβαιωτικών δοκιμασιών

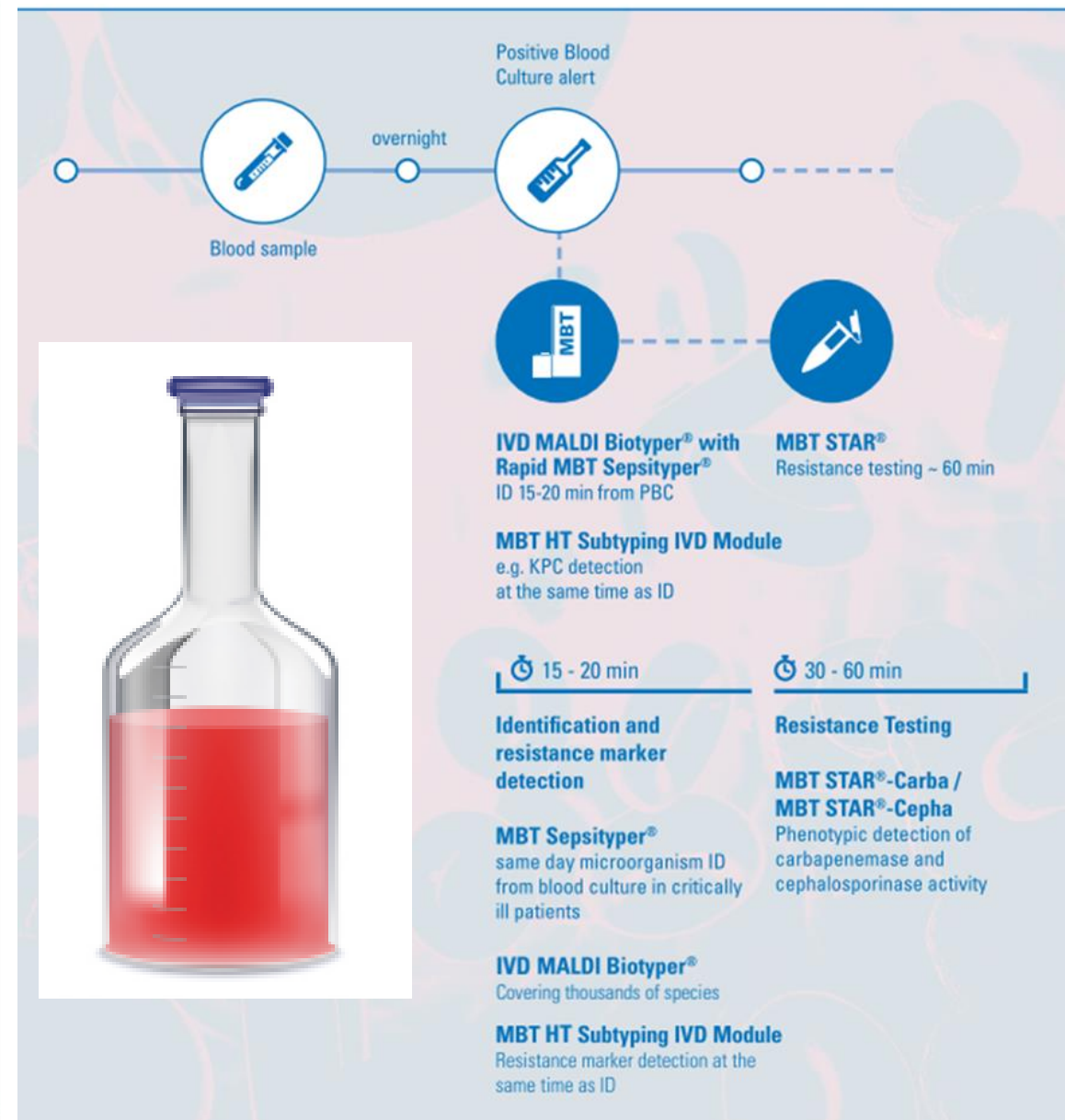
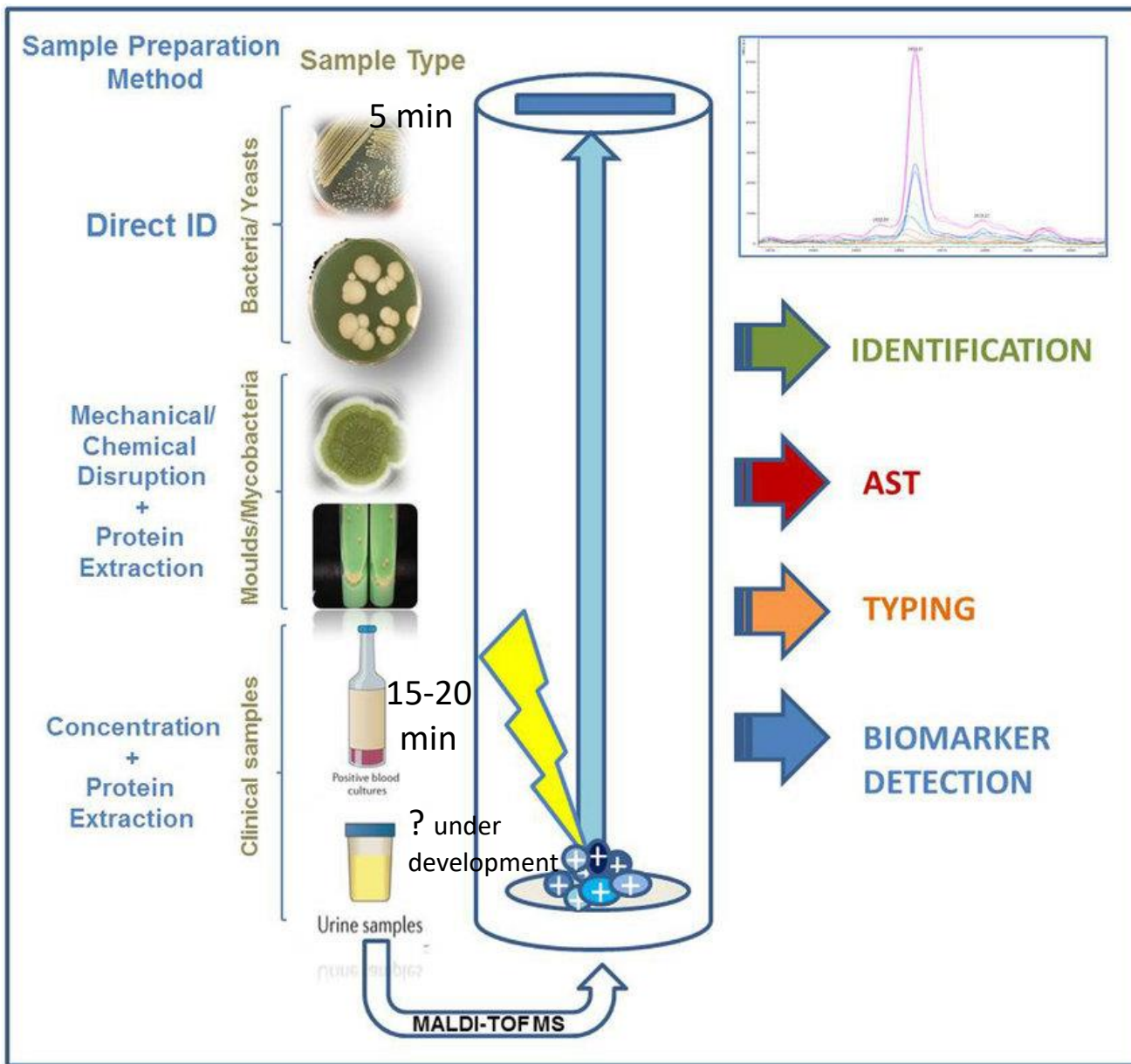
MALDI-TOF Mass Spectrometry και ανίχνευση αντοχής

Matrix-Assisted Laser Desorption Ionization
(MALDI) Time-Of-Flight (TOF)
Mass Spectrometry



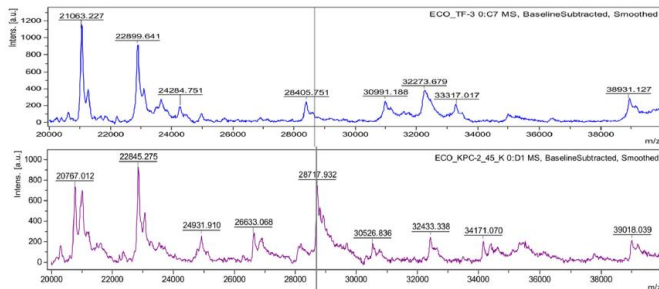
System analysis time to ID result:
95 isolates + 1 QC sample ~5 min





MALDI-TOF Mass Spectrometry και ανίχνευση αντοχής

Identification of the KPC producer



beta-lactamases
Porin defects

Direct detection
of AMR determinants

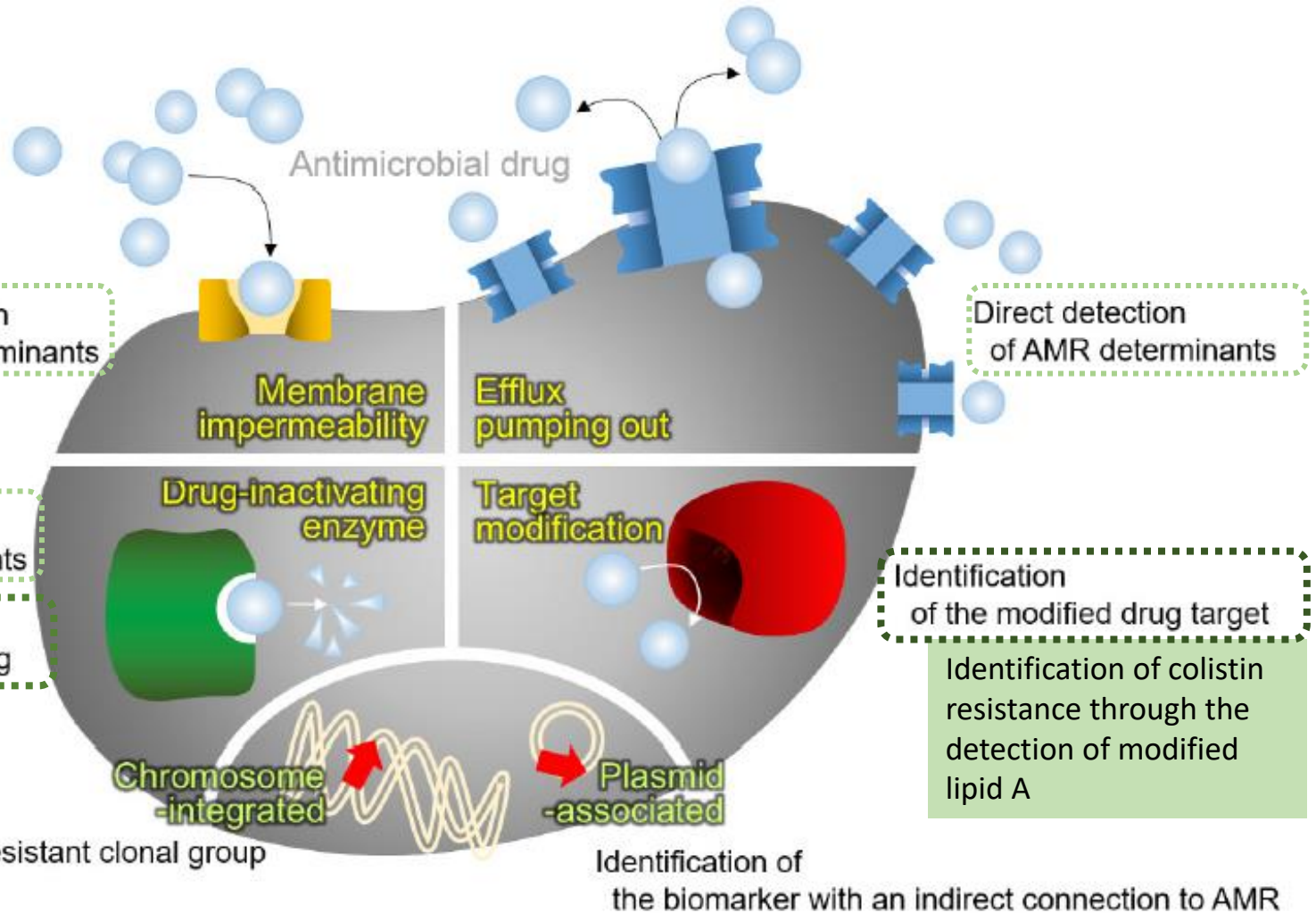
The modified antibiotics
change their molecular
weights. AMR is
identified by detecting the
peak shift using the MALDI-
TOF MS methodology

Direct detection
of AMR determinants

Identification
of the modified drug

MALDI-TOF MS protein
fingerprints of well-
known AMR
clonal groups

Identification of
the antimicrobial-resistant clonal group
Identification of
the biomarker, non-AMR determinants

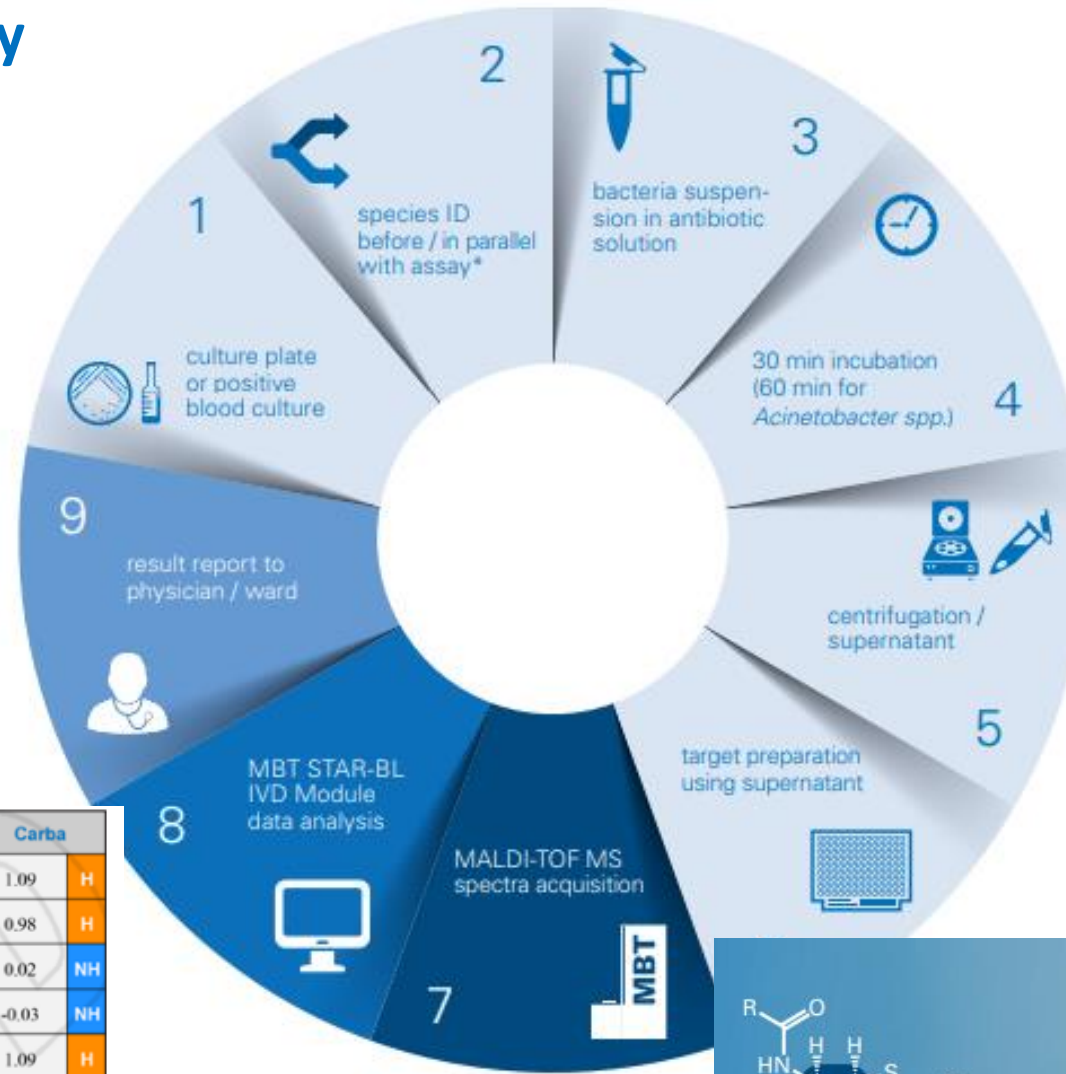


MALDI-TOF Mass Spectrometry και ανίχνευση αντοχής

MBT STAR®-Carba IVD Assay



Sensitivity 93.9%
specificity, 100%



Bacteria with active carbapenemase will inactivate the carbapenem antibiotic by hydrolysis of the β -lactam ring, **which is associated with a detectable mass shift.**

In assays with bacteria without active carbapenemase, only peaks corresponding to the intact antibiotic will be present in the mass spectrum.

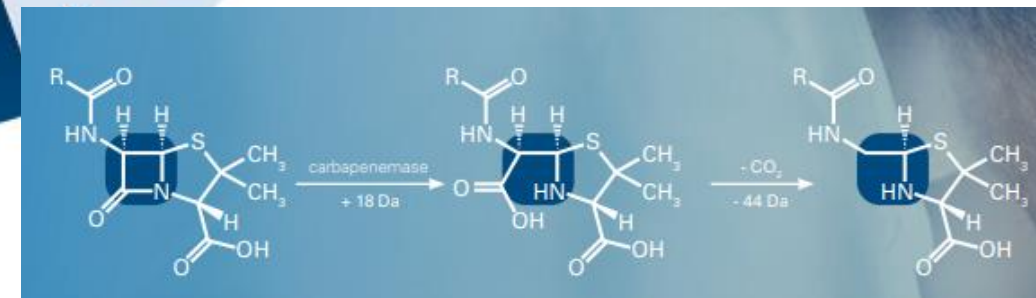
In assays with bacteria with active carbapenemase, peaks of the intact antibiotic will decrease.

Sample	Species	Control ID	Carba
Sample 1	Klebsiella pneumoniae	confirmed	1.09 H
Sample 2	Klebsiella pneumoniae	confirmed	0.98 H
Sample 3	Escherichia coli	confirmed	0.02 NH
neg. control		not performed	-0.03 NH
pos. control		not performed	1.09 H
Δ controls			1.47

H	Hydrolyzed ¹
NH	Non-hydrolyzed ²

Se: 100%, Sp: 100%

Del Corpo O et al. Clin Microbiol Infect.
2023 Sep 16:S1198-743X(23)00425-1.



MALDI-TOF Mass Spectrometry και ανίχνευση αντοχής

MBT STAR®-Cepha IVD Assay

-  Positive blood culture[§] or culture plate
-  Optional species ID before / in parallel (control ID) with assay*
-  Bacteria suspension in antibiotic solution
-  30 min incubation
-  MALDI target plate preparation
-  MALDI-TOF MS spectra acquisition and automated data analysis by MBT STAR-BL IVD Module

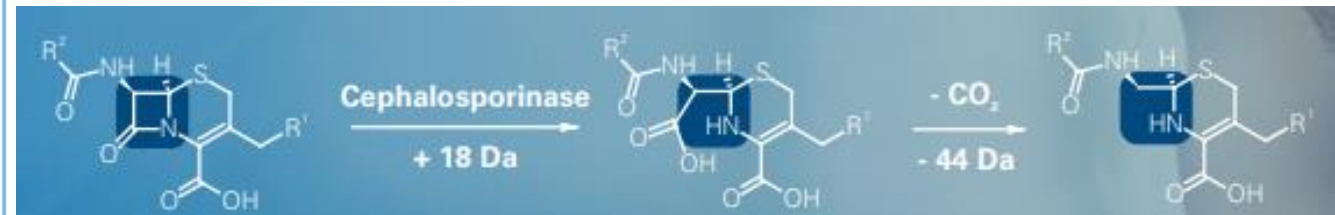


Se: 94%

Sp: 97%

Del Corpo O et al. Clin Microbiol Infect.
2023 Sep 16:S1198-743X(23)00425-1.

Hydrolysis of β -lactam ring leads to mass shifts that can easily be detected by MALDI-TOF mass spectrometry



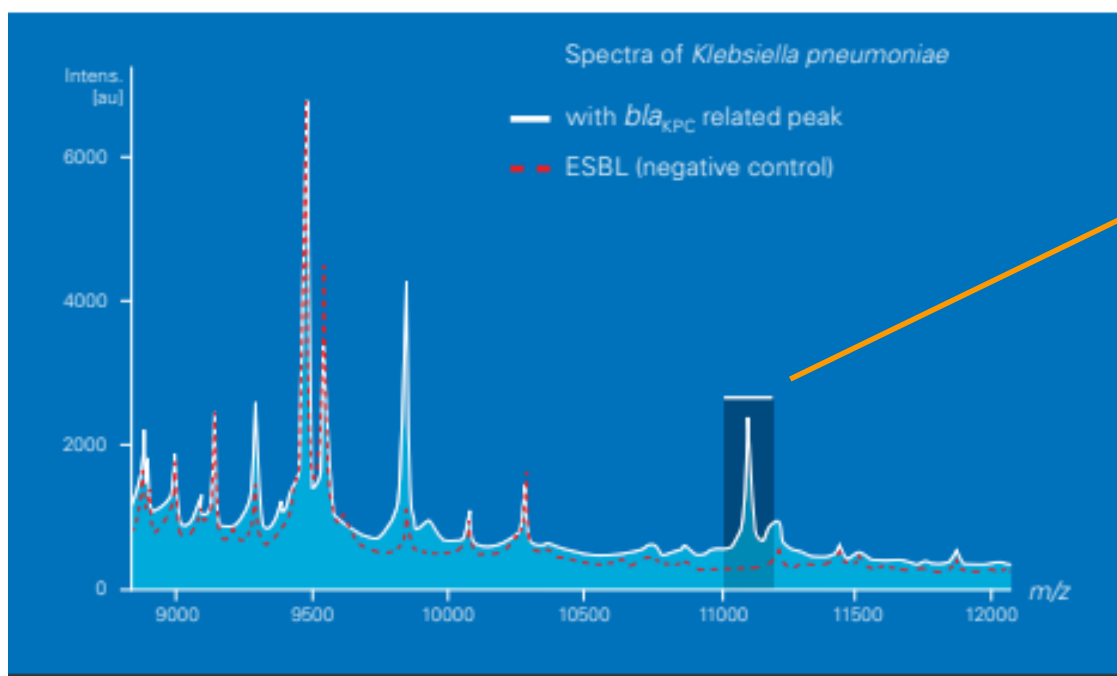
Sample	Species	Control ID	Cepha
Sample 1	Escherichia coli	confirmed	1.28 H
Sample 2	Escherichia coli	confirmed	1.14 H
Sample 3	Klebsiella variicola	confirmed	-0.55 NH
neg.control		not performed	-0.10 NH
pos.control		not performed	1.04 H
Δ controls			1.12

H	Hydrolyzed ¹
NH	Non-hydrolyzed ²

MALDI-TOF Mass Spectrometry και ανίχνευση αντοχής

MBT Subtyping IVD Module

Quick detection of *bla*_{KPC} expressing *K. pneumoniae* and *E. coli*



*A peak in MALDI-TOF mass spectra of KPC-producing *K. pneumoniae*, related to the plasmid carrying *bla*_{KPC} (pKpQIL). This peak at 11,109 m/z is clearly detectable in MALDI-TOF mass spectra.

Prerequisite for the automated detection process is the successful ID.

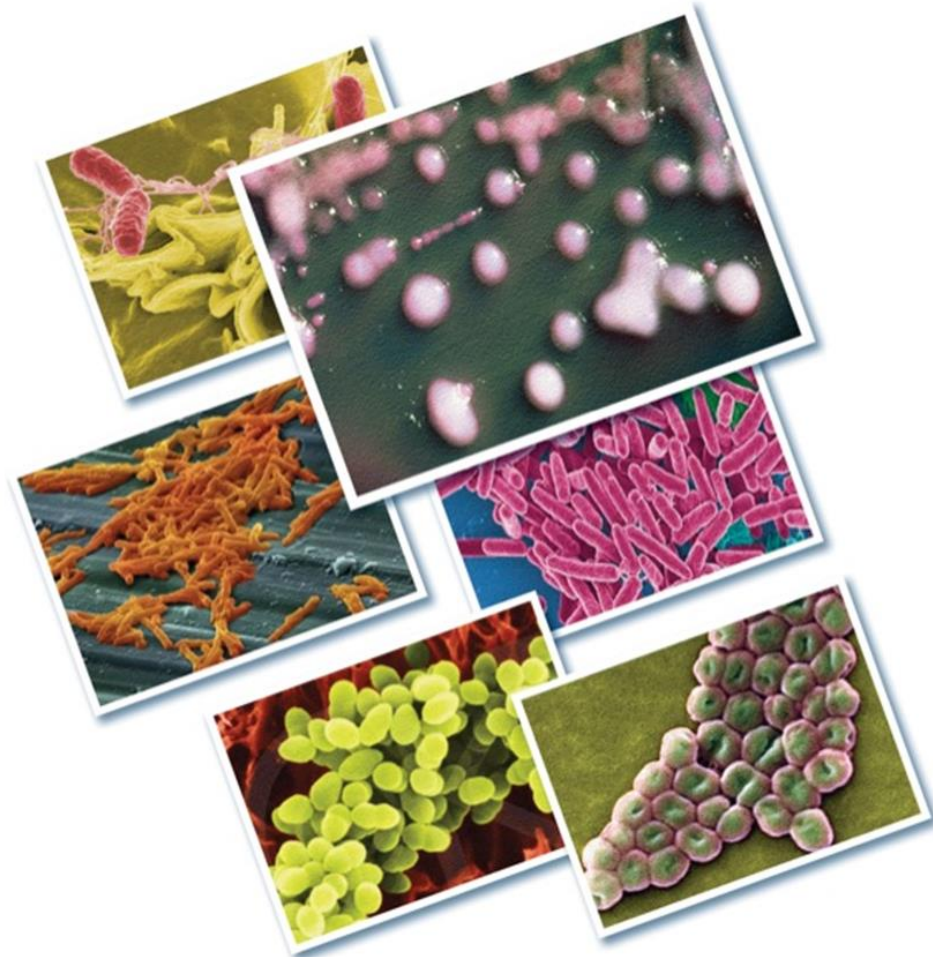
The MBT HT Subtyping IVD Module then looks for the *bla*_{KPC} related peak in the sample spectrum and, if present, the software will report this sample as a KPC positive one.

KPC detection of *K. pneumoniae* and *E. coli* includes only strains with a *bla*_{KPC} pKpQIL plasmid.

If the gene expression rate is low, there will be no characteristic peak and no KPC subtyping alert.

Cultivation media and conditions might suppress or induce a signal at m/z 11,109 which is not related to a KPC resistance. Therefore, Columbia Agar with 5% sheep blood agar must be used for cultivation of *K. pneumoniae* and *E. coli*.

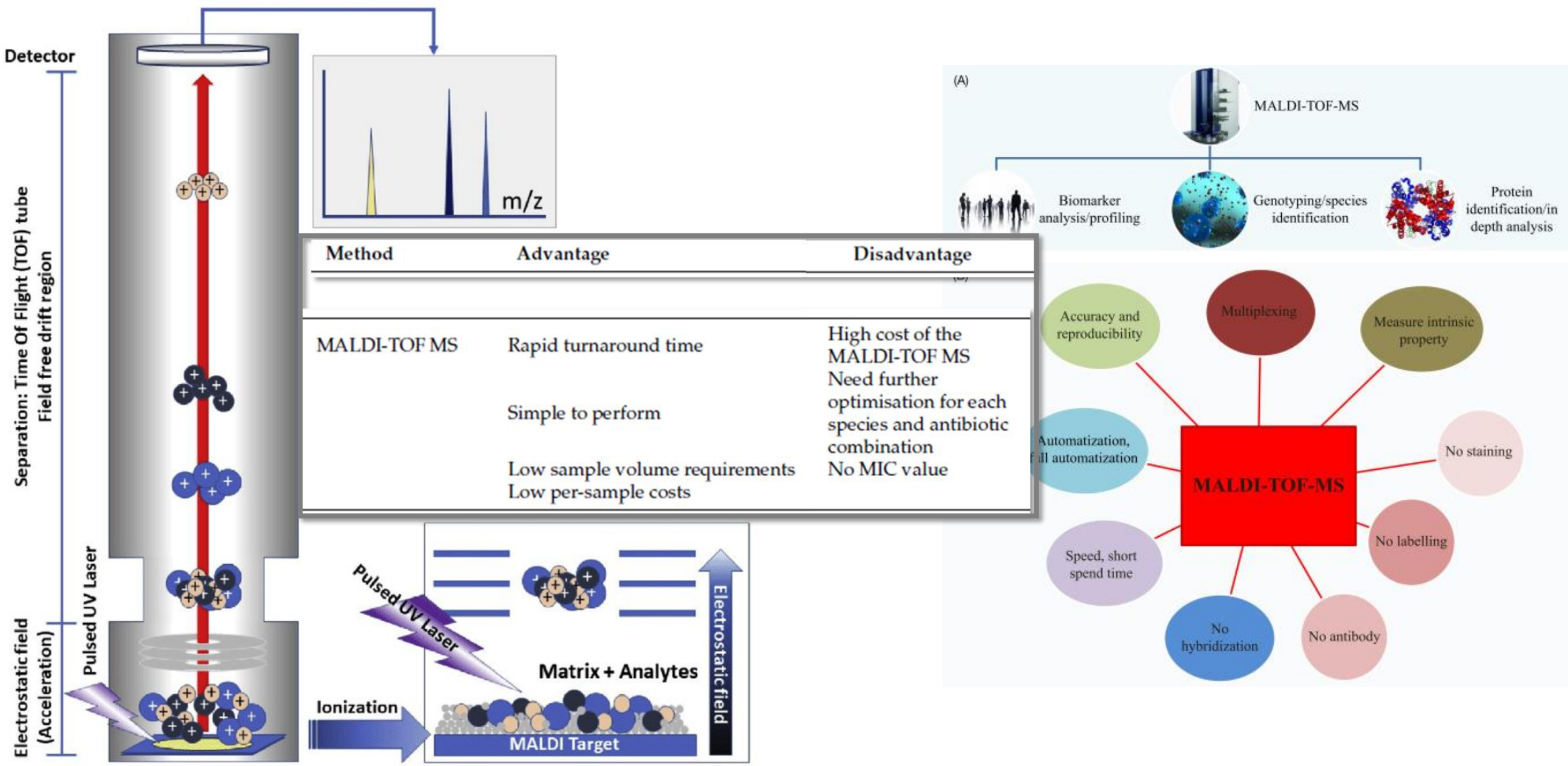
Applications of MALDI-TOF MS for specific AMR detection



Organism	Antibiotic	Year (Reference)
<i>E. coli</i>	Polymyxins	2018 [122]
<i>E. coli</i>	Colistin	2019 [123]
<i>E. coli</i> <i>Klebsiella pneumoniae</i>	Beta-lactams (ESBL-producing isolates)	2019 [124]
<i>Staphylococcus aureus</i> <i>S. intermedius</i> <i>S. pseudintermedius</i>	Novobiocin Polymyxin B Acriflavine	2019 [125]
<i>S. aureus</i>	Methicillin	2019 [126]
<i>Candida auris</i>	Echinocandins	2019 [127]
<i>K. pneumoniae</i> <i>Bacteroides fragilis</i> <i>S. aureus</i>	Carbapenems (carbapenemase-producing isolates) Methicillin	2019 [128]
<i>Enterobacteriaceae</i>	Carbapenems (carbapenemase-producing isolates)	2019 [129]
<i>Enterobacteriaceae</i>	Carbapenem	2019 [130]
<i>Pseudomonas aeruginosa</i>	Beta-lactams (MBL)	2019 [131]
<i>Enterococcus faecium</i>	Vancomycin	2019 [132]
<i>K. pneumoniae</i>	Carbapenems (carbapenemase-producing isolates)	2019 [133]
<i>Acinetobacter baumannii</i>	Colistin	2020 [134]
<i>E. coli</i> <i>K. pneumoniae</i>	Cefotaxime Meropenem, Ciprofloxacin	2020 [135]
<i>S. aureus</i> <i>Enterococcus</i> species <i>E. coli</i> <i>K. pneumoniae</i>	Oxacillin (methicillin) Vancomycin Ceftriaxone Meropenem	2020 [136]
<i>Enterobacterales</i>	Imipenem/Relebactam	2020 [137]
<i>S. aureus</i>	Methicillin	2020 [138]

MALDI-TOF MS approaches for AMR detection in *S. aureus*

MALDI-TOF System Used	Principle of the Method	On Culture/ on Clinical Sample	TAT ¹	Brief Advantages/ Disadvantages +/-	No. and Type of <i>S. aureus</i> Analyzed ²	Major Diagnostic Performance ³	Reference
MALDI Biotyper [®] CA System (Bruker Daltonics)	21 peaks to discriminate MRSA/MSSA	Culture	Few m	+ Use a combination of peaks – Low specificity	181 <i>S. aureus</i>	Sensitivity 87.6% Specificity 71.4%	[64]
MALDI Biotyper [®] CA System (Bruker Daltonics)	Peptide PSM- <i>mec</i> and δ -toxin peaks for MRSA identification	Culture	Few m	+ Sensitive for SCC <i>mec</i> type II, III and VIII – Not adapted for other SCC <i>mec</i> types	35 MRSA	Sensitivity 40%	[65]
MALDI Biotyper [®] CA System (Bruker Daltonics)	Peptide PSM- <i>mec</i> peak for MRSA identification	Culture	Few m	+ Good specificity – SCC <i>mec</i> types of isolates unknown	241 MRSA 106 MSSA	Sensitivity 60.2% Specificity 100%	[66]
MALDI Biotyper [®] CA System (Bruker Daltonics)	m/z 4594 peak for MRSA identification	Culture	Few m	+ Potential MRSA biomarker peak – To be tested in an independent set of data	36 MRSA 31 MSSA	Sensitivity 83.3% Specificity 96.8%	[67]
VITEK MS (BioMérieux)	support vector machine algorithm to identify 38 peaks to discriminate MRSA/MSSA	Culture	Few m	+ Use a combination of peaks – MRSA/MSSA classification model needs optimization	194 MRSA 258 MSSA	Sensitivity 84.0% Specificity 88.0%	[68]
MALDI Biotyper [®] CA System (Bruker Daltonics)	DOT-MGA	Positive blood cultures	6 h	+ Rapid; tested with cefoxitin – Could be optimized for additional antibiotics	30 MRSA 14 MSSA	Sensitivity 100% Specificity 100%	[69]
VITEK MS (BioMérieux)	DOT-MGA	Culture	5 h	+ Tested with oxacillin – Could be optimized for additional antibiotics	20 MRSA 20 MSSA	Sensitivity 100% Specificity 100%	[70]

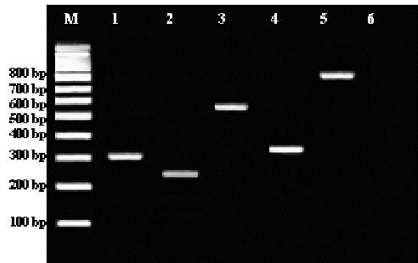
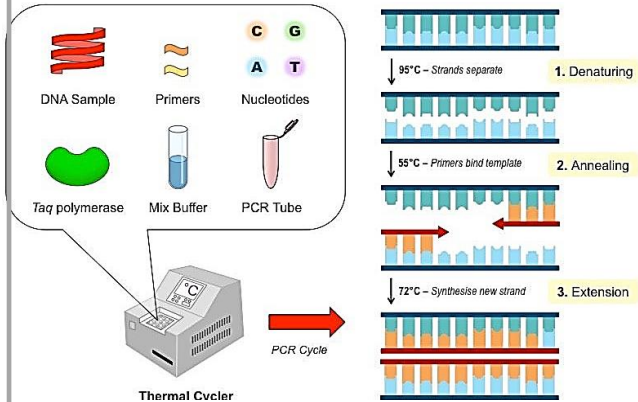




Μοριακές μέθοδοι: από την PCR στο mNGS

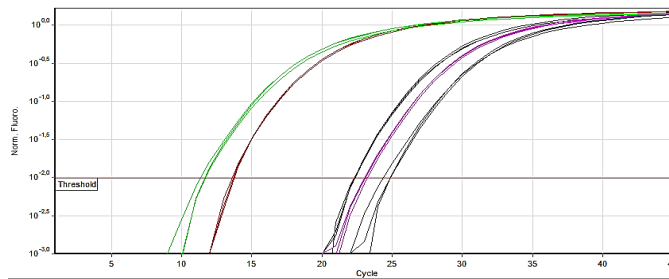
Gene(s) amplification + amplicons detection

Single / Multiplex Endpoint PCR



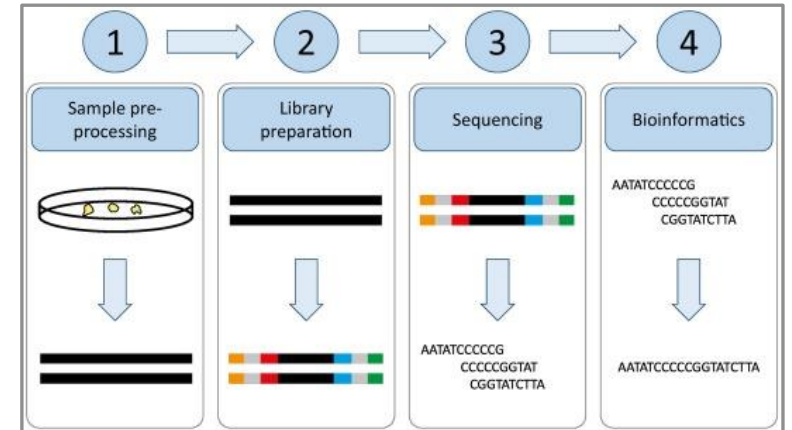
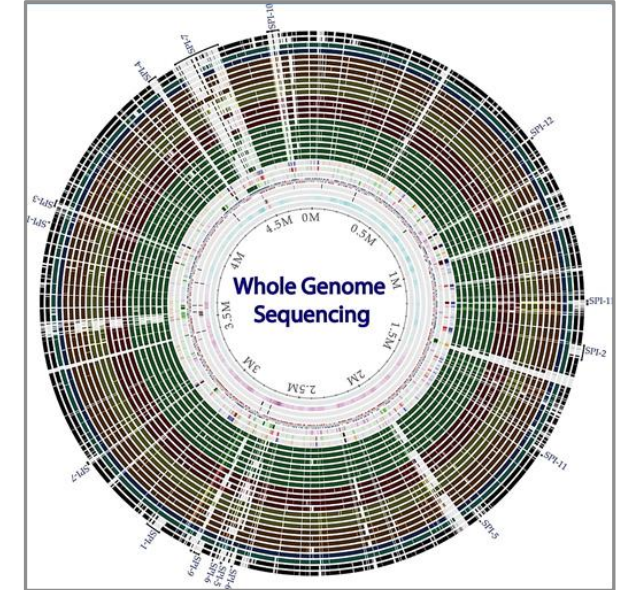
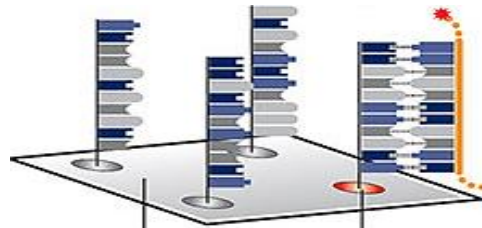
Διαδικασία εργώδης Κίνδυνος επιμολύνσεων

Single / Multiplex Real time PCR



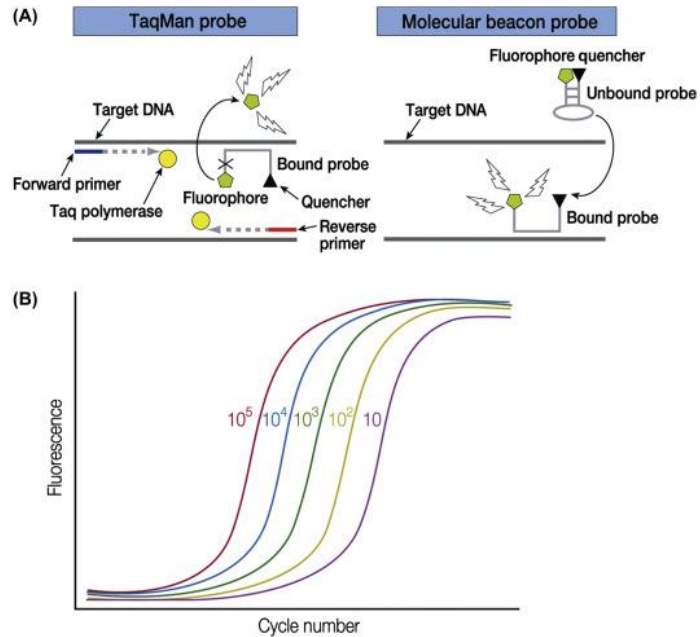
Multiplexing and miniaturization of the hybridization step

PCR / hybridization σε microarray

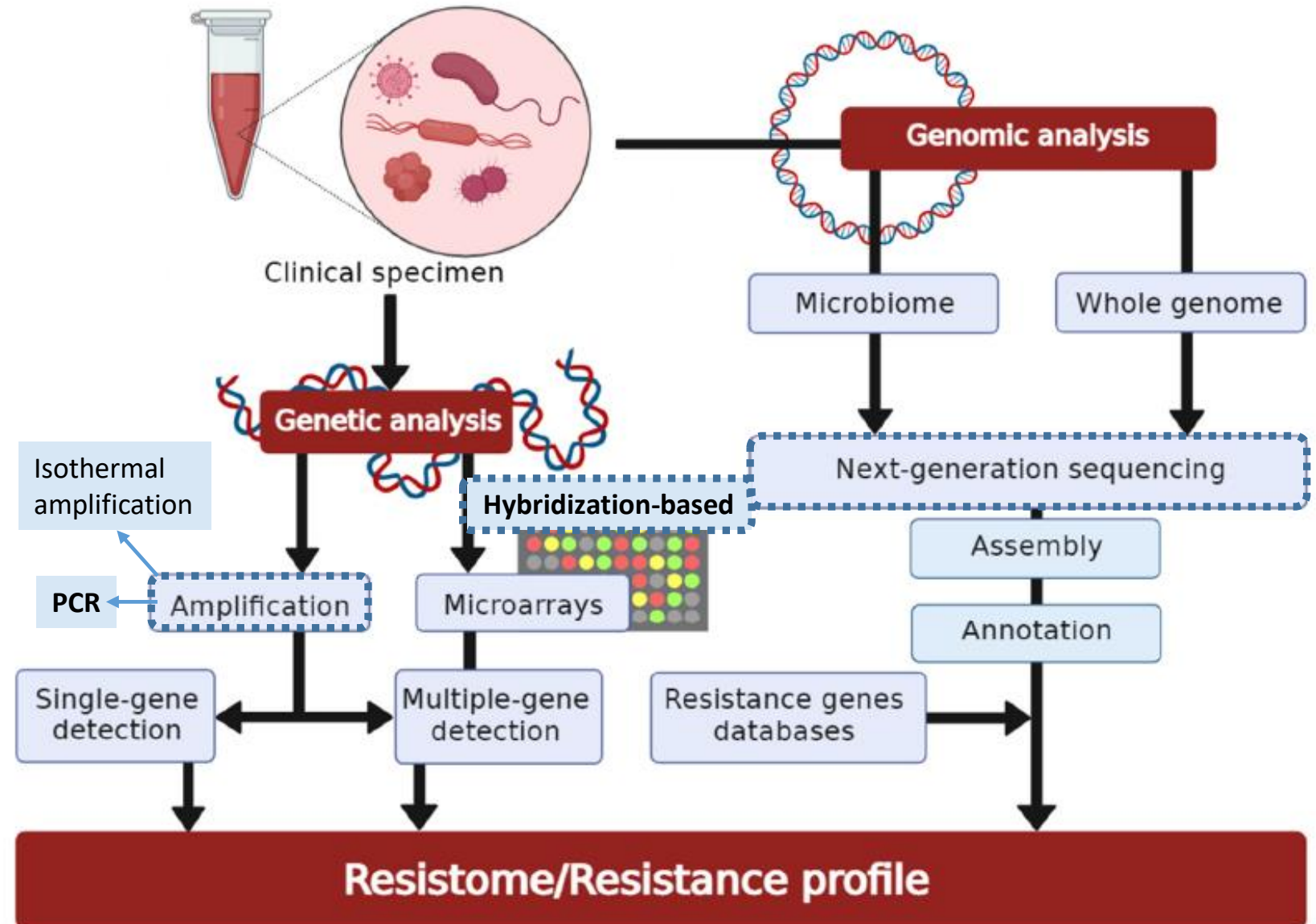


The basic workflow of molecular-based techniques for AMR detection

Real-time PCR (qPCR)



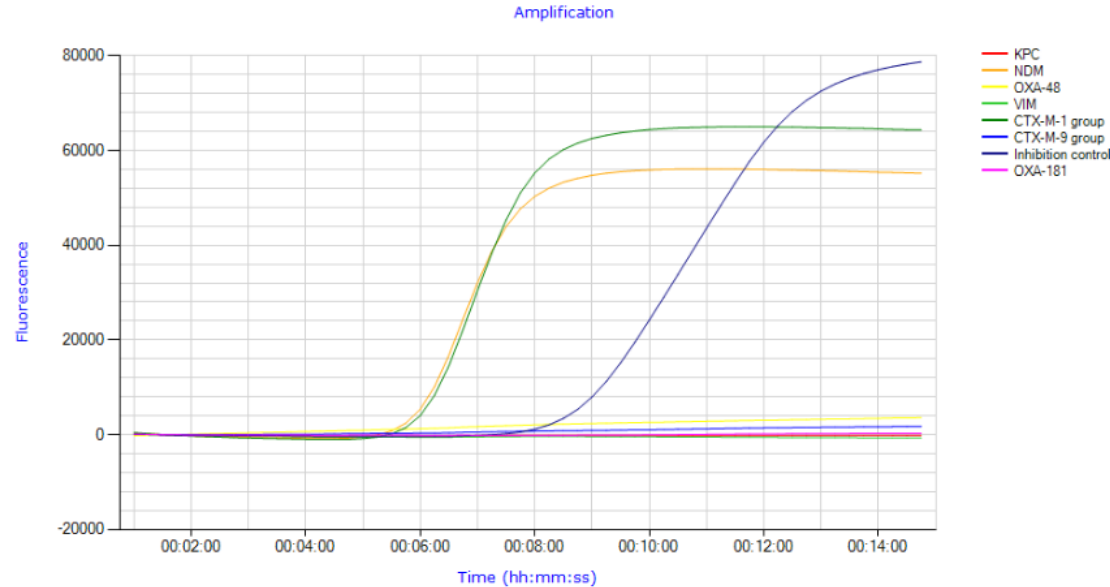
- measurement of data in real-time
- greater sensitivity
- reduced risk of contamination
- greater amenity to multiplexing
- many systems are partially or even completely automated



Ισοθερμικές μοριακές μέθοδοι και ανίχνευση αντοχής: παράδειγμα

Loop-mediated isothermal amplification technology and a real-time PCR platform
No DNA/RNA extraction necessary

eazyplex[®], Amplex



TAT= 20-30 min

Sample: swab, culture

eazyplex[®] SuperBug CRE

KPC, NDM, OXA-48, OXA-181 and VIM
ESBL genes: CTX-M-1 and CTX-M-9 group

TAT= 15-20 min

Direct screening from **rectal swab and urine**
Confirmation from **positive BC and agar plate**

eazyplex SuperBug Acinetobacter

OXA-51, OXA-23-, -40- and -58-like
and NDM
in 15 minutes

	SuperBug complete A	SuperBug complete B	SuperBug complete C
NDM	X	X	X
VIM	X	X	X
KPC	X	X	X
OXA-48	X	X	X
OXA-23	X	X	
OXA-40	X	X	
OXA-58	X		
OXA-181		X	X
IMP			X

Μοριακές μέθοδοι και ανίχνευση αντοχής: παραδείγματα

Xpert® Carba-R, Cepheid

TAT: 48-minutes



Targeted genes

KPC, NDM, VIM, IMP, OXA-48
(covering OXA-181 & OXA-232)

Δείγμα

Ορθικός
στυλεός
Αποικίες

¹Overall percentage agreement with WGS: 92.8%

²Metanalysis of 17 studies involving 15,972 samples
Pooled Se= 95%, Sp= 99%

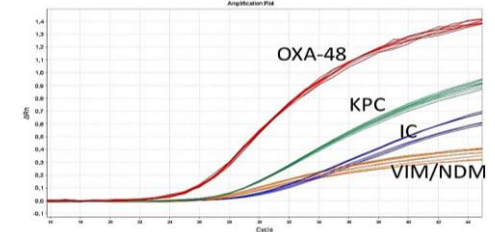
In high-risk populations: Se= 99%, Sp= 98%

1. Khoo BY et al. J Clin Microbiol. 2023;61(9):e0031623.
2. Bai Y et al. J Mol Diagn. 2021;23(11):1534-1544.

Check-Direct CPE® assay, (Checks points)



TAT: 2 h
(+DNA extraction)



Targeted genes

KPC, VIM,
NDM,
OXA-48

Specimen

Rectal swab
Strains

Technology/ instrumentation

Two-step ligation-mediated multiplex real-time PCR amplification – micro array hybridization / non-dedicated platform

867 samples from 627 patients (Reference method: culture + PCR)

CPO assay Se= 95.7%, SP= 96.5%, PPV= 60.8% and NPV= 99.8%

Xpert assay

Se= 97.9%, Sp= 99.8%, PPV= 95.8%, and NPV= 99.9%

The CPO assay: useful when handling many specimens, as tests are conducted in batches.

Positive cases showing high Ct values should be confirmed by another assay to rule out false positivity.

Συμβατική vs. ταχεία διάγνωση

Εμπειρική
αγωγή



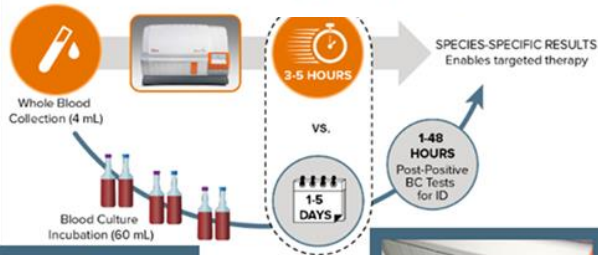
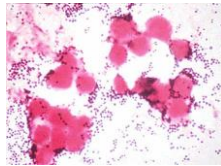
Ολικό αίμα



Θετική
BC



Χρώση Gram



T2Bacteria Panel

Sensitivity: 95%^{1*}
Specificity: 98%^{1*}

E. faecium
S. aureus
K. pneumoniae
P. aeruginosa
E. coli



T2RESISTANCE
PANEL

Gram-negative marker
KPC
OXA-48 Group
NDM /VIM/IMP
CTX-M 14/15
AmpC(CMY/DHA)

Gram-positive marker
vanA/B
mecA/C



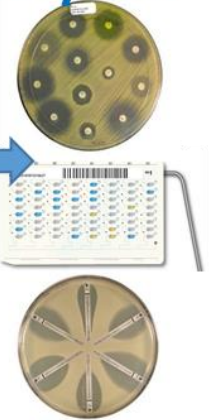
T2Candida Panel

Sensitivity: 91%³
Specificity: 99%³

Candida albicans
Candida tropicalis
Candida krusei
Candida glabrata
Candida parapsilosis

Ροή εργασίας συμβατικών μεθόδων (~48-168 h)

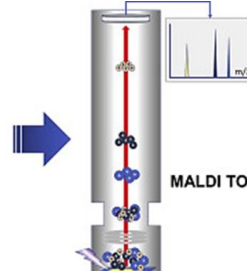
Short subculture → ID /AST



Bacteria Culture



Clinical Specimen



MALDI TOF MS

MALDI Sepsityper

VITEK MS blood culture kit

Antibiotic
Susceptibility
Testing

Microorganism
Identification

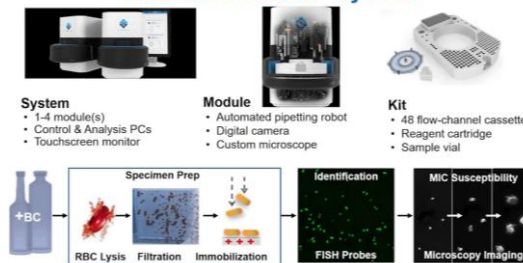
Subtyping

Ροή εργασίας
μοριακών
μεθόδων

Διάγνωση
Στοχευμένη
αγωγή



Accelerate Pheno™ System





Direct MALDI-TOF MS identification from positive blood cultures

ADVANTAGES

LIMITATIONS

Very rapid identification
Easy
Low additional cost

Identification difficulties with some species
Confirmation might be needed
Mixed cultures

MALDI-TOF MS identification from positive blood cultures using short sub-culture on solid medium

ADVANTAGES

LIMITATIONS

Rapid identification
Very easy
Easy to integrate in lab workflow
Confirmation usually not needed

Slow-growing organisms
Mixed cultures

Molecular methods for identification from positive blood cultures

ADVANTAGES

LIMITATIONS

Rapid identification
Easy

High cost
Confirmation needed
Limited pathogen panels

Reduced diagnostic accuracy for polymicrobial cultures

Sensitivity is influenced by:

- the concentration of pathogen-derived nucleic acids
- the presence of PCR inhibitors
- the sample volume

DNA-based identification from whole blood

ADVANTAGES

LIMITATIONS

Time advantage compared to culture
Advantage in pathogen detection under antibiotics

Limited number of pathogens in panel assays
Relatively low sensitivity (only 1-5 ml blood sampled)
Resistance detection ≠ susceptibility testing
Clinical relevance of DNAemia ?
High workload
Highly experienced staff required
High cost
Analytical cut-offs set by some manufacturers
Does not substitute culture-based testing

Ολικό αίμα



Prevalence of Antibiotic-Resistant Pathogens in Culture-Proven Sepsis and Outcomes Associated With Inadequate and Broad-Spectrum Empiric Antibiotic Use

N= 17 430 adults with culture-positive sepsis
admitted to 104 US hospitals
67% received empiric broad-spectrum antibiotics

Is there risk associated with the receipt of empiric broad-spectrum antibiotics?

Resistant GP isolated in 13.6% of patients
Resistant GN in 13.2%
Both undertreatment (failure to cover organisms)
and overtreatment (resistant organisms targeted but not isolated)
were associated with higher mortality after detailed risk adjustment.

Table 2.

Outcomes Associated With Inadequate and Unnecessarily Broad Empiric Antibiotic Therapy^a

Outcome	Inadequate vs adequate empiric therapy						Unnecessarily broad vs not unnecessarily broad empiric therapy ^b					
	No./total No. (%)		Unadjusted OR (95% CI)	P value	Adjusted OR (95% CI)	P value	No./total No. (%)		Unadjusted OR (95% CI)	P value	Adjusted OR (95% CI)	P value
	Inadequate	Adequate empiric therapy					Unnecessarily broad	Not unnecessarily broad				
In-hospital death	488/2785 (17.5)	2011/12 388 (16.3)	1.10 (0.98-1.22)	.09	1.19 (1.03-1.37)	.02	1575/8405 (18.7)	436/3993 (10.9)	1.88 (1.68-2.11)	<.001	1.22 (1.06-1.40)	.007
Hospital-onset acute kidney injury	486/2785 (17.5)	2196/12 398 (17.7)	0.98 (0.88-1.09)	.74	1.02 (0.90-1.16)	.72	1641/8405 (19.5)	555/3993 (13.9)	1.50 (1.35-1.67)	<.001	1.12 (1.00-1.26)	.05
<i>Clostridioides difficile</i>	207/2785 (7.4)	498/12 398 (4.0)	1.92 (1.63-2.27)	<.001	1.19 (0.98-1.45)	.09	367/8405 (4.4)	131/3993 (3.3)	1.34 (1.10-1.65)	.004	1.26 (1.01-1.57)	.04

These findings underscore the need for better tests to rapidly identify patients with resistant pathogens

FilmArray BCID2: 43 Targets, TAT 1 h

GRAM-NEGATIVE BACTERIA

Acinetobacter baumannii complex

Bacteroides fragilis

Enterobacterales

Enterobacter cloacae complex

Escherichia coli

Klebsiella aerogenes

Klebsiella oxytoca

Klebsiella pneumoniae group

Proteus spp.

Salmonella spp.

Serratia marcescens

Haemophilus influenzae

Neisseria meningitidis

Pseudomonas aeruginosa

Stenotrophomonas maltophilia

GRAM-POSITIVE BACTERIA

•*Enterococcus faecalis*

•*Enterococcus faecium*

•*Listeria monocytogenes*

•*Staphylococcus* spp.

Staphylococcus aureus

Staphylococcus epidermidis

Staphylococcus lugdunensis

•*Streptococcus* spp.

Streptococcus agalactiae

Streptococcus pneumoniae

Streptococcus pyogenes

YEAST:

•*Candida albicans*

•*Candida auris*

•*Candida glabrata*

•*Candida krusei*

•*Candida parapsilosis*

•*Candida tropicalis*

•*Cryptococcus*

(*C. neoformans*/*C. gattii*)

AMR GENES

Carbapenemases

IMP

KPC

OXA-48-like

NDM

VIM

Colistin Resistance: *mcr-1*

ESBL: CTX-M

Methicillin Resistance

mecA/C

mecA/C and *MREJ*

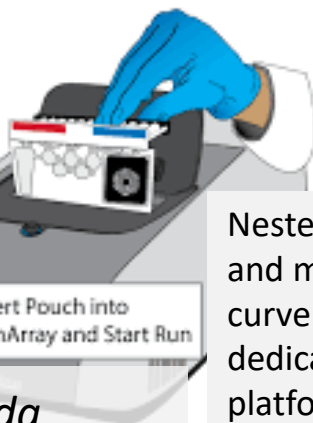
(MRSA)

Vancomycin Resistance:

vanA/B



5 Inject Sample into Pouch



6 Insert Pouch into FilmArray and Start Run

Nested-PCR and melting curve analysis dedicated platform

ePlex® BCID Panels



*Se 97-99%

Sp 96-99%

*Huang TD et al.

J Clin Microbiol. 2019;57(2).

Gram-positive Organisms

21 παθογόνα
6 γονίδια αντοχής
Gram-Negative Organisms

20 παθογόνα
4 γονίδια αντοχής

15
παθογόνα

Fungal Organisms

VERIGENE® Bloodstream Infection Testing Panels

Gram-Positive Blood Culture Test (BC-GP)

Species

Staphylococcus aureus
Staphylococcus epidermidis
Staphylococcus lugdunensis
Streptococcus agalactiae
Streptococcus pneumoniae
Streptococcus pyogenes
Enterococcus faecalis
Enterococcus faecium
Streptococcus anginosus

Genus

Staphylococcus spp.
Streptococcus spp.
Micrococcus spp.*
Listeria spp.

Resistance

mecA (methicillin)
vanA (vancomycin)
vanB (vancomycin)

Group

Gram-Negative Blood Culture Test (BC-GN)

Species

*Escherichia coli**
Klebsiella pneumoniae
Klebsiella oxytoca
Pseudomonas aeruginosa
*Serratia marcescens***

Resistance

CTX-M (ESBL)
IMP (carbapenemase)
KPC (carbapenemase)
NDM (carbapenemase)
OXA (carbapenemase)
VIM (carbapenemase)

Genus

Acinetobacter spp.
Citrobacter spp.
Enterobacter spp.
Proteus spp.

Se: 98.1% GN, 97.3% GP, 99.2% *Candida*

Sp: 99.9% GN, 99.8% GP, 99.9% *Candida*

Se/Sp για AMR γονίδια: 98.4-100/98.3-100%

Salimnia H et al J Clin Microbiol 2016.

Micro-array hybridization
dedicated platform



TAT 2-2.5 h



Συνδρομική διάγνωση και ανίχνευση γονιδίων αντοχής: Blood Culture Panels (positive BC bottle)

Parameter	FilmArray BCID	Verigene	
		Gram-positive blood culture	Gram-negative blood culture
Total no. of targets	27	15	14
Ability to detect presence of resistance gene			
<i>mecA</i>	✓	Se 93.9%	✓
<i>vanA</i>	✓	Se: 90.9	✓
<i>vanB</i>	✓		✓
<i>bla</i> _{KPC}	✓		
<i>bla</i> _{NDM}	✓		
<i>bla</i> _{OXA}	✓		
<i>bla</i> _{VIM}	✓		
<i>bla</i> _{IMP}	✓		
<i>bla</i> _{CTX-M}	✓		
<i>mcr-1</i>	✓		
Time to result (h)	~1	~2.5	~2

Sp > 98 %

Se 93.9%

Se: 90.9

NPA >99.9%

Se: 94.9%

PPA 100%

PPA 96.2%

PPA 94.3%

PPA 100%

PPA 100%

PPA 98.9%

ePlex BCID, GenMark



Resistance
Genes

CTX-M
KPC
NDM
VIM
IMP
OXA

Resistance
Genes

mecA
mecC
vanA
vanB

Multiplexed NAAT and a
electrochemical detection technology
with a dedicated platform

Peri AM et al. BMC Infect Dis. 2022;22(1):794.

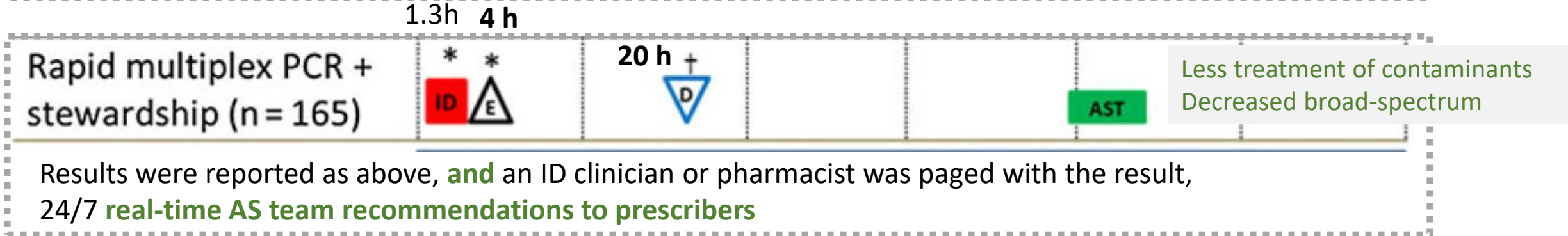
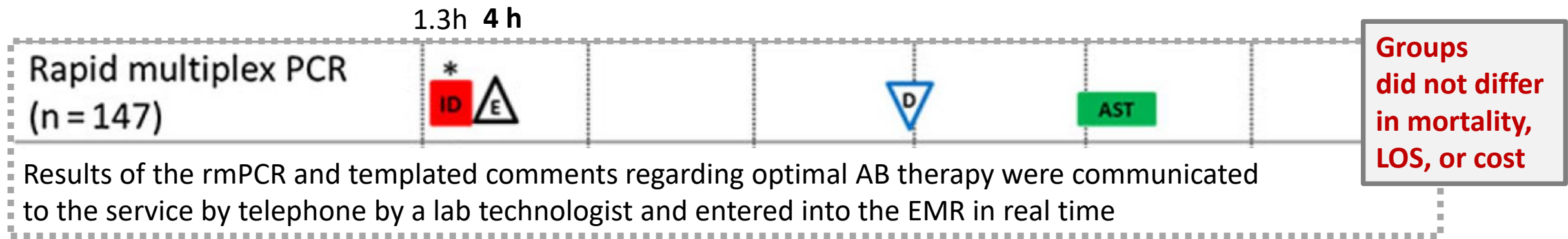
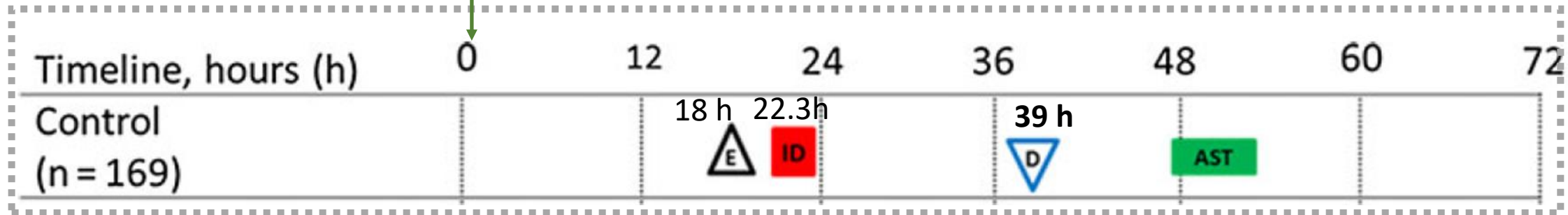
Ramanan P et al. Clin Microbiol Rev. 2017;31(1) (με επικαιροποίηση).

Salimnia H et al. J Clin Microbiol. 2016;54(3):687-98.

Ledeboer NA et al. J Clin Microbiol. 2015;53(8):2460-72.

Prospective RCT, 617 patients with BSI

Gram result reported



ID Organism identification
 AST Phenotypic antimicrobial susceptibility report
 D De-escalation
 E Escalation

2022;75(12):2066-2075.

Impact of a Rapid Molecular Test for *Klebsiella pneumoniae* Carbapenemase and Ceftazidime-Avibactam Use on Outcomes After Bacteremia Caused by Carbapenem-Resistant Enterobacterales

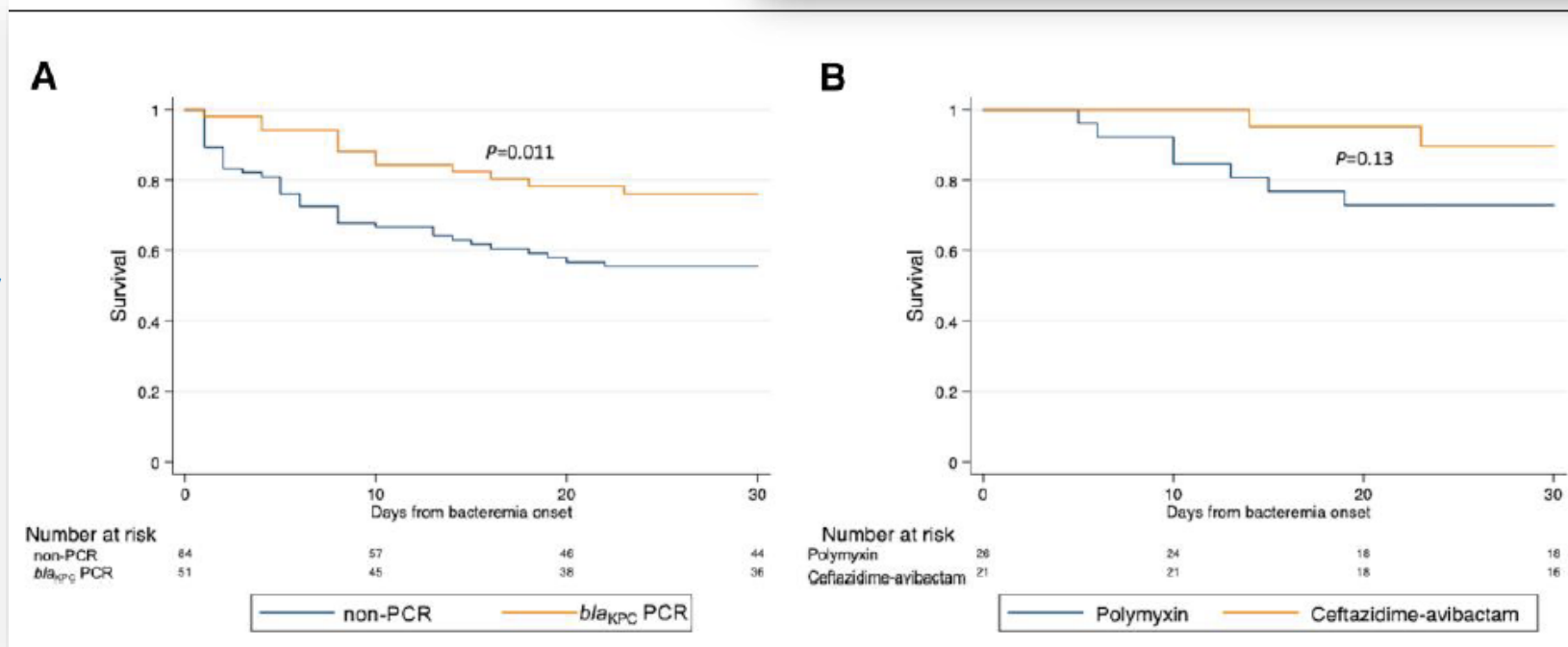
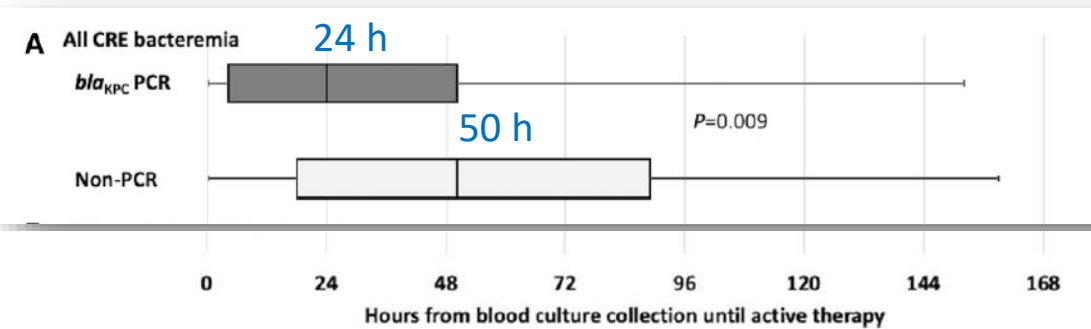
Michael J. Satlin,^{1,2} Liang Chen,^{3,4} Angela Gomez-Simmonds,⁵ Jamie Marino,² Gregory Weston,⁶ Tanaya Bhowmick,⁷ Susan K. Seo,⁸ Steven J. Sperber,^{9,10} Angela C. Kim,¹¹ Brandon Eilertson,¹² Sierra Derti,¹ Stephen G. Jenkins,^{1,2} Michael H. Levi,¹³ Melvin P. Weinstein,^{7,14} Yi-Wei Tang,¹⁵ Tao Hong,¹⁶ Stefan Juretschko,¹⁷ Katherine L. Hoffman,¹⁸ Thomas J. Walsh,¹ Lars F. Westblade,^{1,2} Anne-Catrin Uhlemann,⁹ and Barry N. Kreiswirth^{3,4}

N= 137 patients with CRE bacteremia

[77% CP-CRE (65% KPC)]

2016-2018, 8 New York and New Jersey medical centers

3/8 study hospitals used the FilmArray BCID



30-day mortality
24% vs 47%;
 $P=.007$

30-day
mortality 10% with
caz-avi monotherapy
vs. 31% with
polymyxin
monotherapy
($P=.08$)

Impact of an Antimicrobial Stewardship Program Intervention Associated with the Rapid Identification of Microorganisms by MALDI-TOF and Detection of Resistance Genes in ICU Patients with Gram-Negative Bacteremia

216 episodes of GN bacteremia, ICU

MALDI-TOF και ανίχνευση των γονιδίων AMR απευθείας από τη φιάλη της θετικής αιμοκ/ας

Table 4. Antimicrobial consumption in DOT per 1000 days, present and collection (21 days).

Antimicrobial Consumption and Cost	Pre-Intervention	Intervention	p-Value
Consumption (DOT/1000 days present, median, IQR)			
All antimicrobials	N = 114 1.381 (1.103–2.251)	N = 102 1.262 (1.063–1.662)	0.032 ^a
Antimicrobial for gram-negative bacteria	N = 114 1.281 (1.004–1.775)	N = 102 1.172 (1.006–1.427)	0.067 ^a
Antimicrobial for gram-positive bacteria	N = 52 475 (238–761)	N = 43 270 (139–467)	0.004 ^a
Carbapenems	N = 76 836 (504–1056)	N = 72 543 (301–991)	0.040 ^a
Colistin/Polymyxin B	N = 45 722 (390–932)	N = 40 881 (462–1001)	0.299 ^a
Ceftazidime/avibactam	N = 3 931 (725–1022)	N = 8 911 (823–940)	0.865 ^a

Καμιά επίδραση στη θνητότητα 30-ημερών
Σημαντικά μικρότερη η συνολική διάρκεια νοσηλείας (44 vs. 39 ημέρες, $p = 0.005$) και ο χρόνος νοσηλείας στη ΜΕΘ (17 vs. 13 ημέρες, $p = 0.033$)



Whole Blood



EXPERT REVIEW OF MEDICAL DEVICES
2021, VOL. 18, NO. 5, 473–482
<https://doi.org/10.1080/17434440.2021.1919508>



META-ANALYSIS

OPEN ACCESS

Antimicrobial and resource utilization with T2 magnetic resonance for rapid diagnosis of bloodstream infections: systematic review with meta-analysis of controlled studies

Blood Culture Incubation (60 mL)

61.4% των αιτίων
μικροβιαμίας



T2BACTERIA PANEL

Sensitivity: 90%⁷
Specificity: 98%⁷

E. faecium
S. aureus
K. pneumoniae
P. aeruginosa
E. coli



T2RESISTANCE PANEL

Gram-negative marker
KPC
OXA-48 Group
NDM /VIM/IMP
CTX-M 14/15
AmpC(CMY/DHA)

Gram-positive marker
vanA/B
mecA/C



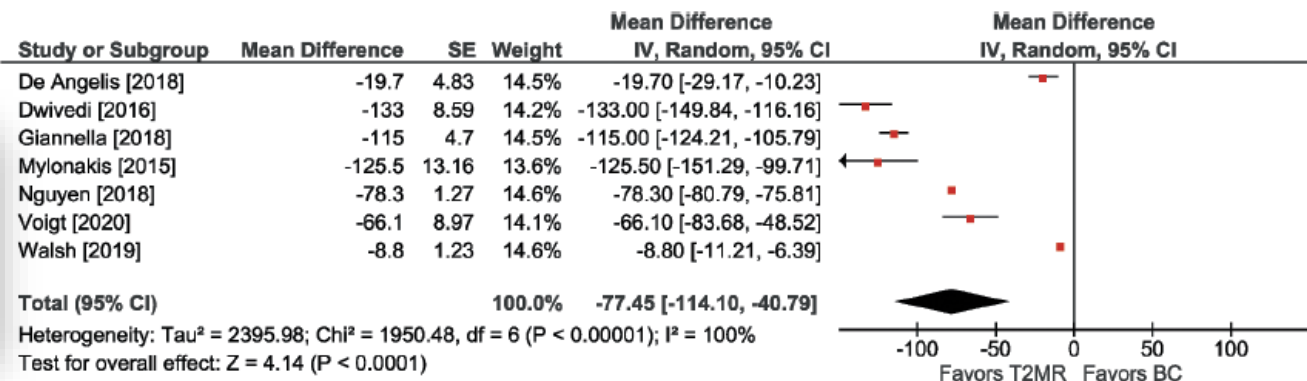
T2CANDIDA PANEL

Sensitivity: 91%⁵
Specificity: 99%⁵

Candida albicans
Candida tropicalis
Candida krusei
Candida glabrata
Candida parapsilosis

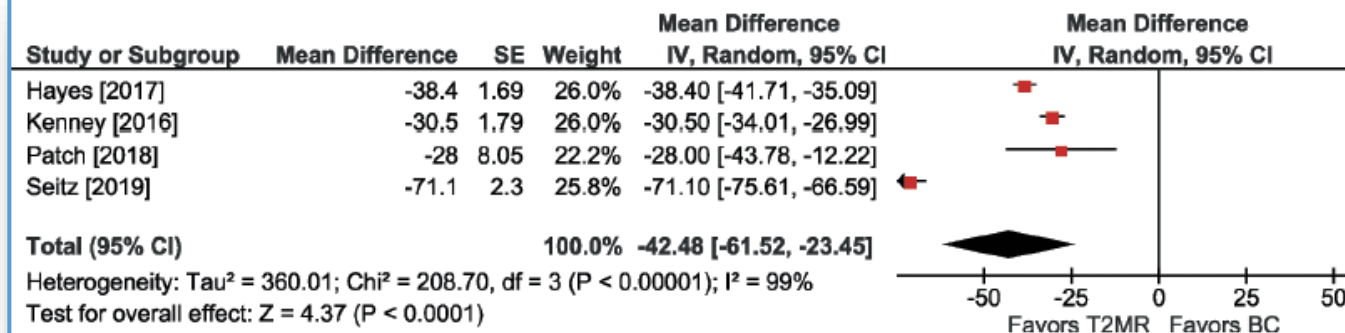
1-3 CFU/mL LoD

Hours to species identification with T2MR vs. BC



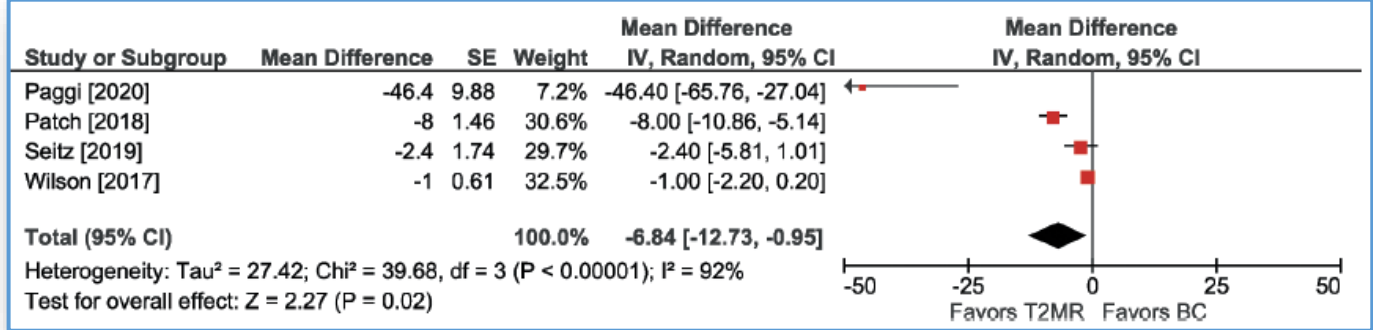
Pooled mean difference = -77 hours ($p < 0.001$)

Hours to targeted therapy among T2MR positive cases vs. BC



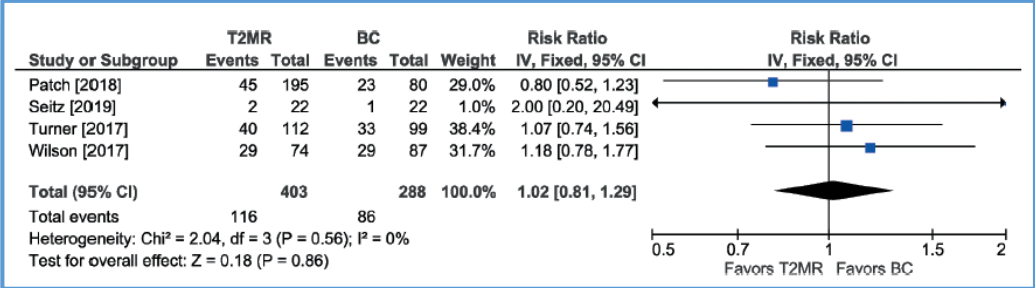
Pooled mean difference = -42 hours ($p < 0.001$)

Hours to empirical therapy de-escalation among T2MR negative cases vs. BC



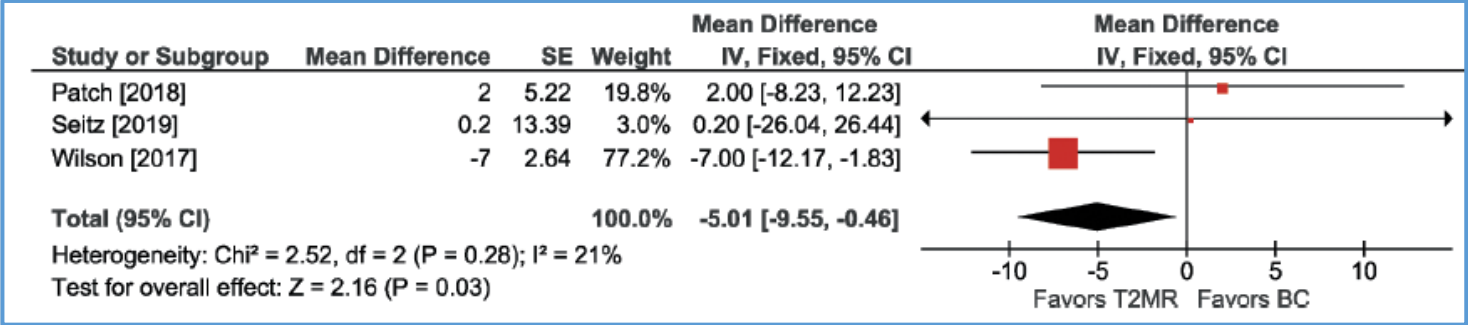
Pooled mean difference = -7 hours (p = 0.02).

Mortality with T2MR vs. BC

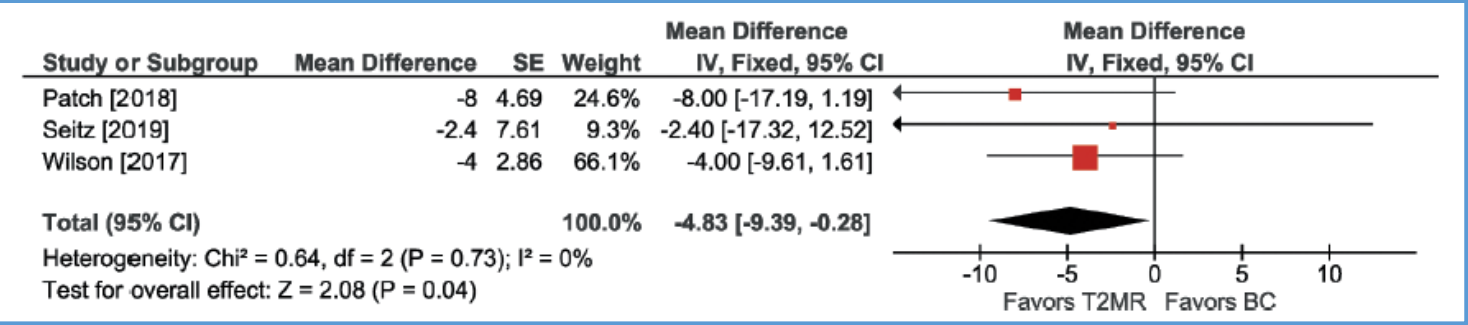


28.9% vs. 29.9%, RR = 1.02, p = 0.86

ICU stay with T2MR vs. BC. Pooled mean difference = -5.0 days (p =0.03)



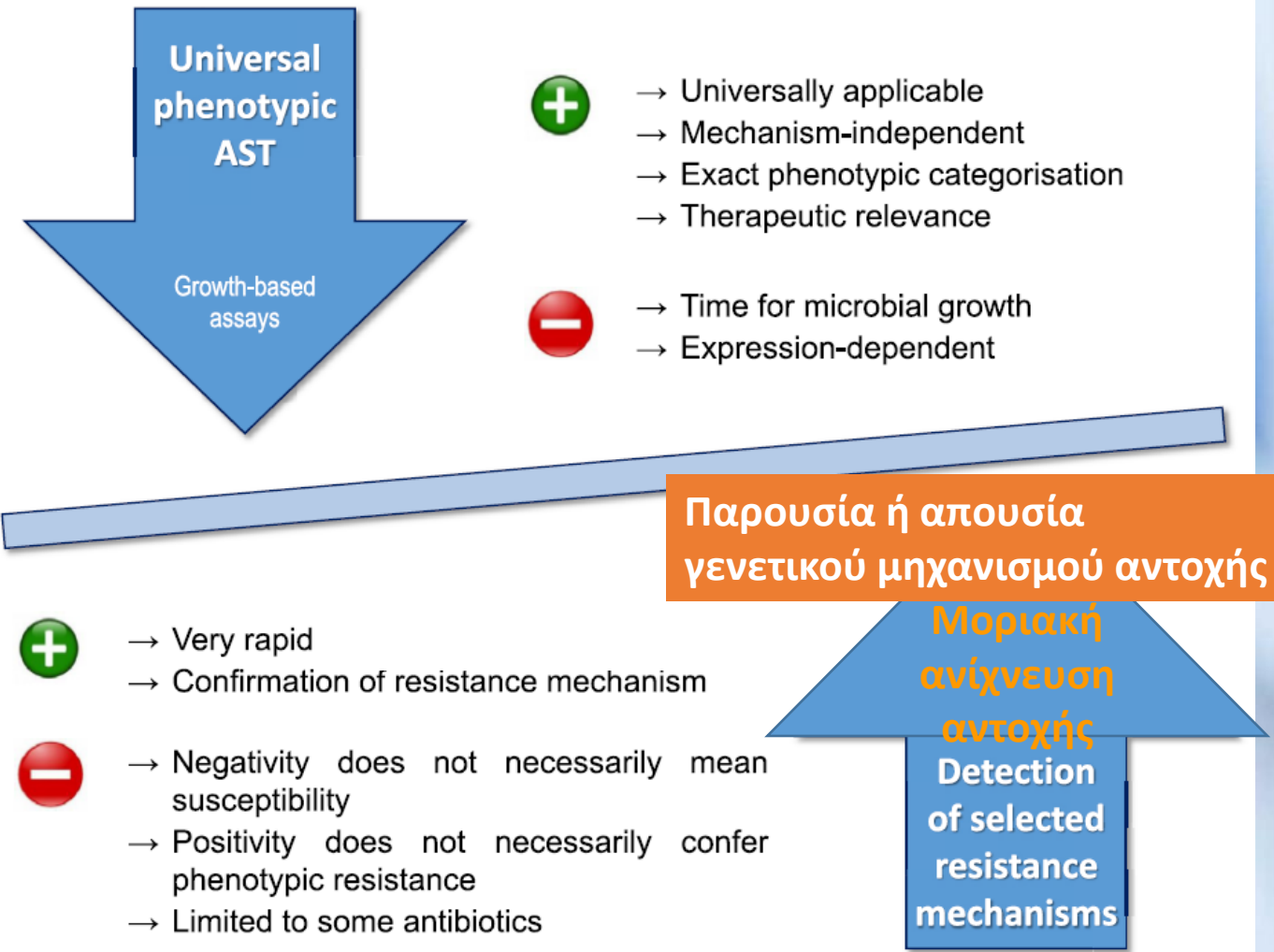
Hospital stay with T2MR vs. BC. Pooled mean difference = -4.8 days (p =0.04)



Considering hospital costs for sepsis patients range from 2,000 USD to 5,000 USD per day depending on sepsis severity, implementation of T2MR could theoretically reduce hospital costs by 10,000 USD to 25,000 USD per patient tested

Μοριακή ανίχνευση αντοχής: προβληματισμοί

Detection of particular resistance mechanisms vs. phenotypic AST



-
- Πόσα και ποια γονίδια περιλαμβάνονται στα panel
 - Επιλογή γονιδίων σε σχέση με την τοπική επιδημιολογία τη δεδομένη χρονική στιγμή
 - Μη αναγνώριση κάποιων variants ή αναγνώριση variants με διαφορετικό υδρολυτικό προφίλ
 - Μη ανίχνευση νέων αναδυόμενων γονιδίων αντοχής
 - **Σύνθετοι μηχανισμοί αντοχής**
 - **Πολυμικροβιακά δείγματα και γονίδια αντοχής**

Είναι ασφαλής η αποκλιμάκωση;



An Antibiotic Stewardship Program Blueprint for Optimizing Verigene BC-GN within an Institution: a Tale of Two Cities

Δεν υπάρχει σαφής κατεύθυνση για τις περιπτώσεις που δεν ανιχνεύονται καθοριστές αντοχής

1046 GN bloodstream isolates

TABLE 2 Verigene BC-GN analytic results by bug-drug combination^a

Institution and target organism	n	No. (%) of isolates	No. of isolates	P (%)	PPV (%)	NPV (%)
Detroit Medical Center						
<i>E. coli</i>	38			99	97	98
<i>K. pneumoniae</i>	14			9	97	94
	14			00	100	99
<i>K. oxytoca</i>	2			00	100	95
<i>Proteus</i> spp.	5			00	100	94
<i>Enterobacter</i> spp.	6			00	100	97
<i>Citrobacter</i> spp.	10					100
<i>Acinetobacter</i> spp.	39			3	80	93
<i>Pseudomonas</i> spp.	5					88



The Clinical Impact of a Negative Molecular β -Lactamase Gene Test for *Enterobacteriaceae*: Let's Not Let Perfect Be the Enemy of Really Good

Jason M. Pogue,^a Emily L. Heil^b

Με εξαίρεση την *P. aeruginosa*, η απουσία καθοριστών αντοχής μπορούσε να προβλέψει την ευαισθησία στα αντιβιοτικά στόχους



ID and phenotypic AST using the Accelerate Pheno System (RAPID)

RCT

Patients (N= 448)

with positive BC with GNB

RANDOMIZED

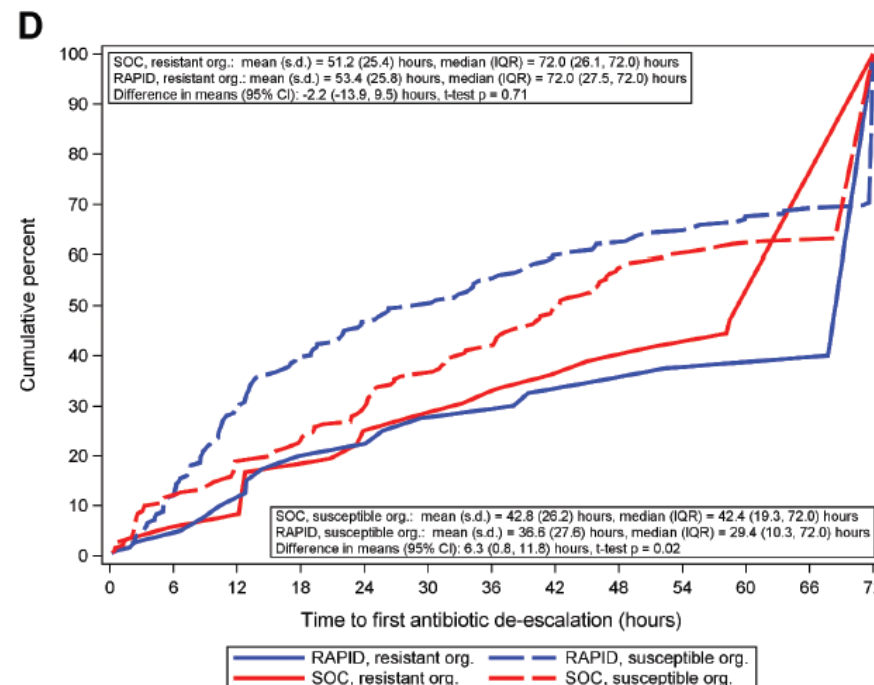
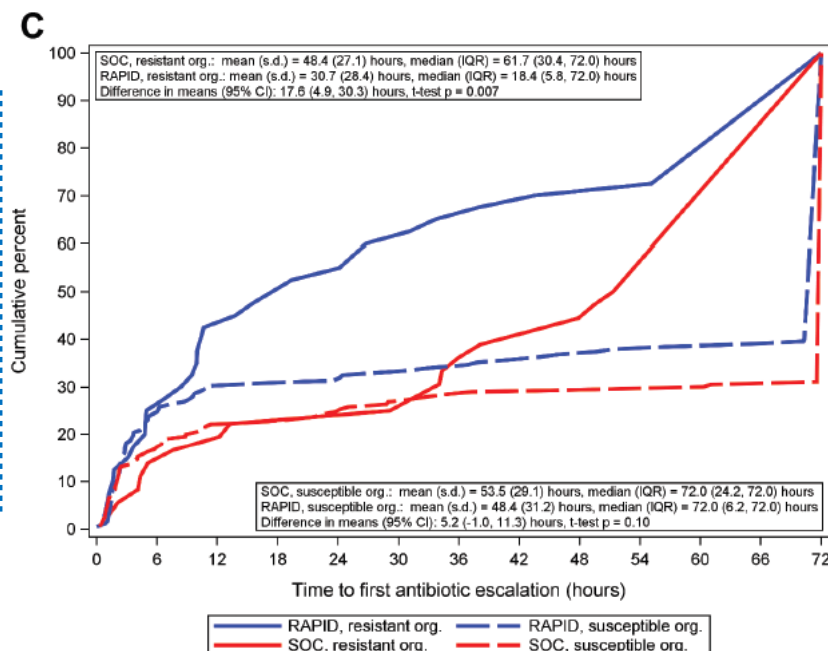
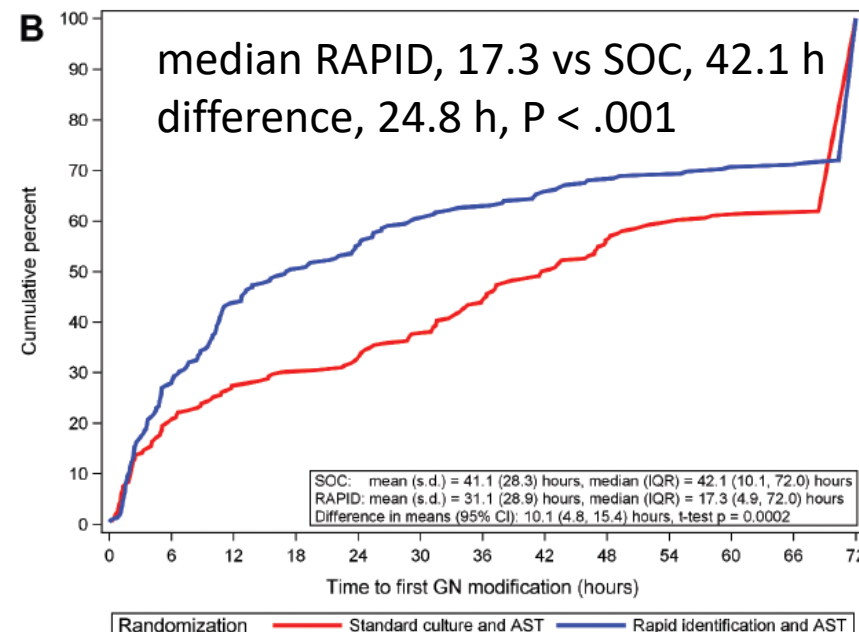
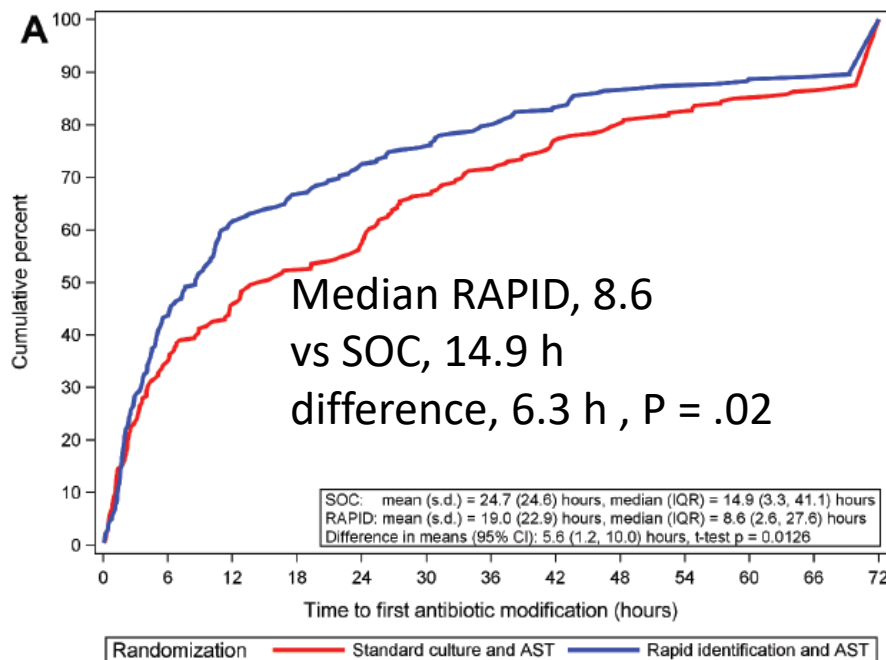
SOC testing with AS review
or RAPID with AS

Patients with MDRO, escalation

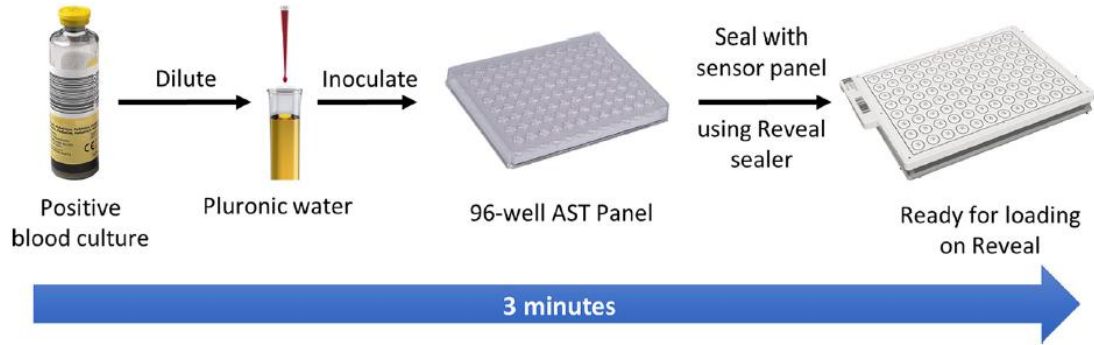
Median RAPID, 18.4 vs SOC, 61.7 h
(difference, 43.3 h; $P = .01$)

Patients with S organisms, de-escalation

median RAPID, 29.4 vs SOC, 42.4 h
(difference, 13 h, $P = .02$)



Rapid AST



RAPID AST SYSTEM

5.5 hours



TABLE 1 Overall performance of the Reveal AST system

Parameter or detail	Performance of Reveal AST against:	
	Sensititre	Vitek 2
Parameter, % (no. positive/total no.)		
EA ^c	98.0 (2,129/2,173)	97.0 (1,482/1,528)
CA ^c	96.3 (2,174/2,258)	96.2 (1,554/1,615)
mE	3.5 (78/2,258)	3.3 (54/1,615)
ME	0.3 (5/1,889)	0.3 (4/1,342)
VME	1.3 (4/313)	1.3 (3/232)

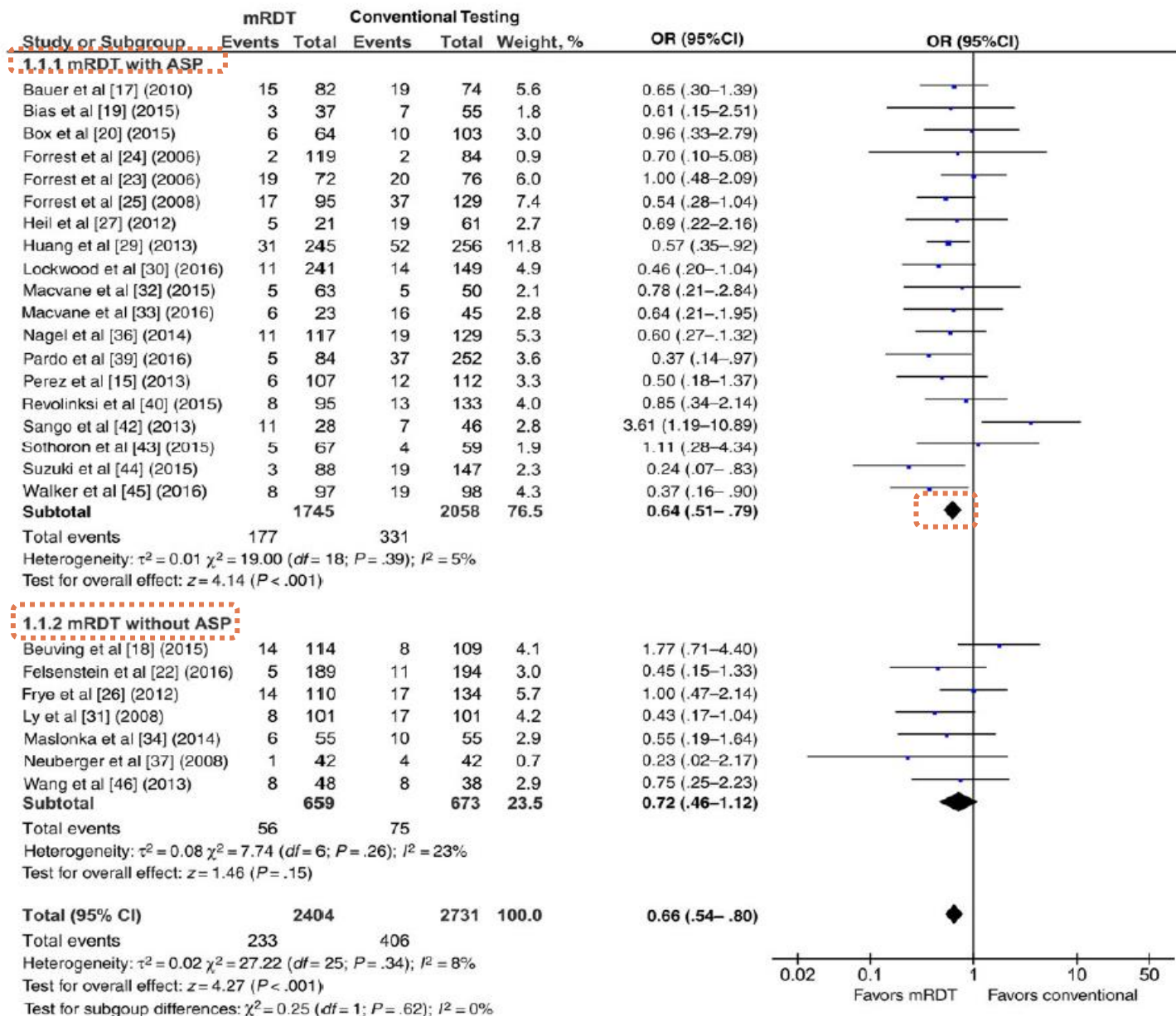
Wide antimicrobial coverage for Gram-negative bloodstream infections, with 176 bug drug combinations.

ANTIMICROBIAL DRUG	MIC CALLING RANGE (µg/ml)	A. baumannii	C. freundii	C. koseri	E. cloacae	E. coli	K. aerogenes	K. oxytoca	K. pneumoniae	P. mirabilis	P. aeruginosa	
Amikacin	4 - 16	✓	✓	✓	✓	✓	✓	✓	✓	✓	10	
Amoxicillin / Clavulanate	4/2 - 16/2			✓		✓		✓	✓		5	
Ampicillin	4 - 8				✓				✓		2	
Aztreonam	1 - 16		✓	✓	✓	✓		✓		✓	8	
Cefepime	0.125 - 64		✓	✓	✓	✓		✓	✓	✓	8	
Cefotaxime	0.125 - 4		✓	✓	✓	✓		✓	✓		8	
Cefoxitin	8 - 16			✓		✓		✓	✓		5	
Certazidime	0.125 - 64		✓	✓	✓	✓		✓	✓	✓	9	
Certazidime / Avibactam	0.25/4, 1/4 - 16/4		✓	✓	✓	✓		✓	✓	✓	9	
Certazidime / Clavulanate	0.25/4 - 8/4				✓			✓			3	
Ceftolozane / Tazobactam	1/4 - 4/4				✓	✓	✓	✓		✓	6	
Ciprofloxacin	0.06, 0.25 - 1	✓	✓	✓	✓	✓	✓	✓	✓	✓	10	
Ertapenem	0.125 - 1		✓	✓	✓	✓	✓	✓	✓		8	
Gentamicin	2 - 4	✓	✓	✓	✓	✓	✓	✓	✓		9	
Imipenem	1 - 8	✓	✓	✓	✓	✓	✓	✓		✓	9	
Levofloxacin	0.25 - 1	✓	✓	✓	✓	✓	✓	✓	✓	✓	10	
Meropenem	0.125 - 8	✓	✓	✓	✓	✓	✓	✓	✓	✓	10	
Meropenem / Vaborbactam	2/8 - 8/8		✓	✓	✓	✓	✓	✓	✓	✓	9	
Piperacillin	8 - 16		✓		✓	✓	✓	✓	✓	✓	8	
Piperacillin / Tazobactam	4/4 - 16/4		✓	✓	✓	✓	✓	✓	✓	✓	9	
Tigecycline	0.5 - 1			✓		✓					2	
Tobramycin	2 - 4	✓	✓	✓	✓	✓	✓	✓	✓	✓	10	
Trimethoprim / Sulfamethoxazole	2/38 - 4/76	✓	✓	✓	✓	✓	✓	✓	✓		9	
TOTAL		8	17	19	18	23	17	21	21	18	14	176

System reports results with S/I/R interpretation according to CA-SFM 2019 and EUCAST 2021 Breakpoints

Mortality outcomes with molecular rapid diagnostic testing (mRDT) versus conventional testing in bloodstream infection

mRDT και ASP



31 μελέτες, 5920 BSI pts

Mortality risk decreased significantly with mRDT in the presence of ASP

Number needed to treat: 20 patients with BSI to prevent 1 death within 30 days

Time to effective therapy decreased by a weighted mean difference of –5.03 hours

Length of stay decreased by –2.48 days



The Cost-Effectiveness of Rapid Diagnostic Testing for the Diagnosis of Bloodstream Infections with or without Antimicrobial Stewardship

Elina Eleftheria Pliakos,^a Nikolaos Andreatos,^a Fadi Shehadeh,^a Panayiotis D. Ziakas,^a Eleftherios Mylonakis^a

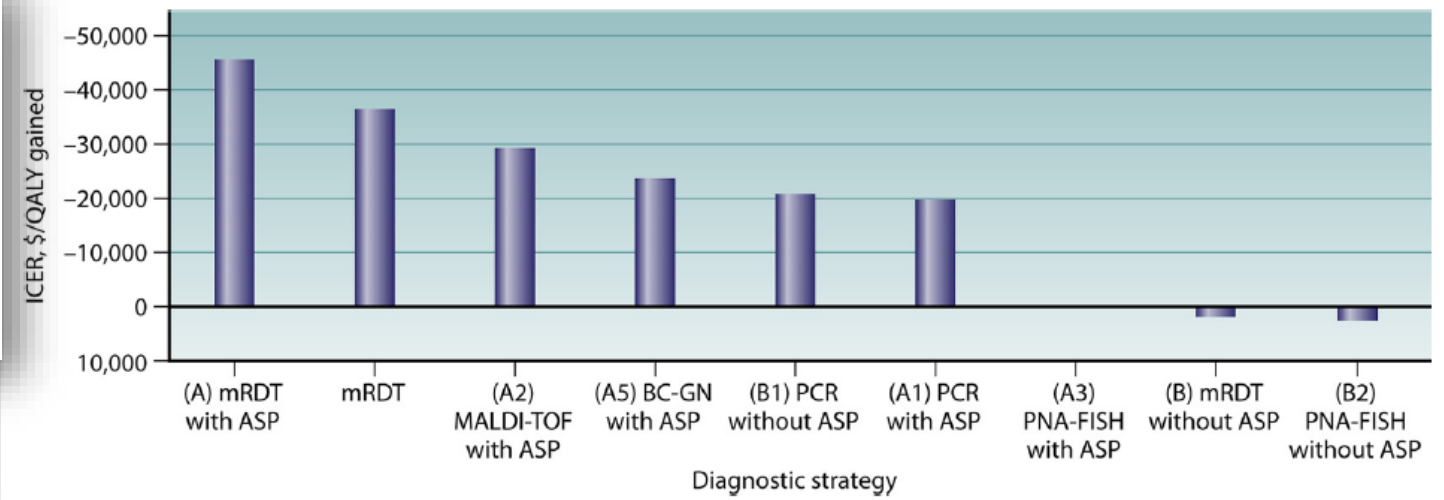


TABLE 3 Base-case analysis results for competing strategies

Strategy	Base-case estimate				
	Cost (\$)	Effect		ICER	
		Probability of survival	QALY value	\$/death averted	\$/QALY gained
mRDT (with and without ASP)	36,301.50	0.89	11.9883	−490,763	−36,434
mRDT with ASP (A)	31,274.24	0.89	11.9883	−616,445	−45,764
mRDT without ASP (B)	57,220.14	0.90	12.1230	25,762	1,913
PCR with ASP (A1)	47,917.58	0.88	11.8536	−267,148	−19,833
MALDI-TOF analysis with ASP (A2)	28,394.21	0.92	12.3924	−393,397	−29,205
PNA-FISH with ASP (A3)	53,226.09	0.85	11.4495	0	0
BC-GP with ASP (A4)	24,904.91	0.84	11.3148	Dominated	Dominated
BC-GN with ASP (A5)	33,691.47	0.92	12.3924	−317,722	−23,587
PCR without ASP (B1)	47,430.21	0.88	11.8536	−283,394	−21,039
PNA-FISH without ASP (B2)	58,284.16	0.92	12.3924	33,602	2,495
Conventional laboratory methods with ASP (C)	41,723.98	0.84	11.3148	Dominated	Dominated
Conventional laboratory methods without ASP (D)	55,932.02	0.85	11.4495	Baseline	Baseline



Ταχεία μοριακή διάγνωση και diagnostic stewardship

Right test



Right patient



Right time



TABLE 1 Key diagnostic stewardship considerations for implementation of rapid infectious disease diagnostics

Goal	Key question	Key considerations and potential strategies
Right test	Is the test appropriate for the clinical setting?	Sensitivity and specificity Predictive values Testing volumes Diagnostic yield Laboratory feasibility Cost Clinical impact
Right patient	Will the clinical care of the patient be affected by the test result?	Laboratory test utilization committee Automatic laboratory reflex CPOE decision support Appropriate use criteria Indication selection Prior authorization Benchmarking Specimen rejection
Right time	Will the result be available in time to optimally affect care?	Time to specimen receipt Centralized vs point-of-care testing On-demand vs batched testing Specimen preparation time Run time Result reporting time

Λειτουργεία 24/7?

Τρόπος παρουσίασης αποτελεσμάτων

Μοριακή ανίχνευση: αναφορά αποτελέσματος

Reporting

“detected” or “not detected” for a specific molecular target

+

interpretative comments with therapeutic guidance

Simner PJ et al. Clin Infect Dis. 2023;76(9):1550-1558.

Organisms Detected: Enterobacterales Klebsiella pneumoniae group		Controls: Passed
Applicable Antimicrobial Resistance Genes Detected: CTX-M NDM		
Note: Antimicrobial resistance can occur via multiple mechanisms. A Not Detected result for antimicrobial resistance gene(s) does not indicate antimicrobial susceptibility. Subculturing is required for species identification and susceptibility testing of isolates.		
Result Summary		
Gram Negative Bacteria		
✓	Not Detected	Acinetobacter calcoaceticus-baumannii complex
	Not Detected	Bacteroides fragilis
	Detected	Enterobacterales
	Not Detected	Enterobacter cloacae complex
	Not Detected	Escherichia coli
	Not Detected	Klebsiella aerogenes
	Not Detected	Klebsiella oxytoca
✓	Detected	Klebsiella pneumoniae group
	Not Detected	Proteus spp.
	Not Detected	Salmonella spp.
	Not Detected	Serratia marcescens
	Not Detected	Haemophilus influenzae
	Not Detected	Neisseria meningitidis
	Not Detected	Pseudomonas aeruginosa
	Not Detected	Stenotrophomonas maltophilia
Yeast		
	Not Detected	Candida albicans
	Not Detected	Candida auris
	Not Detected	Candida glabrata
	Not Detected	Candida krusei
	Not Detected	Candida parapsilosis
	Not Detected	Candida tropicalis
	Not Detected	Cryptococcus neoformans/gattii

Organisms Detected: Enterobacterales Klebsiella pneumoniae group		Controls: Passed
Applicable Antimicrobial Resistance Genes Detected: CTX-M NDM		
Note: Antimicrobial resistance can occur via multiple mechanisms. A Not Detected result for antimicrobial resistance gene(s) does not indicate antimicrobial susceptibility. Subculturing is required for species identification and susceptibility testing of isolates.		
Result Summary		
Antimicrobial Resistance Genes		
✓	Detected	CTX-M
	Not Detected	IMP
	Not Detected	KPC
	Not Detected	mcr-1
⊘	N/A	mecA/C
⊘	N/A	mecA/C and MREJ (MRSA)
✓	Detected	NDM
	Not Detected	OXA-48-like
	N/A	vanA/B
	Not Detected	VIM
Gram Positive Bacteria		
	Not Detected	Enterococcus faecalis
	Not Detected	Enterococcus faecium
	Not Detected	Listeria monocytogenes
	Not Detected	Staphylococcus spp.
	Not Detected	Staphylococcus aureus
	Not Detected	Staphylococcus epidermidis
	Not Detected	Staphylococcus lugdunensis
	Not Detected	Streptococcus spp.
	Not Detected	Streptococcus agalactiae (Group B)
	Not Detected	Streptococcus pneumoniae
	Not Detected	Streptococcus pyogenes (Group A)

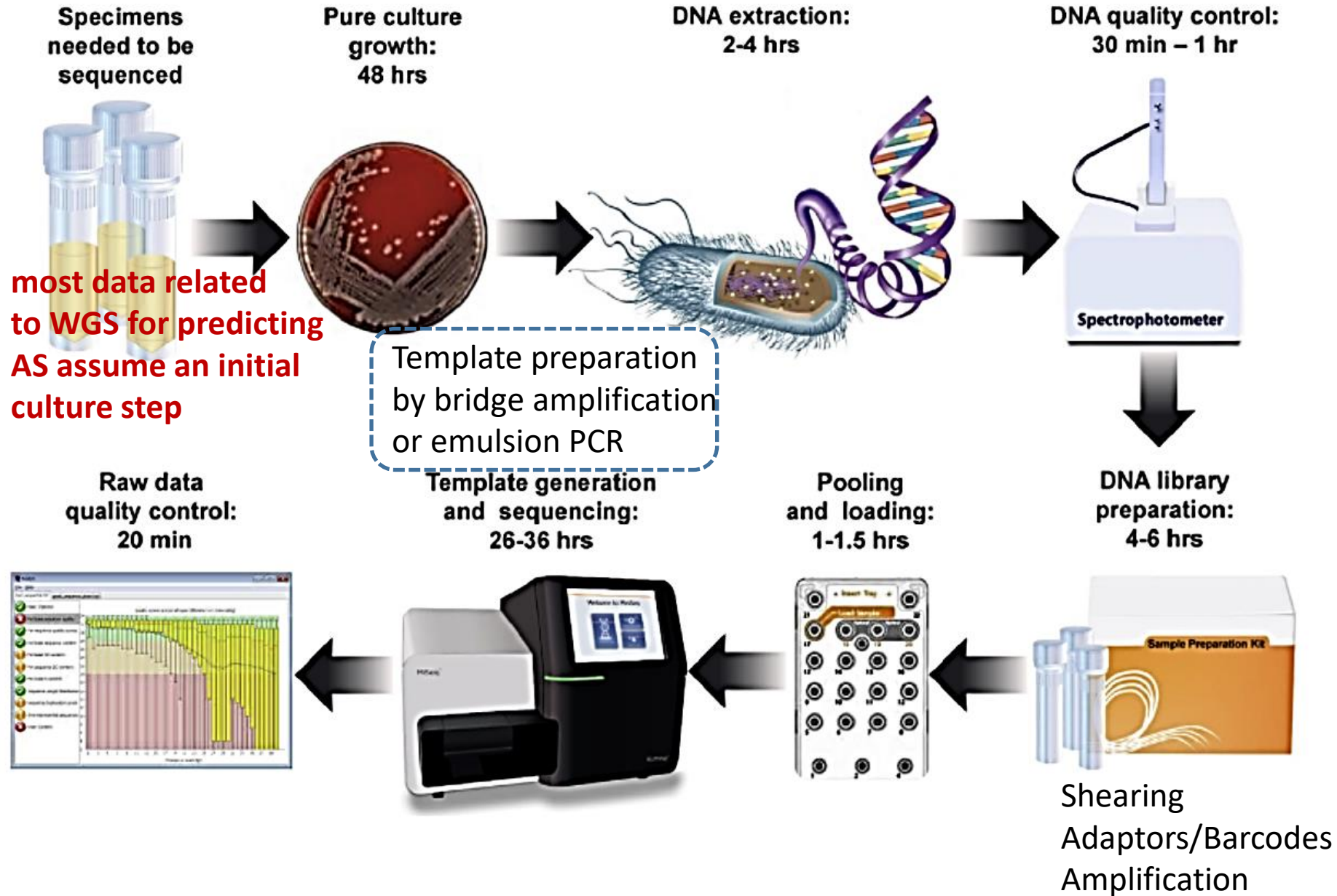
Μοριακή ανίχνευση αντοχής: σχόλια για την αναφορά των αποτελεσμάτων

Table 3. Reporting comments for molecular blood culture diagnostics.

Adapted from Banerjee et al.²⁹

<i>Staphylococcus aureus</i> , <i>mecA</i> detected	Probable methicillin-resistant <i>Staphylococcus aureus</i> (MRSA); further testing in progress. MRSA is predictably resistant to beta-lactam antibiotics (except ceftaroline). Patient requires contact precautions if hospitalized.	<i>Enterobacter cloacae</i> complex	This organism may contain an inducible β -lactamase. Penicillin or second- or third-generation cephalosporin monotherapy may result in emergence of high-level resistance.
<i>Staphylococcus aureus</i> , <i>mecA</i> not detected	Methicillin (oxacillin)-susceptible <i>Staphylococcus aureus</i> . Preferred therapy is an anti-staphylococcal β -lactam antibiotic, unless clinically contraindicated.	<i>Escherichia coli</i> , <i>bla</i> _{CTX-M} detected	Extended-spectrum β -lactamase-producer. Carbapenems are drugs of choice for ESBL-producers.
<i>Staphylococcus</i> , coagulase-negative, <i>mecA</i> detected	Methicillin (oxacillin)-resistant coagulase-negative <i>Staphylococcus</i> . Possible blood culture contaminant (unless isolated from more than one blood culture draw or clinical case suggests pathogenicity). No antibiotic treatment is indicated for blood culture contaminants.	<i>E. coli</i> , <i>bla</i> _{KPC} detected	Carbapenemase producer. Patient requires contact precautions if hospitalized. This organism is resistant to carbapenems and other β -lactam antibiotics. Consult infectious diseases.

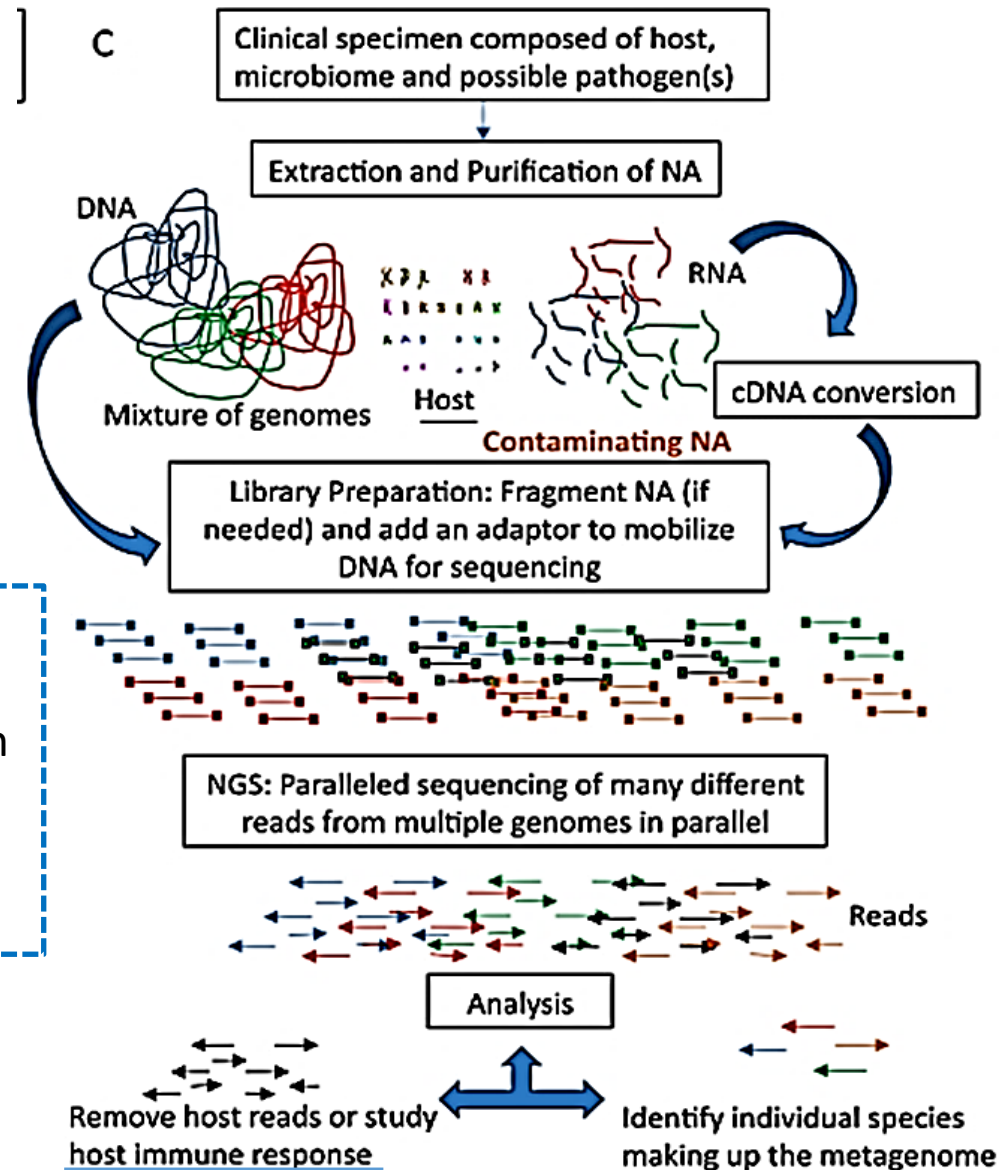
WGS: τα βήματα της διαδικασίας



Metagenomic NGS (agnostic NGS): το επόμενο βήμα

hypothesis-free,
culture-independent,
pathogen detection

evaluate the immunologic
response associated
with the presence and type of infection
e.g. biomarkers
provide insight into the pathogenesis
of the microorganisms detected



Ερμηνεία αποτελεσμάτων

WGS:
**“one test fits all”
approach**



NAAT versus Genomic data

Tweet versus... Tome

Leo Tolstoy, War and Peace

Napoleon invades Russia.

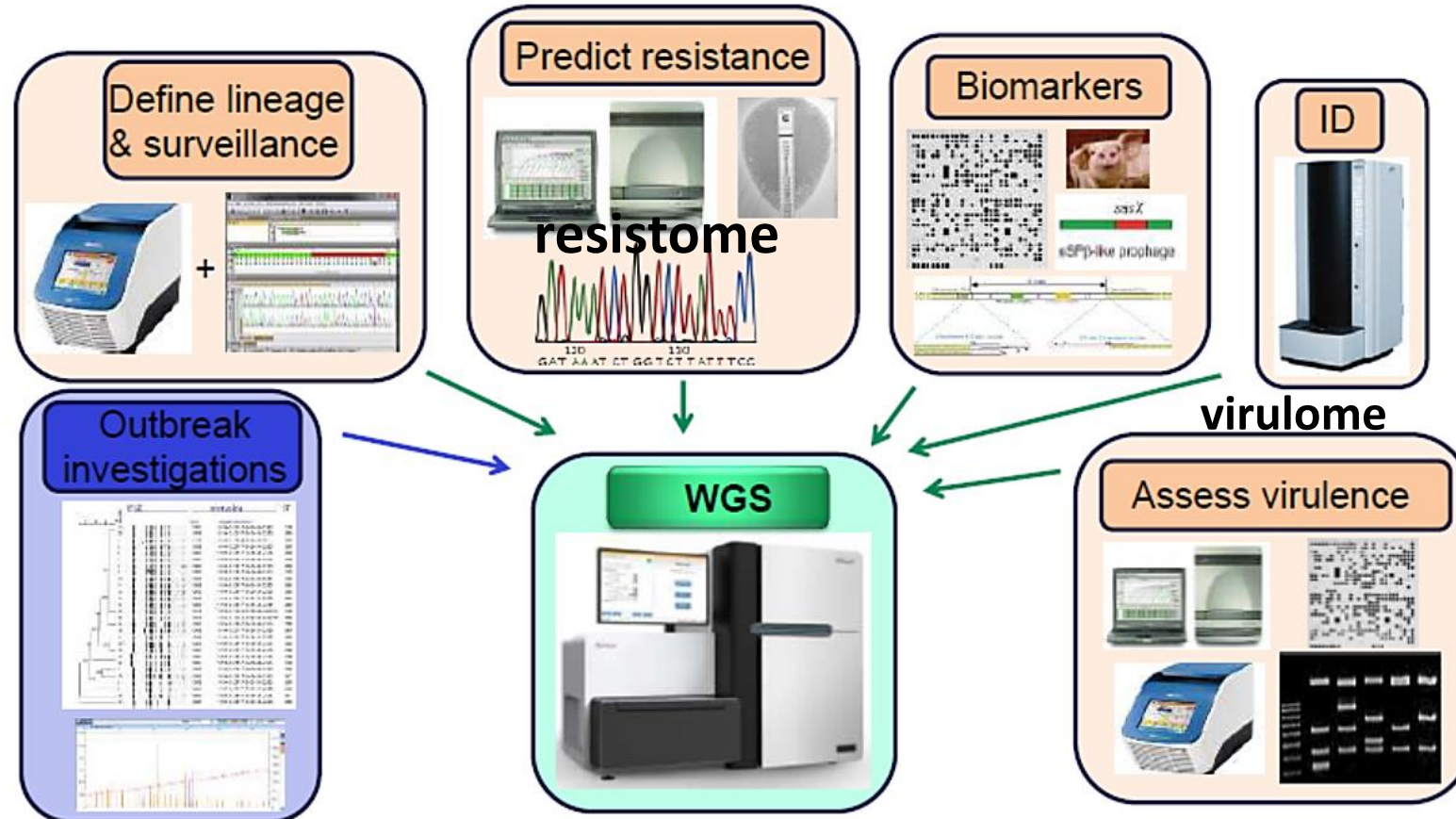
Russian aristocratic families sent into a tizz

War ensues. French retreat.

Russians celebrate. Lots of them marry.



Reed T. ECCMID, 2016.

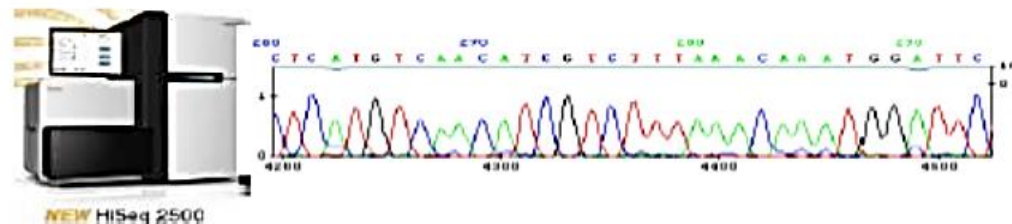


Woodford N. ECCMID, 2016.

	Phenotypic AST	WGS-based AST
Measures susceptibility	✓	✗
Resistance mechanisms	✓ (limited)	✓✓✓
ECOFF (WT vs. non-WT)	✓	✓
Clinical resistance (S vs. R)	✓	? (must be inferred)
Additional data	✗	✓✓✓
Suitable speed	✓ (most) ✗ (e.g. TB)	✗ (most) ✓ (e.g. TB)
Cost	✓	✗

M. tuberculosis: the speed of WGS results offers advantage over traditional AST

Woodford N. ECCMID, 2016.



- >95% of pathogen ID and AST in present CM labs is based on around 20 bacterial species and a limited number of AB
- may not be necessary to cover all possibilities to provide useful data rapidly, but rather we might focus on a limited number of agents for each pathogen initially and let more detailed data become available later.



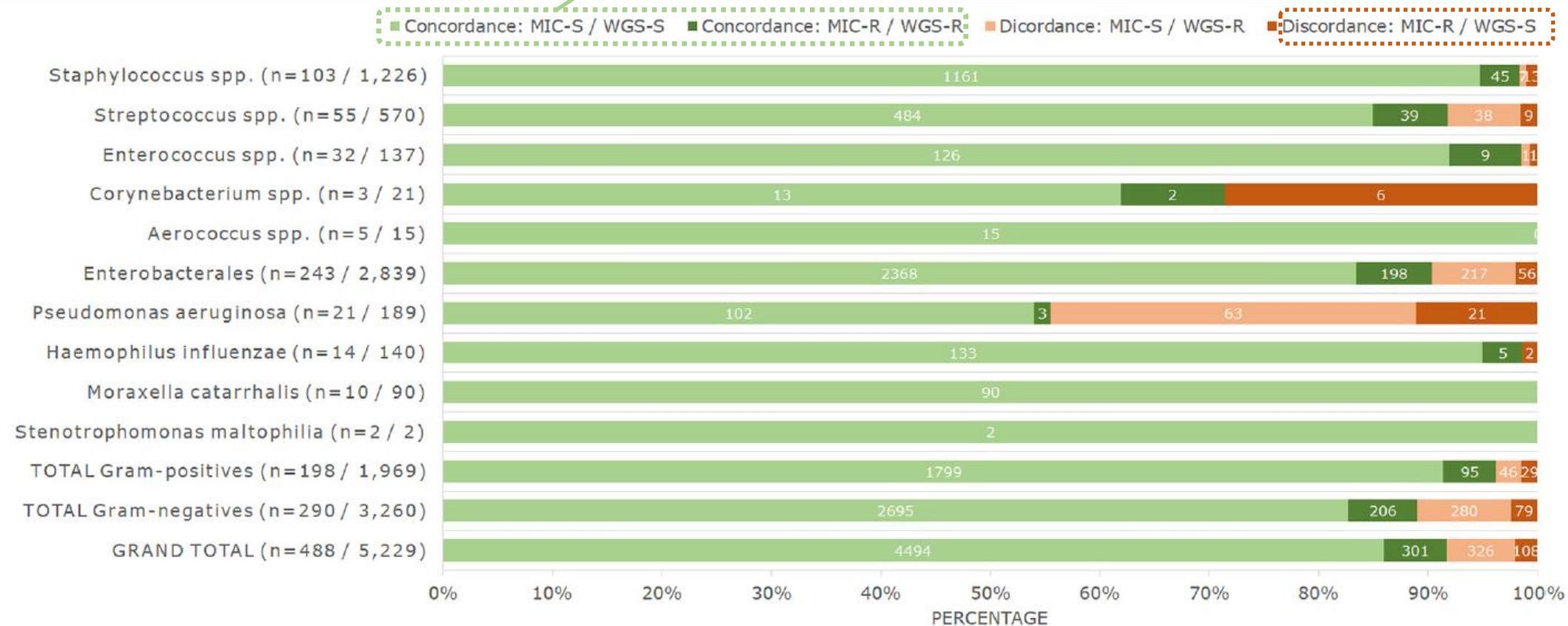
Rebelo AR et al. 2022;13:804627.

One Day in Denmark: Comparison of Phenotypic and Genotypic Antimicrobial Susceptibility Testing in Bacterial Isolates From Clinical Settings

concordance of 91.7%

cases of phenotypically resistant isolates without any known genetic resistance mechanism: 2.1% of all combinations analyzed

Percentage and number of genotype–phenotype concordances and discordances observed for all bacteria analyzed in this study, when comparing MIC and WGS results.



One size does not fit all



stewardship

Noun: the activity or job of
protecting
and being **responsible**
for something



Our time with antibiotics
is running out.

CHANGE CAN'T WAIT



THE FUTURE OF
ANTIBIOTICS
DEPENDS ON ALL OF US



World AMR Awareness Week

18-24 November 2023

Campaign guide



World Antimicrobial Awareness Week
becomes
World AMR Awareness Week



Go blue!

Join us in raising awareness of one of the biggest global public health threats: antimicrobial resistance

Every one, everywhere, at all ages is affected

Be an AMR Awareness Champion and Go Blue for AMR!