

# **System and Method Troubleshooting**

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# Troubleshooting

## (Αντιμετώπιση Προβλημάτων)

There is no standard troubleshooting procedure.

### General Pattern:

- Locate the problem by ranking (κατάταξη) possible causes.
- Verify the presence of the most probable cause.
- If present – fix the problem, otherwise verify the existence of the next possible cause.

First try to distinguish

System problem

or

Method problem

# Method vs. System Troubleshooting

## System

### Parameters

- Flow stability
- Backpressure (οπισθοπίεση)
- Clogging (απόφραξη)
- Detector problems
- Injection suitability
- Injection volume
- Temperature

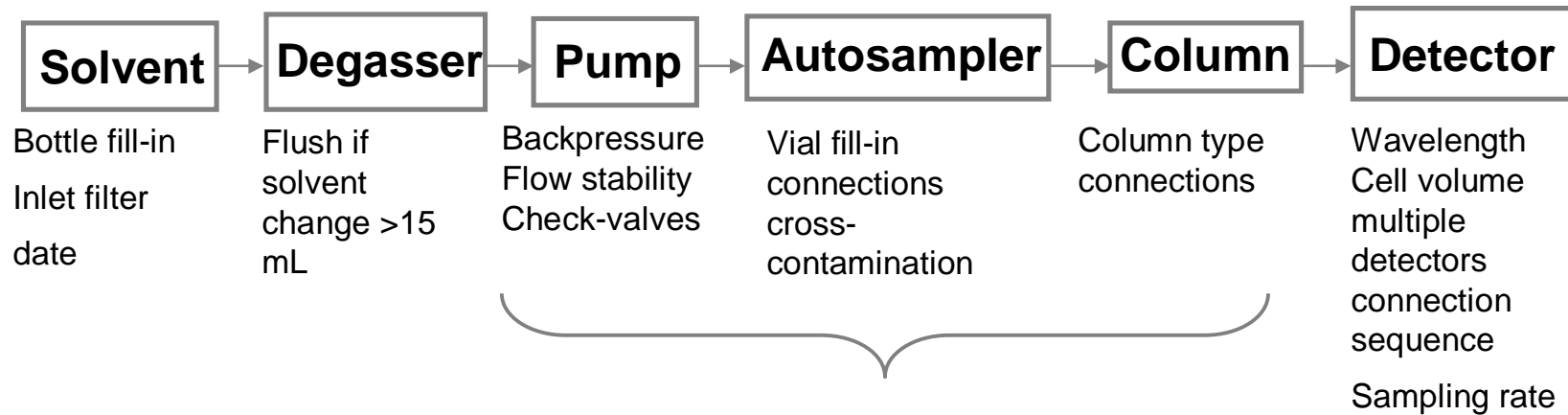
## Method

### Parameters

- Flow rate
- Eluent (εκλουστικό) type
- Eluent composition
- pH
- pH modifier (τροποποιητής) (type)
- Injection volume
- Temperature
- Gradient profile

# System Parameters

- Simple preliminary verification of system setup can save time.



**Critical connections. Minimize tubing length**

# System Suitability

## Available HPLC system set margins (περιθώρια) for column selection.

- 20  $\mu\text{l}$  detector flow-cell incompatible with  $<3$  mm I.D. columns
- 10  $\mu\text{l}$  sample loop incompatible with  $<1$  mm I.D. columns.
- 0.2  $\mu\text{l}$  micro-injector is useless for conventional columns.

## Suitability Rule

Injection volume  $<$  Cell volume

Column Dead Volume  $\approx 0.65$  of the empty column volume

# System Suitability

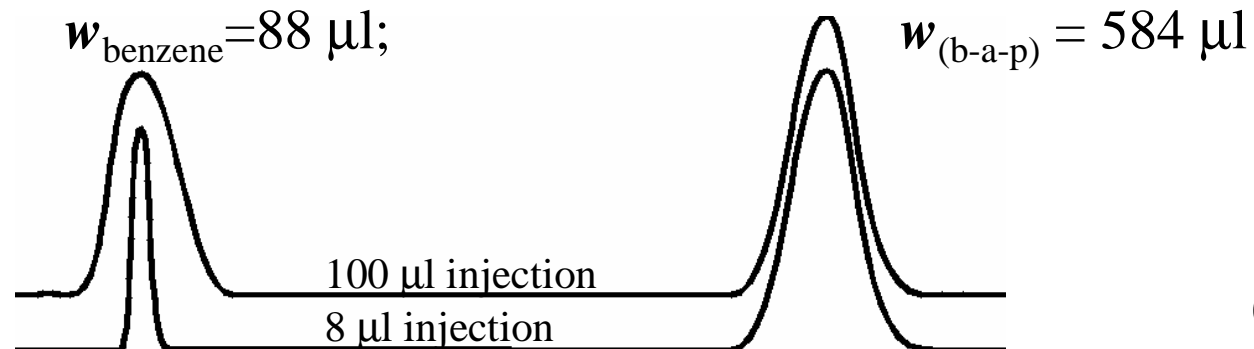
## (Injection Volume)

**Column:** 150 x 4.6 mm (C18),  $V_o = 1.7$  ml

**Efficiency:** 10,000 t.p. **Eluent:** MeCN/Water 70/30

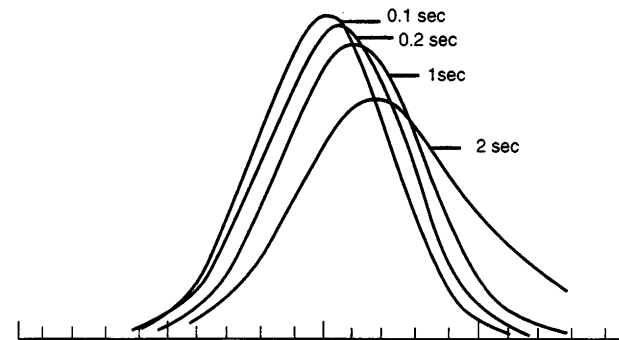
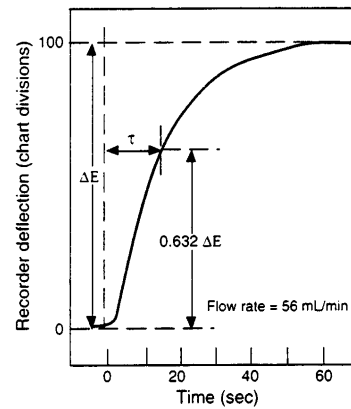
$V_{R(\text{benzene})} = 2.2$  ml;       $V_{R(\text{benz-a-pyrene})} = 14.6$  ml

$$N = 16 \left( \frac{V_R}{w_b} \right)^2 \quad \text{or} \quad w_b = \frac{4V_R}{\sqrt{N}}$$

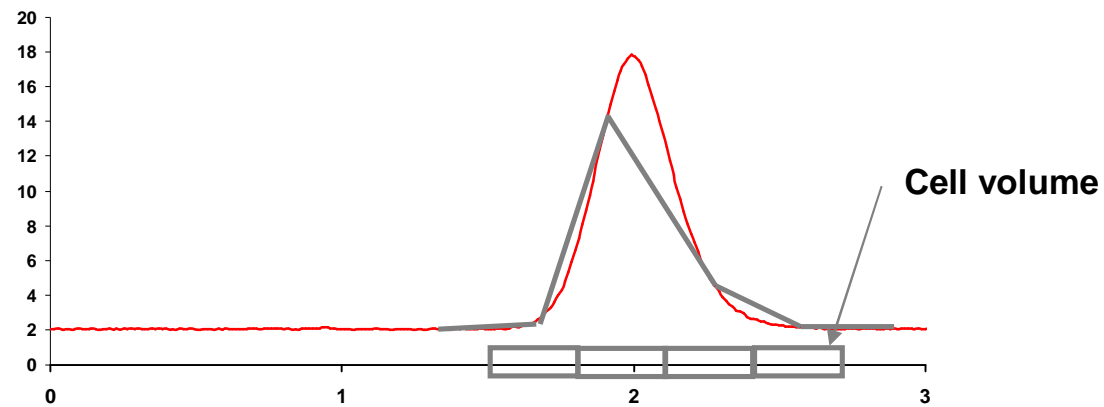


# Effect of flow-cell volume and sampling rate

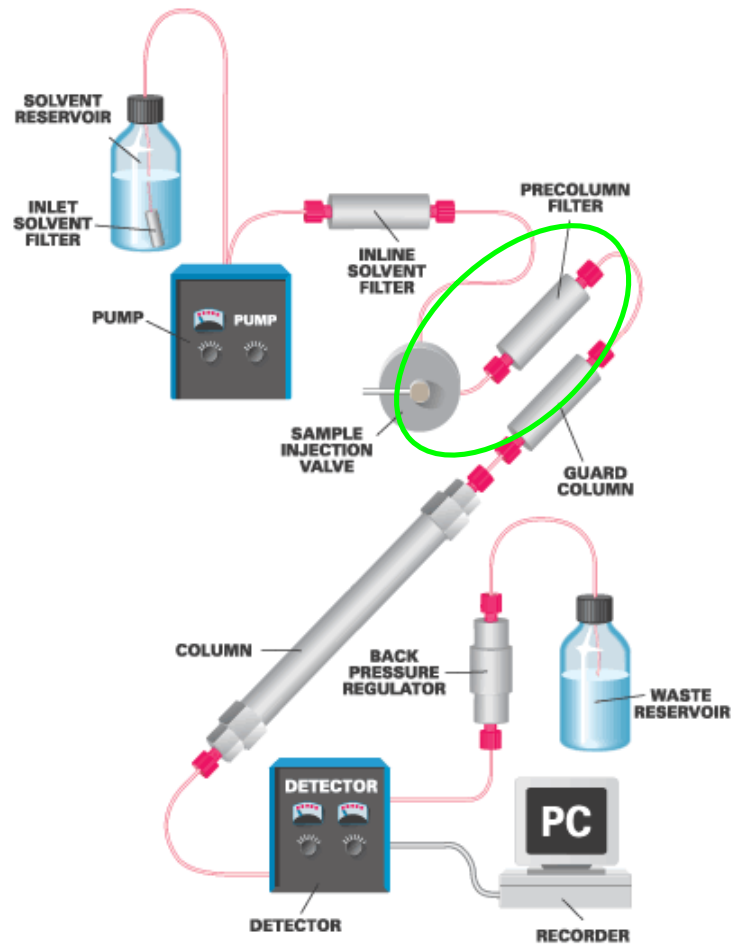
## Response time (σταθερά απόκρισης) effect



## Flow cell volume



# HPLC System set up



- Minimize the volume and connections between autosampler, column, and detector.
- No guard (προστασία), no prefilter

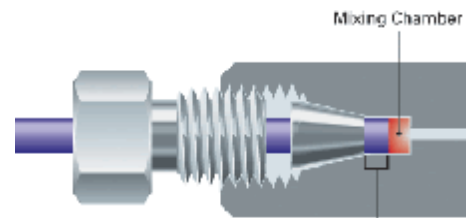
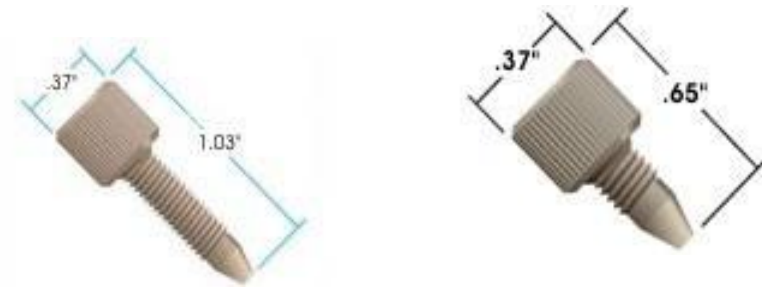




# Tubing & connections

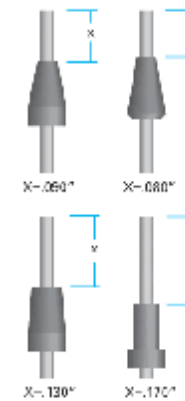


1560	.0025" (65µm) ID	Natural	7,000 psi (483 bar)*
1561	.004" (100µm) ID	Black	7,000 psi (483 bar)*
1535	.005" (125µm) ID	Red	7,000 psi (483 bar)*
1562	.006" (150µm) ID	Purple	7,000 psi (483 bar)*
1536	.007" (175µm) ID	Yellow	7,000 psi (483 bar)*
1531	.010" (.25mm) ID	Natural	7,000 psi (483 bar)*
1531B	.010" (.25mm) ID	Blue	7,000 psi (483 bar)*
1565	.015" (.40mm) ID	Gray	7,000 psi (483 bar)*
1532	.020" (.50mm) ID	Orange	7,000 psi (483 bar)*



If Dimension X is too short, a dead-volume or mixing chamber, will occur.

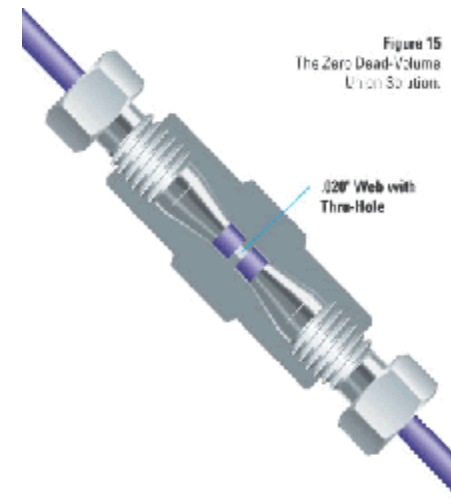
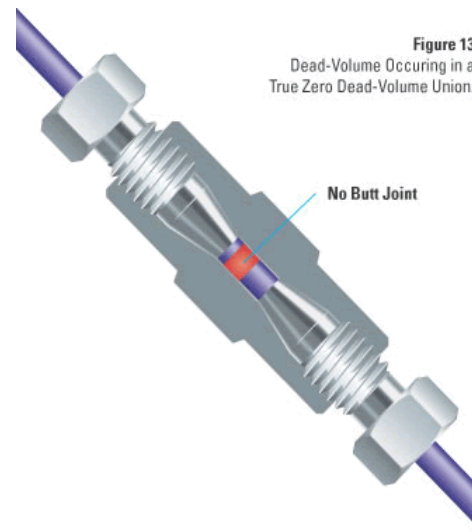
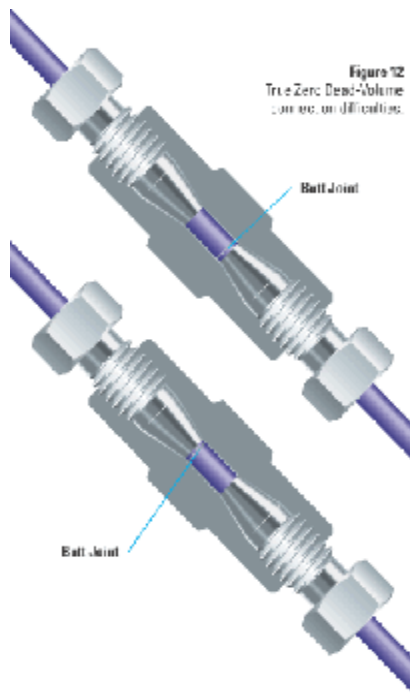
Figure 3b



Dimension X ranges from .035" to .170" among various manufacturers.

Figure 2

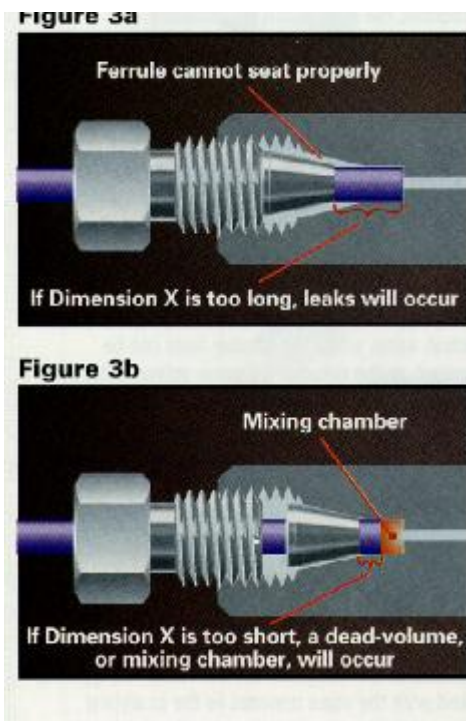
# Unions (Butt Joint = σύνδεσμος αρμού)



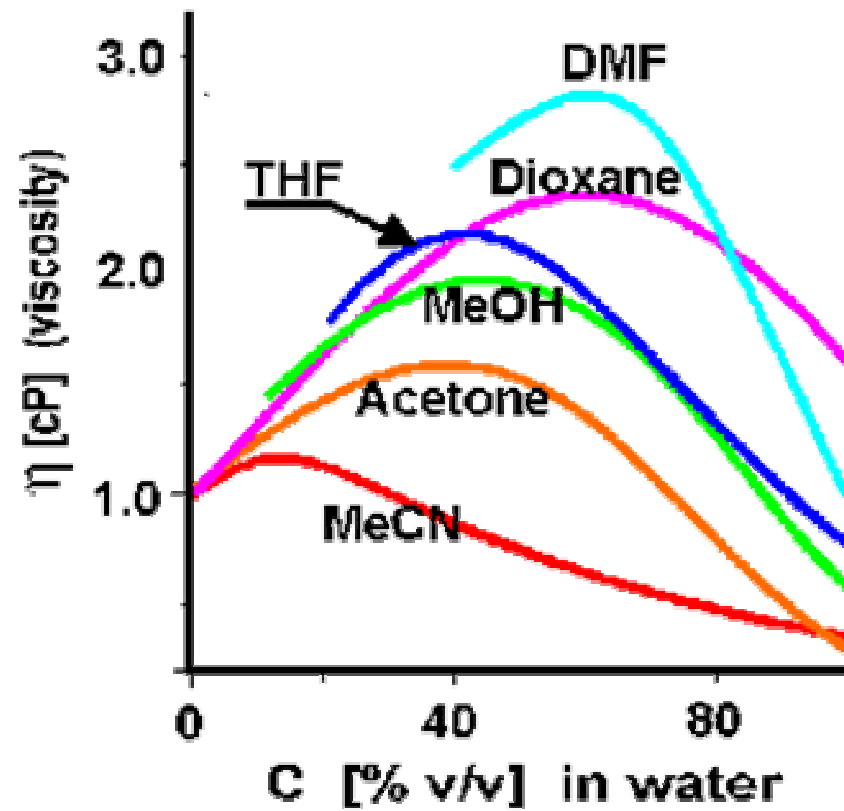
# Critical Connections

Injector - Column, Column - Detector

No unions, in-line filters, guard columns  
Single piece tubing (0.007" and smaller I.D.)



## Eluent Composition Effects on the Column Back Pressure



# Guard Columns

## Στήλες Προστασίας

Purpose - trapping retentive impurities (παγίδευση συγκρατούμενων ακαθαρσιών)

Disadvantage - introduces extra-connections in critical zone

Sample has 1% impurity. How many injections will kill 1% of column surface with 1% sample solution and 10  $\mu$ l injection volume?

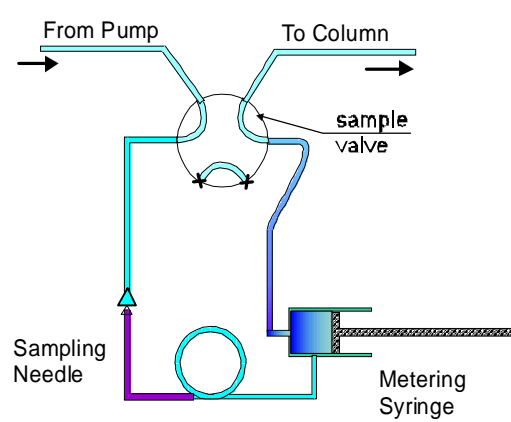
1% column surface  $\sim$  2-3  $m^2$ , it could adsorb  $\sim$  0.1  $\mu$ Mole

**300** injections will reach this level.

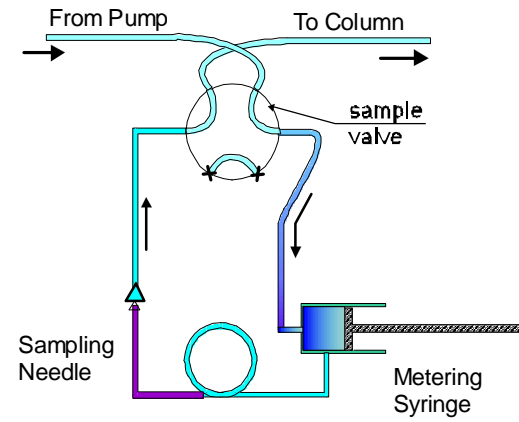
Guard column always decreases system efficiency

	Retention time		W1/2		Theoretical Plates	
	Guard	No Guard	Guard	No Guard	Guard	No Guard
aniline	2.743	2.696	.1047	0.083	3802	5845
Methyl aniline	3.898	3.734	0.0865	0.0832	11250	11159
NN-dimethyl aniline	4.274	4.188	0.0952	0.0879	11166	12576

# Autosampler – Column/Pump Connections



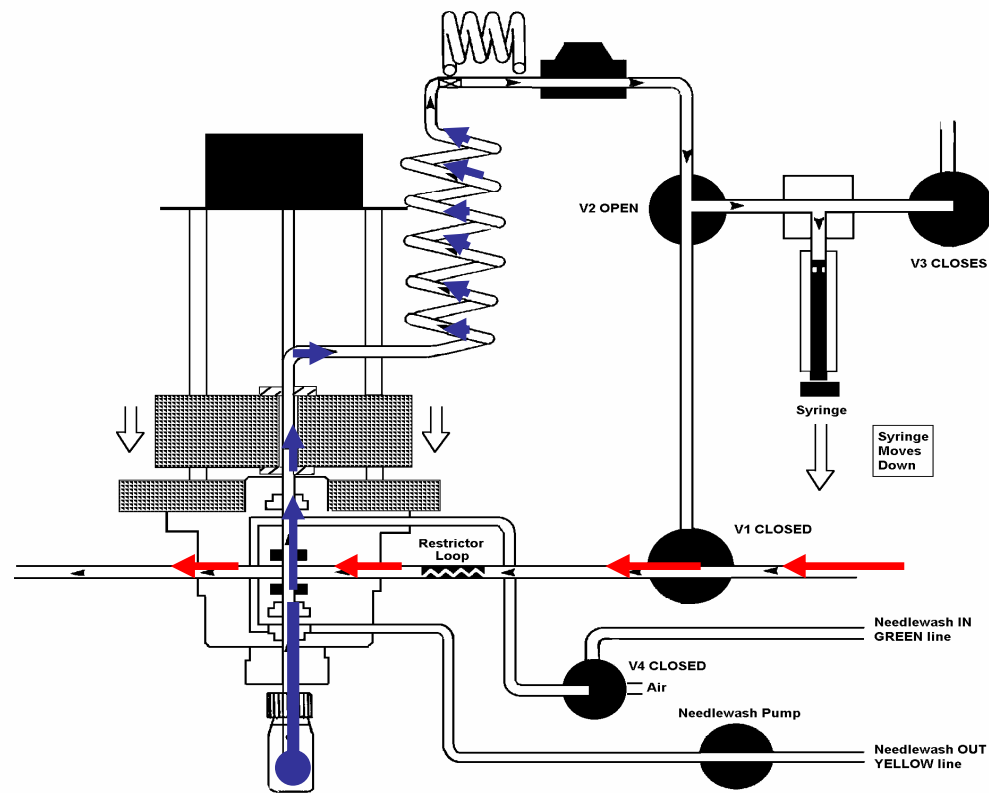
Wrong connection



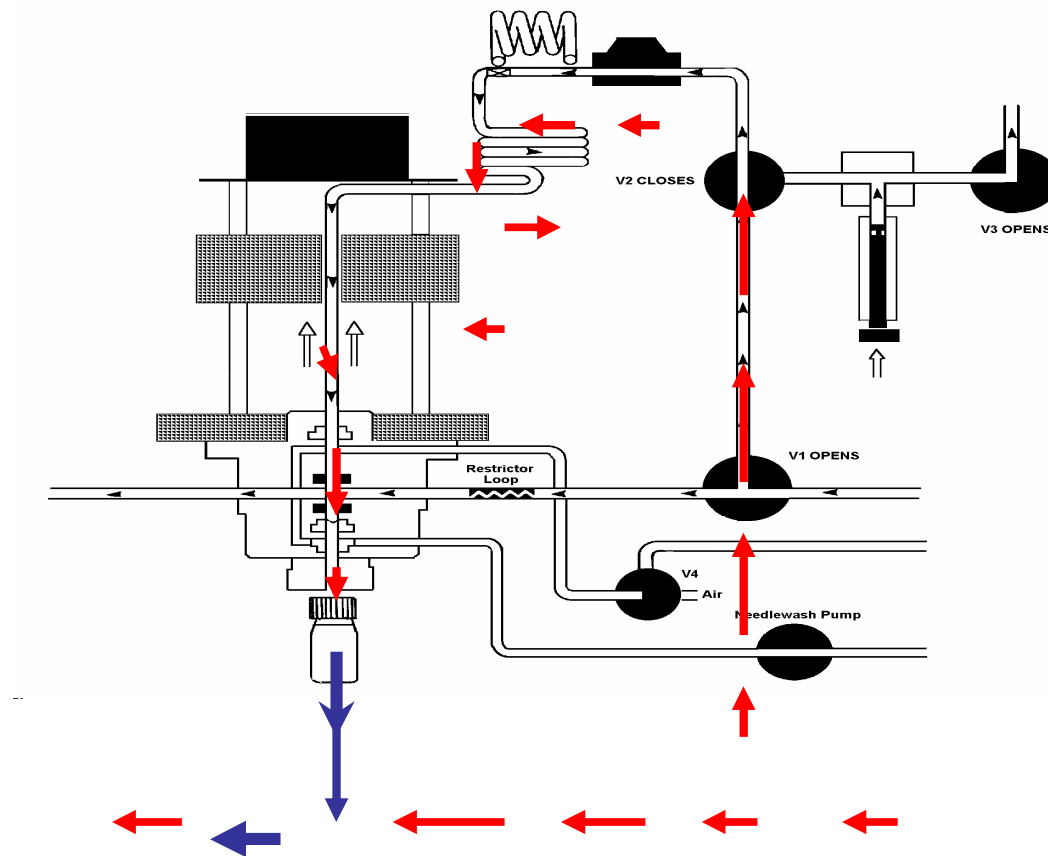
Correct connection



# Waters system (Injection, Drawing Sample)



# Waters system (Injection, Injecting Sample)





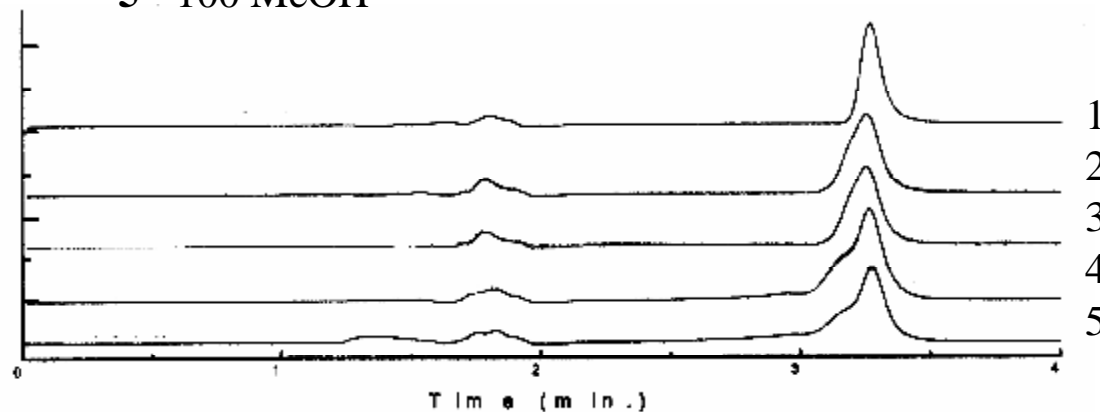
## Sample Diluent (Αραιωτής) Effect

Sample diluent:

- 1- 50/50 MeOH/Water
- 2 - 80/20 MeOH/Water
- 3 - 90/10 MeOH/Water
- 4 - 95/5 MeOH/Water
- 5 - 100 MeOH

50/50 Buffer/MeOH

Buffer: 20 mM Citrate, pH=4.6



**Incompatible solvents may cause sample precipitation and column clogging**

Different eluent pH and composition may cause peak splitting

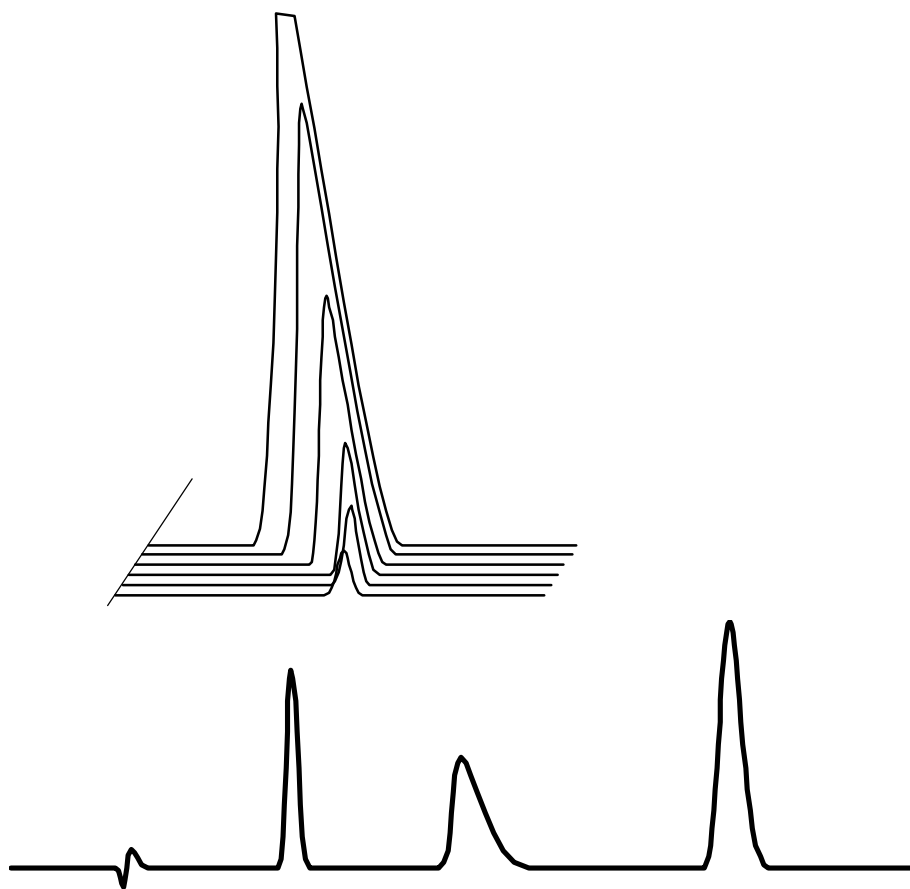
# Column Length

- Column length is a compromise (συμβιβασμός) between the efficiency and backpressure
- Column efficiency is proportional to the column length
- **Specific efficiency** (# of particles per one plate) decreases with length increase.

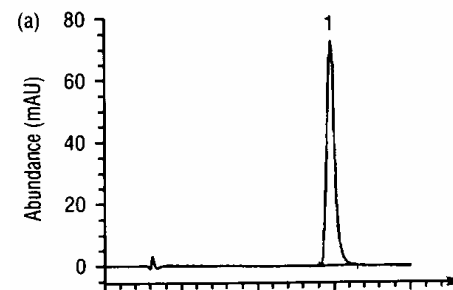
Length [cm]	Particle Dia. [um]	Efficiency, N	Specific Efficiency, <i>h</i>
10	3	11111	3
10	5	10526	1.9
15	5	13636	2.2
25	5	15625	3.2
25	10	10000	2.5

# Column Overloading

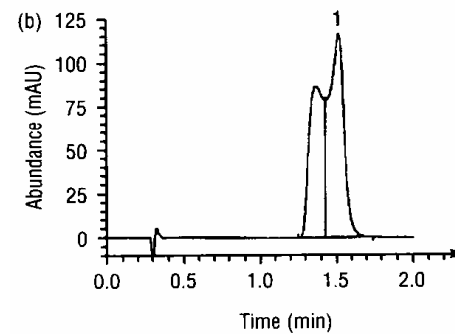
## Υπερφόρτωση Στήλης



1  $\mu\text{l}$

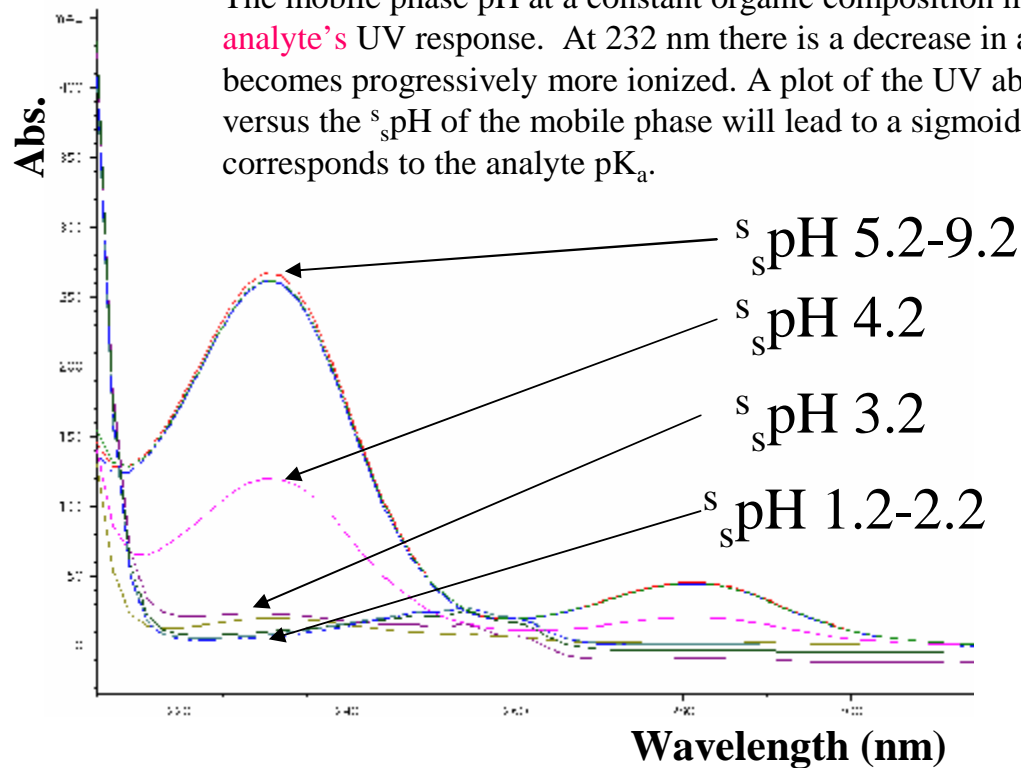


5  $\mu\text{l}$



# Effect of pH on Aniline ( $pK_b = 9,42$ , $pK_a = 4,58$ ) UV absorbance

The mobile phase pH at a constant organic composition may have an effect on an **ionizable analyte's** UV response. At 232 nm there is a decrease in aniline's absorbance as this analyte becomes progressively more ionized. A plot of the UV absorbance at a particular wavelength versus the  $s_p$ pH of the mobile phase will lead to a sigmoidal dependence. The inflection point corresponds to the analyte  $pK_a$ .



## Chromatographic Conditions

Column: 15 cm x 0.46 cm Luna C18(2)

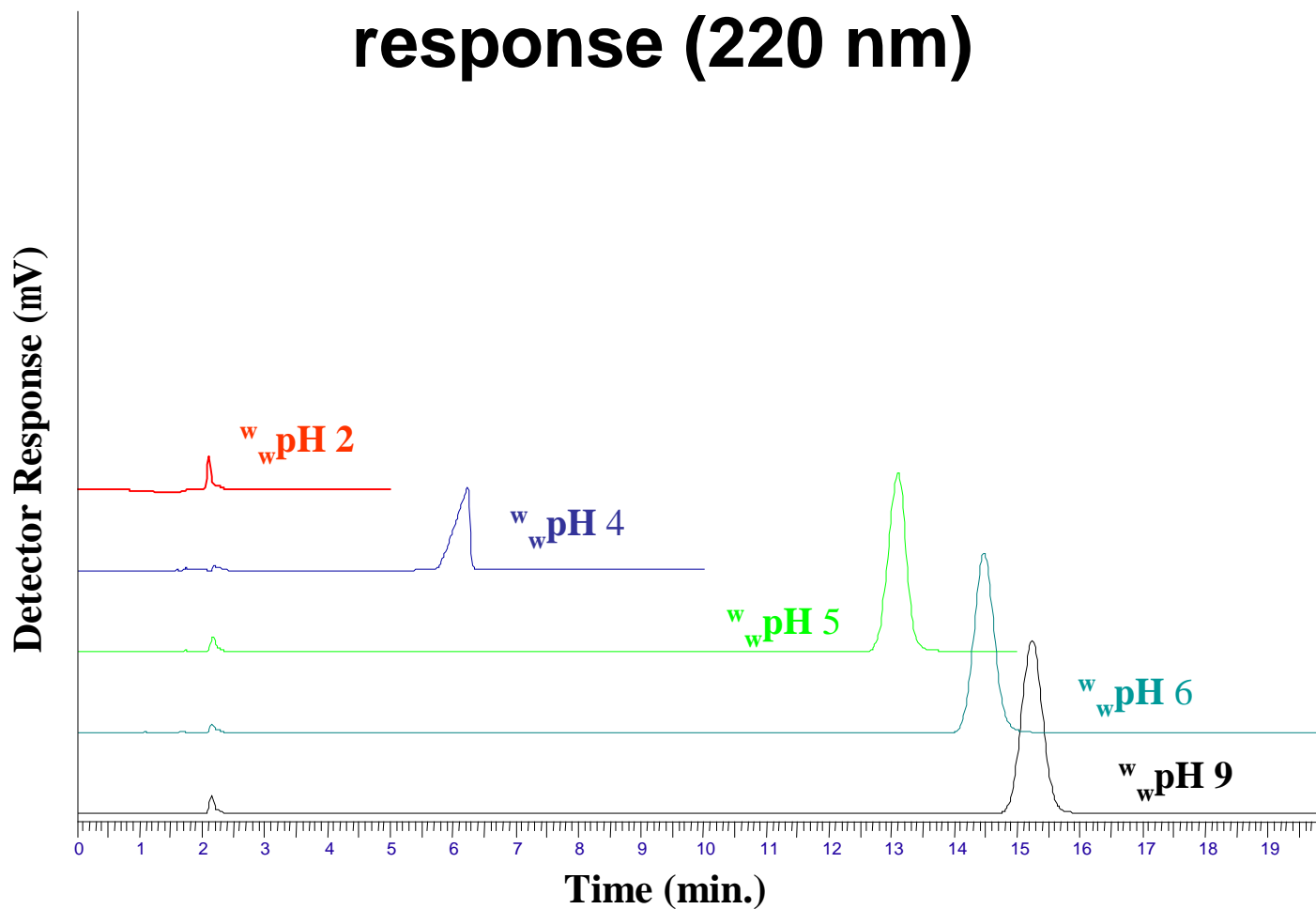
Eluent: 90% Aqueous:10% MeCN

Aqueous: 15 mM  $K_2HPO_4 \cdot 7H_2O$  adj. to  
 $w_w$  pH 1 - 9 with  $H_3PO_4$

Flow rate: 1 ml/min

Temp: 25°C

# Effect of pH on Aniline Retention and UV response (220 nm)



## Chromatographic Conditions

Column: 15 cm x 0.46 cm Luna C18(2) Eluent: 90% Aqueous: 10% MeCN

Aqueous: 15 mM  $K_2HPO_4 \cdot 7H_2O$  adj. to  $w_w$ pH 1.5 - 9 with  $H_3PO_4$

Flow rate: 1 ml/min Temp: 25°C

- Enhanced sensitivity is obtained by analyzing aniline in its neutral state 21

# Column Equilibration

- Column equilibrates (εξισορροπεί) within 30 min in normal eluent composition range.
- Check retention time stability by injecting standard mixture 3 - 4 times.
- Very high organic (>98%) or very high aqueous (>80%) need ~1 - 2 h equilibration at 1 ml/min.
- In pure water after ~20 h equilibration all analytes elute with void volume. “Chain collapse”? - No. After 20 h of water pumping all organic removed from adsorbent pores. Water is not wetting the alkylated hydrophobic surface. There is no flow through adsorbent particles, only around.

# Solvent Purity

How much solvent (0.1 ppm total impurity) will contaminate 10% of adsorbent surface?

Average column - 200 m<sup>2</sup>/g

Assume molecular area of 100 Å<sup>2</sup>

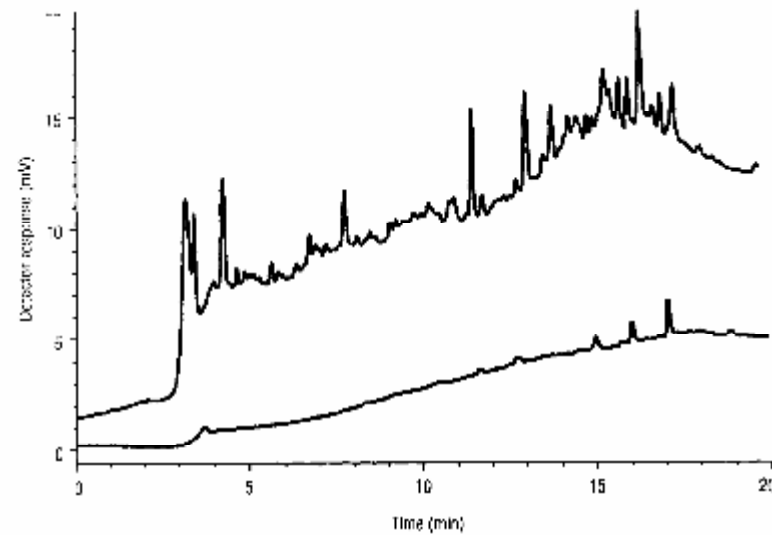
$$n_{\text{(moles)}} = \frac{S}{A \cdot N_A} = \frac{20m^2}{100\text{\AA}^2 \cdot 6 \cdot 10^{23}} \approx 30 \quad \text{mMole}$$

Assume average 100 g/mole - 3 mg total accumulation  
this comes from **30 L** of solvent with 0.1 ppm total purity

Column has to be cleaned at least once a week

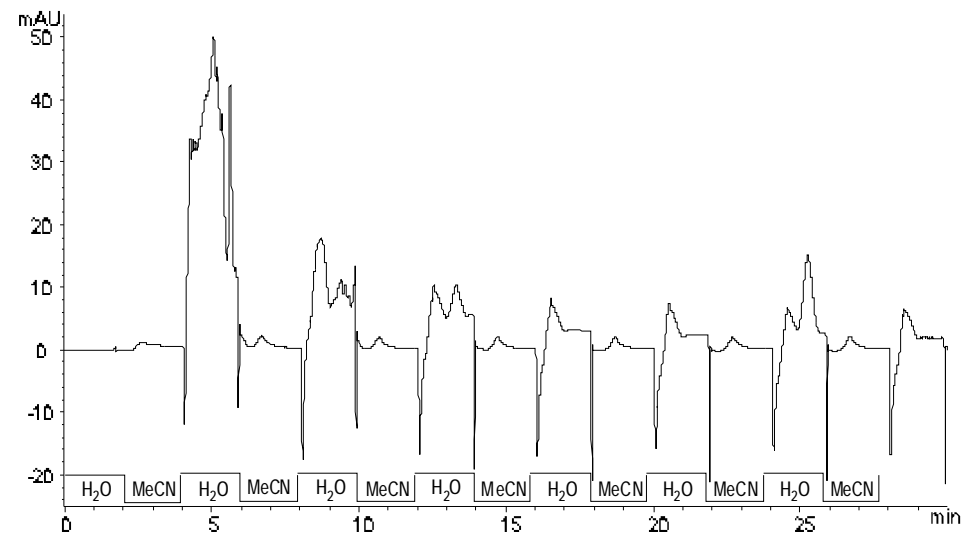
# Gradient

- High pressure vs. low pressure mixing
- System dwell (νεκρός) volume effect





# Column Cleaning



Solvent front (μέτωπο διαλύτη) disturbs phase equilibrium  
Release of trapped (παγιδευμένες) impurities

# Method troubleshooting

- Problems are usually related to one of the following:
  1. **System**
  2. **Column**
  3. **Sample**
  4. **Mobile Phase**

# System

- System-to-system compatibility
  - Differences in configuration (detector sequence, etc.)
  - Different dwell volume
  - Detector sensitivity always different
  - Wavelength accuracy
  - Bandwidth
  - Environment effects

# Sample

## Avoid particulate in the sample

Typical cause of inlet filter clogging

### Filter

Sample filtration can change composition

### Centrifuge

Usually cumbersome (δυσκίνητη)

## Sample vials

Type of the vial cap and septa affect contamination and carry-over

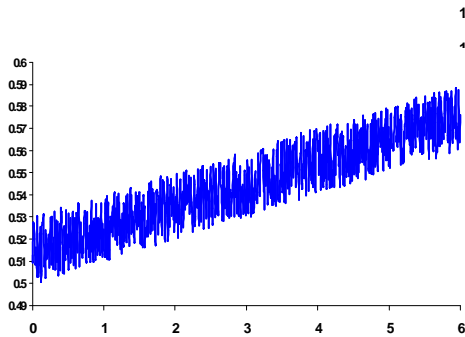
Waters systems require 75% filling of 2 mL vial

# Troubleshooting sequence

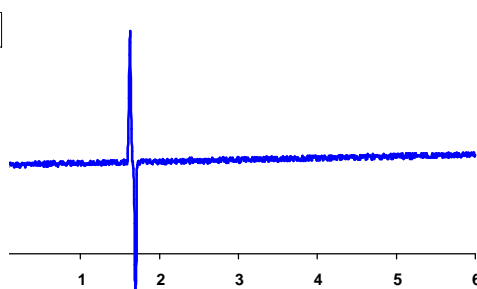
- Pump
  - Any reciprocal pattern (ανάποδη εικόνα) on chromatogram
  - Pressure fluctuations
  - Baseline drift (possible contamination of the solvent)
- Autosampler
  - Injection marks (baseline disturbance)
  - Cross-contamination
  - Vial fill-in (sample level)
- Detector
  - Response (baseline noise, drift, etc.)
  - Wavelength (bandwidth, accuracy, etc.)

# Troubleshooting sequence

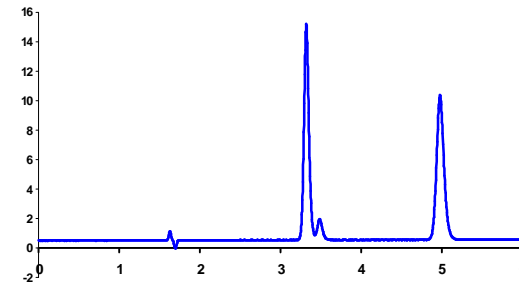
- First check is always the plumbing (σωληνώσεις) (leak, flow rate, pressure)
- Output (chromatogram) evaluation



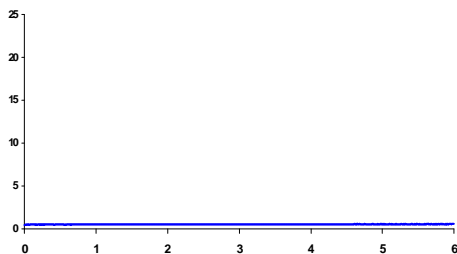
1



2



3



- 1- flow or detection problem
- 2 – possible injection problem
- 3 – correct chromatogram

# Troubleshooting sequence

- Analysis of chromatogram
  - Compare with previous results
  - Peak tailing
  - Retention shift
  - Reverse elution

