Male infertility

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Conflict of interests

- During the last three years, I have received research, educational and travel grants from scientific societies and pharmaceutical companies.
- I am currently or have been:
 - President, Hellenic Society of Andrology (2013 2017)
 - President-Elect, European Menopause and Andropause Society (2019 2021)
 - Secretary General, European Academy of Andrology (2014 2018)
 - Director, Training Centre European Academy of Andrology (2013 2019)
 - Member of the Executive Board, Hellenic Society of Endocrinology (2011 2013)
 - Member of the Executive Board, Hellenic Society of Climacteric and Menopause (2013 2019)
 - Associate Editor, Human Reproduction (2009 2013)
 - Associate Editor, Hormones (2012 2019)
 - Member of the Editorial Board, Andrology (2012 2019)
 - Member of the Editorial Board, Maturitas (2015 2019)
 - Member of the Editorial Board, Metabolism (2015 2019)
 - Member of the Editorial Board, Journal of Endocrinological Investigation (2014 2019)
 - Associate Editor, Human Reproduction Open (2017 2019)
 - Associate Editor, Human Andrology (2017 2019)
- None of the above can be considered as a conflict of interest for today's lecture.

Aims

- Evaluation and etiologic approach of male infertility
- Selection of the optimal management method

Diagnostic approach



Diagnostic approach



Causes of male infertility

Cause	Prevalence (%)
 Idiopathic infertility 	32
Varicocele	17
 Endocrine causes 	9
 Infections 	9
 Cryptorchidism 	8
 Sexual dysfunction 	6
 Systematic diseases 	5
 Anti-sperm antibodies 	4
 Testicular tumors 	2
 Obstruction 	1
 Other causes 	7

Diagnostic evaluation

- Clinical
- Hormonal
- Seminal
- Imaging
- Histologic
- Genetic

Diagnostic evaluation

Clinical

- Hormonal
- Seminal
- Imaging
- Histologic
- Genetic

• History

 Primary or secondary infertility, duration of infertility, mumps, cryptorchidism, trauma, surgical procedures, infections, recent febrile episodes, chemotherapy or radiotherapy, medications, family history of infertility, cystic fibrosis, mental retardation, female factor

Clinical evaluation

• Testicular size, secondary sexual characteristics, presence and consistence of epididymides and vas deferens, varicocele, digital examination

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 Primary or secondary infertility, duration of infertility, mumps, cryptorchidism, trauma, surgical procedures, infections, recent febrile episodes, chemotherapy or radiotherapy, medications, family history of infertility, cystic fibrosis, mental retardation, female factor

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Diagnostic evaluation

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Hormonal evaluation

- FSH
- LH
- Testosterone, total
- Prolactin
- Thyroid evaluation
- Inhibin B (Inh B)
- Anti-Müllerian hormone (AMH)



- Differential diagnosis between central (hypothalamus, pituitary) and peripheral (testicular) failure
- Variation <10%
- Strong correlation with:
 - Testicular histology
 - Sperm count
 - GnRH stimulation

Inh B

- TGF-β family glycoprotein
- Exclusive Sertoli cell product
- Endocrine action:
 - FSH inhibition
- Paracrine effects
- Positive correlation with sperm count and testicular size
- Prognostic factor for TESE

AMH

- TGF-β family glycoprotein
- Sertoli cell product
- Endocrine action:
 - Müllerian duct reversal
- Paracrine and autocrine actions
- Prognostic factor for TESE

Testicular histology



Nieschlag E, Behre HM. Andorlogy, 1997

Diagnostic evaluation

- Clinical
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Semen reference ranges

- Liquefaction
 - < 60 min
- Appearance
 - Non-translucent
- Viscosity
 - Filaments < 2 cm
- pH
 - > 7.2

- Volume
 - > 1.5 ml
- Concentration
 - > 15 millions/ ml
- Motility
 - > 40 (a + b + c)
 - > 32 (a + b)
- Normal morphology
 - >4%

Semen reference ranges

Table II Distribution of values, lower reference limits and their 95% CI for semen parameters from fertile men whose partners had a time-to-pregnancy of 12 months or less

	N	Centi	es									
		2.5	(95% CI)	5	(95% CI)	10	25	50	75	90	95	97.5
Semen volume (ml)	1941	1.2	(1.0–1.3)	1.5	(1.4–1.7)	2	2.7	3.7	4.8	6	6.8	7.6
Sperm concentration (10 ⁶ /ml)	1859	9	(8-11)	15	(12-16)	22	41	73	116	169	213	259
Total number (10 ⁶ /Ejaculate)	1859	23	(18–29)	39	(33–46)	69	142	255	422	647	802	928
Total motility (PR + NP, %)*	1781	34	(33–37)	40	(38–42)	45	53	61	69	75	78	81
Progressive motility (PR, %)*	1780	28	(25–29)	32	(31–34)	39	47	55	62	69	72	75
Normal forms (%)	1851	3	(2.0-3.0)	4	(3.0–4.0)	5.5	9	15	24.5	36	44	48
Vitality (%)	428	53	(48–56)	58	(55–63)	64	72	79	84	88	91	92

*PR, progressive motility (WHO, 1999 grades a + b); NP, non-progressive motility (WHO, 1999 grade c).

The values are from unweighted raw data. For a two-sided distribution the 2.5th and 97.5th centiles provide the reference limits; for a one-sided distribution the fifth centile provides the lower reference limit.

Cooper TG, et al. Hum Reprod Update 2010, 16:231

Definitions

- Oligo-astheno-teratozoospermia
 - Low sperm number motility morphology
- Azoospermia
 - No presence of sperm, even after centrifugation of semen
 - Transient permanent
- Cryptozoospermia
 - No presence of sperm, after the initial inspection
 - Presence of sperm, after centrifugation of semen
- Aspermia
 - No presence of semen

Sperm concentrations



Concentration



Concentration



Concentration



Motility



Specimen 524

Motility



Aggregations



Agglutinations



Normal sperm



Head defect



Neck defect



Tail defect



Tail defect



Cytoplasmic residual


Semen reference ranges

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• pH

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- Normal morphology
 > 4%

Anatomy of male reproductive system



Diagnostic algorithm



Oates R. Asian J Androl 2012, 14:82

Anatomy of male reproductive system



Diagnostic algorithm



Anatomy of male reproductive system



Diagnostic algorithm



Oates R. Asian J Androl 2012, 14:82

Anatomy of male reproductive system



Diagnostic algorithm



Oates R. Asian J Androl 2012, 14:82

Anatomy of male reproductive system



Semen evaluation

- Spermiogram
- Biochemical evaluation of seminal plasma
- Semen culture
- Immunological evaluation
- Acrosome rection
- Sperm DNA fragmentation
- Functional tests

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Pathophysiology

- Mechanisms
 - Oxidative: Reactive Oxygen Species (ROS)
 - Anti-oxidative: Antioxidant Scavenging Systems (ASS)
- Oxidative stress in male reproductive system results in sperm membrane damage and sperm DNA fragmentation

Oxidative stress

FIGURE 1

Major mechanisms of inducing DNA damage in spermatozoa during either the production or the transport of sperm cells: (i) apoptosis during the process of spermatogenesis; (ii) DNA strand breaks produced during the remodelling of sperm chromatin during the process of spermiogenesis; (iii) post-testicular DNA fragmentation induced, mainly by oxygen radicals, during sperm transport through the seminiferous tubules and the epididymis (increasing DNA damage is indicated by size of red flashes and gradient darkening in tract); (iv) DNA fragmentation induced by endogenous caspases and endonucleases; (v) DNA damage induced by radiotherapy and chemotherapy; and (vi) DNA damage induced by environmental toxicants.



Sakkas D, et al. Fertil Steril 2010, 93:1027

Sperm DNA fragmentation



Kantartzi P-D. PhD thesis, AUTh, 2012

Evaluation methods

- Direct
 - COMET
 - TUNEL
 - NT
 - DBD-FISH

- Indirect
 - SCSA
 - AOT
 - Halosperm





Anti-oxidative substances

- Glutathione and vitamin E
- Free or total carnitine, α -glycosidase
- Carnitine and acetyl-carnitine
- Selenium and vitamin E
- Zinc and folid acid

Bhardwaj A, et al. Asian J Androl 2000, 2:225

Zopfgen A, et al. Hum Reprod 2000, 15:840

Vicari E, et al. Hum Reprod 2001, 16:2338

Keskes-Ammar L, et al. Arch Androl 2003, 49:83

Wong WY, et al. Fertil Steril 2002, 77:491

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Testicular ultrasound



Testicular Triplex



Transrectal ultrasound



Diagnostic evaluation

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Testicular FNA and TESE



Normal spermatogenesis





Hypospermatogenesis





Spermatogenesis arrest



TESE

Sertoli cell-only syndrome







Prognosis of testicular extraction



Tuettelmann F, et al. Int J Androl 2010, 34:291

microTESE





Sperm cryopreservation



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Yq microdeletions



• Three main areas Yq

Vogt PH, et al. Hum Mol Genet 1996, 7:933

• Azoospermia / severe OAT: 3%

Osterlund C, et al. Int J Andr 2000, 23:225
Cystic fibrosis

- Congenital bilateral agenesis of vas deferens (CBAVD)
- Obstructive azoospermia
- Congenital bronchiectasis
- Chronic pancreatitis

Claustres M, et al. Hum Mutat 2000, 16:143

Jarvi K, et al. Lancet 1995, 345:1578

Girodon E, et al. Eur J Hum Genet 1997, 5:149

Sharer N, et al. N Eng J Med 1998, 339:645

Genetics of male infertility

Human Reproduction, Vol.25, No.6 pp. 1383-1397, 2010

Advanced Access publication on April 8, 2010 doi:10.1093/humrep/deq081

human reproduction **ORIGINAL ARTICLE Andrology**

Evaluation of 172 candidate polymorphisms for association with oligozoospermia or azoospermia in a large cohort of men of European descent

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Sperm FISH



Sperm disomy: 24,XY

Aims

- Evaluation and etiologic approach of male infertility
- Selection of the optimal management method

Male infertility - 1

Etiology

- Kallmann syndrome
- Prolactinoma
- Hyperthyroidism
- Hypothyroidism

Etiologic approach

- Gonadotropins
- Dopamine agonists
- Anti-thyroid drugs
- L-thyroxine

Male infertility - 2

Etiology

- Infections
- Varicocele
- Obstruction

Oriented approach

- Antibiotics
- Surgery
- Microsurgical approach

Male infertility - 3

Etiology

Idiopathic infertility

Empirical approach

- Citric clomiphene / tamoxifen
- Gonadotropins
- Testosterone
- Anti-oxidants / Vitamins
- Insemination (IUI)
- |CS|

Citric clomiphene



Testicular dysgenesis

Human Reproduction Vol.16, No.5 pp. 972–978, 2001 OPINION

Testicular dysgenesis syndrome: an increasingly common developmental disorder with environmental aspects

N.E.Skakkebæk¹, E.Rajpert-De Meyts and K.M.Main

Department of Growth and Reproduction, Copenhagen University Hospital, Copenhagen, Denmark

Testicular dysgenesis



Skakkebaek NE et al. Hum Reprod (2001) 16:972

Testicular dysgenesis



Skakkebaek NE et al. Hum Reprod (2001) 16:972

Male infertility

- Application of modern diagnostic methods for the etiologic diagnosis of the cases
- Appropriate evaluation of the patients and their classification into subgroups to select the cases for the application of the optimal therapeutical approaches

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- C. Tsametis (endocrinologist)
- P. Poulakos (endocrinologist)
- P. Iliadou (endocrinologist)
- C. Dimopoulou (endocrinologist)
- E. Kintiraki (endocrinologist)
- S. Paschou (endocrinologist)
- P. Anagnostis (endocrinologist)
- I. Litsas (endocrinologist)
- G. Kanakis (endocrinologist)
- G. Mintziori (endocrinologist)
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